

Wip1 phosphatase: between p53 and MAPK kinases pathways

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ABSTRACT

Cells undergoing oncogenic transformation frequently inactivate tumor suppressor pathways that could prevent their uncontrolled growth. Among those pathways p53 and p38MAPK pathways play a critical role in regulation of cell cycle, senescence and cell death in response to activation of oncogenes, stress and DNA damage. Consequently, these two pathways are important in determining the sensitivity of tumor cells to anti-cancer treatment. Wild type p53-induced phosphatase, Wip1, is involved in governance of both pathways. Recently, strategies directed to manipulation with Wip1 activity proposed to advance current day anticancer treatment and novel chemical compounds synthesized to improve specificity of manipulation with Wip1 activity. Here we reviewed the history of Wip1 studies in vitro and in vivo, in genetically modified animal models that support Wip1 role in tumorigenesis through regulation of p53 and p38MAPK pathways. Based on our knowledge we propose several recommendations for future more accurate studies of Wip1 interactions with other pathways involved in tumorigenesis using recently developed tools and for adoption of Wip1 manipulation strategies in anti-cancer therapy.

The PP2C delta phosphatase Wip1 plays an important role in normal homeostasis and pathogenesis of several human diseases [1]. These features are mainly connected with Wip1 ability to regulate signaling in MAPK kinases pathway p53 network and DNA damage response (DDR). Recently published data on the genetic polymorphism in Wip1 gene PPM1D [2] and the presence of genetic amplification and mutations of this gene in cancer [3] additionally confirmed Wip1 importance for tumorigenesis, making even more attractive an idea of Wip1 as potential oncotarget for developing new protocols in anti-cancer therapy [4]. In presented review, we will try to sort out facts from more than fifteen-year old history of Wip1 research pointing to possible benefits and side effects of manipulation with Wip1 levels and activity.

Wip1 was first found as p53 target gene and named, Wip1 (wild-type p53-induced phosphatase), in the laboratory of Ettore Appella [5]. Interestingly, that in this first report authors presented evidences that Wip1 overexpression negatively regulates clonal survival in two tumor cell lines, T98G and Saos2. However, further studies by several groups identified targets of Wip1-dependent dephosphorylation and proved that Wip1 revealed features

of mild oncogene rather than tumor suppressor. The initial controversy as we currently understand was due to the p53-negative status of both cell lines, T98G and Saos2, used in the original study. The p53 status, with active or compromised p53 pathway, is crucial for prediction of effects from manipulation with Wip1 activity or levels.

Below, we compare effects of increasing Wip1 activity versus Wip1 inhibition focusing on data obtained mainly in experiments with genetically modified mice.

WIP1 DELETION AND INHIBITION

Wip1, DDR and p53 pathway

Currently, Wip1 is considered as a bona fide negative regulator of p53 pathway. p53 itself is the most potent transcriptional activator regulating transcription of Wip1 gene, PPM1D, compared to other identified Wip1 regulating transcriptional factors, such as NFκB [6], CREB [7], ER [8], c-Jun [9] and others. The cascade of events after DNA damage leads to p53 activation,

subsequent binding Wip1 promoter and turning ON Wip1 transcription that resulted in elevated levels of Wip1 protein. Wip1 directly dephosphorylates p53 on Ser 15 [10], hence reducing, p53 activity and levels, but more importantly in our opinion Wip1 down regulates the whole system of positive signaling to p53 from damaged DNA. ATM [11], Chk1 [10], Chk2 [12], γ H2AX [13, 14, 15, 16] proteins, which are responsible for sensing DNA damage and activating DDR, were shown to be targeted and negatively regulated by Wip1 (Figure 1). It was also shown by the group of L. Donehower that Wip1 could potentiate MDM2 function to further reduce p53 levels [17].

This feedback mechanism of p53 - Wip1 relations is thought to be necessary for cell recovery after DNA damage and some forms of stress. The fine-tuning of p53 response is also vital for the prevention of cell death or senescence especially during development and in stem cell homeostasis.

p53 and members of DDR system play a significant role in tumor suppression and many anti-cancer strategies are based on their activation. In agreement with this fact, more active p53 pathway in mice with genetic deletion of Wip1 significantly attenuated tumorigenesis in several tumor models, APC (Min) intestinal tumorigenesis [18], c-myc induced lymphoma [10] and others. In these models the mediating role of p53 was confirmed by reversing tumor suppressive phenotype of Wip1^{-/-} mice by deletion of p53. At the same time, Wip1 deletion was unable to prevent lymphomas and sarcomas associated with p53^{-/-} phenotype. The negative consequences of Wip1 deletion such as premature aging [19], lymphopenia [20] also could be linked to the abnormal regulation of p53 pathway and in many cases cured by p53 inhibition.

These facts lead to the conclusion that Wip1 inhibition strategy could sensitize p53 to upstream signaling, including DDR and oncogenic stress, and thus prevent tumorigenesis. However, one has to take into account possible side effects in normal tissues with elevated activity of DDR, high proliferation pace and therefore highly sensitive to DNA damage.

Interestingly, Wip1 deletion protected mice from mammary tumorigenesis [21]. From three models of breast cancer studied by Bulavin and others Wip1 deletion attenuated mammary gland tumorigenesis only in MMTV-ErbB2 and MMTV-HRAS1 mice, but not in MMTV-WNT1 mice [22]. The authors did not study deeply effects of p53 deletion in mammary gland on the Wip1-dependent tumor suppression, but linked the observed effect to another direct target of Wip1 - p38 MAPK.

WIP1, P38 AND OTHERS

Takekawa et al 2000 found that Wip1 directly binds and inactivates p38 MAPK by dephosphorylation of Thr182 activatory site under stress conditions such as

H₂O₂ treatment or UV-irradiation [23]. The activation of p38 can parallel with the simultaneous activation of p53 and plays direct role in positive posttranslational regulation of p53. Despite possible crosstalk between MAPK pathway and p53 pathway we foresee that it will be important to distinguish p53-dependent and p38-dependent effects of Wip1 deletion and/or activation. In the study mentioned above (Bulavin and colleagues [22]) the absence of inhibitory effect of Wip1 on p38 leads to elevated levels of two products of Cdkn2a gene, p16 and p14^{arf}, which were responsible for tumor suppression in mammary gland epithelium. Pietersen's group followed up study on mammary gland development in Wip1^{-/-} mice deepened our knowledge showing that Wip1 effect is restricted to the estrogen receptor positive cells of luminal layer of mouse mammary gland epithelium. This finding correlates with data from patients with breast cancer. It has been shown that the amplification of and mutations in the PPM1D gene occurred mainly in the luminal subtype of human breast cancer [24]. This group also showed that Wip1 could regulate activity of Stat5 transcriptional factor and another member of MAPK family - ERK kinase.

In addition to intrinsic interplay between Wip1, p53, p38 and other factors in tumors cells, tumorigenesis could be affected by changes in immune system of Wip1^{-/-} mice (Figure 2).

Though defect in T-cell and B-cell maturation was attributed largely to the function of p53 with no considered as p53-dependent effect with no involvement of p38 [19, 23], the increase in granulocyte compartment of hematopoietic system was linked to p38 proficiency in Wip1^{-/-} mice [25]. This disturbance in immune system, regulation of stromal cells [26], involvement of p38 pathway in cytokine secretion and described proinflammatory phenotype of Wip1^{-/-} mice [27, 28] allow us to hypothesize that tumorigenesis could be affected not only at the level of tumor cells themselves, but by tumor microenvironment as well. This hypothesis requires new approaches and tools such as conditional deletion of Wip1 in specific cells or tissues. Single reports on Wip1 involvement in regulation of other MAPK kinases: ERK [29] and JNK [8, 30], NF κ B [31, 32] pathway additionally motivate community to conduct such experiments.

Unfortunately, at the present moment the data on genetic deletion of Wip1 in mice can not be fully supported by studies which used Wip1 inhibitors instead of genetic deletion or siRNA/shRNA depletion shRNA depletion due to reported off-target effects of current day Wip1 inhibitors [33]. The careful adjustment of inhibitor concentration could be performed in experiments with cell lines in culture, but more complex pharmacological aspects of in vivo experiments makes interpretation of data obtained in such studies with animals a difficult task. Recently, two new promising Wip1 inhibitors reported to have a high specificity towards Wip1 inhibition. They can

potentiate cytotoxic effects of chemotherapeutic agents in wild type p53 tumor cells, though their potential side effects and usefulness for anti-cancer therapy await further studies in vivo [34, 35, 36].

As an additional remark to the data obtained from knockout animals, we would like to mention, that in all experiments we conducted on mice with genetically modified Wip1 no haploinsufficiency was observed. It seems that expression of Wip1 only from one allele is sufficient for proper function and does not disturb phenotype at least in normal development and tumorigenesis. On the other hand, it is important to mention that Wip1 deletion (Wip1^{-/-}) in heterozygous p53^{+/-} animals applies selective pressure on p53 pathway during tumorigenesis and frequently leads to the complete loss of p53 in tumors [11].

WIP1 OVEREXPRESSION

Recently, significant number of studies published on Wip1 overexpression in various tumors that correlates or not with status of positive p53 status in these tumors. Some studies emphasize on very important fact that Wip1 overexpression predominantly observed in tumors without p53 mutations highlighting a fact that Wip1 overexpression gives no advantages to the tumors without functional p53

tumor suppressor pathway. The critical assessment of such studies indicated that presented data especially on protein Wip1 levels could be sometime confusing and contaminating informational landscape. Different poorly described and verified anti-Wip1 antibodies from different companies were used in these studies frequently showing figures with cytosolic localization of mainly nuclear Wip1 protein. The new verified anti-Wip1 antibodies recently announced by the companies with long-time expertise in antibodies production will allow verification and refining of previously published data.

Currently, two mouse models with Wip1 overexpression were reported: ubiquitous Wip1 transgenic mouse, Ub-Wip1 [37], and mammary gland specific Wip1 transgenic mouse, MMTV-Wip1 [38].

Despite the fact that Wip1 overexpression was reported in many human cancers, and correlated with the more aggressive stage and poor prognosis, no spontaneous tumor appearance was observed in mice with Wip1 overexpression. One of the explanations to this phenomenon can be that the life span of mice is not sufficient to accumulate secondary to Wip1 mutations required for tumor formation. Thus, it can be assumed that overexpression of Wip1 has potentiating effect but not sufficient to trigger tumorigenesis. In other words, it only sets up the stage for cancer-driving mutations.

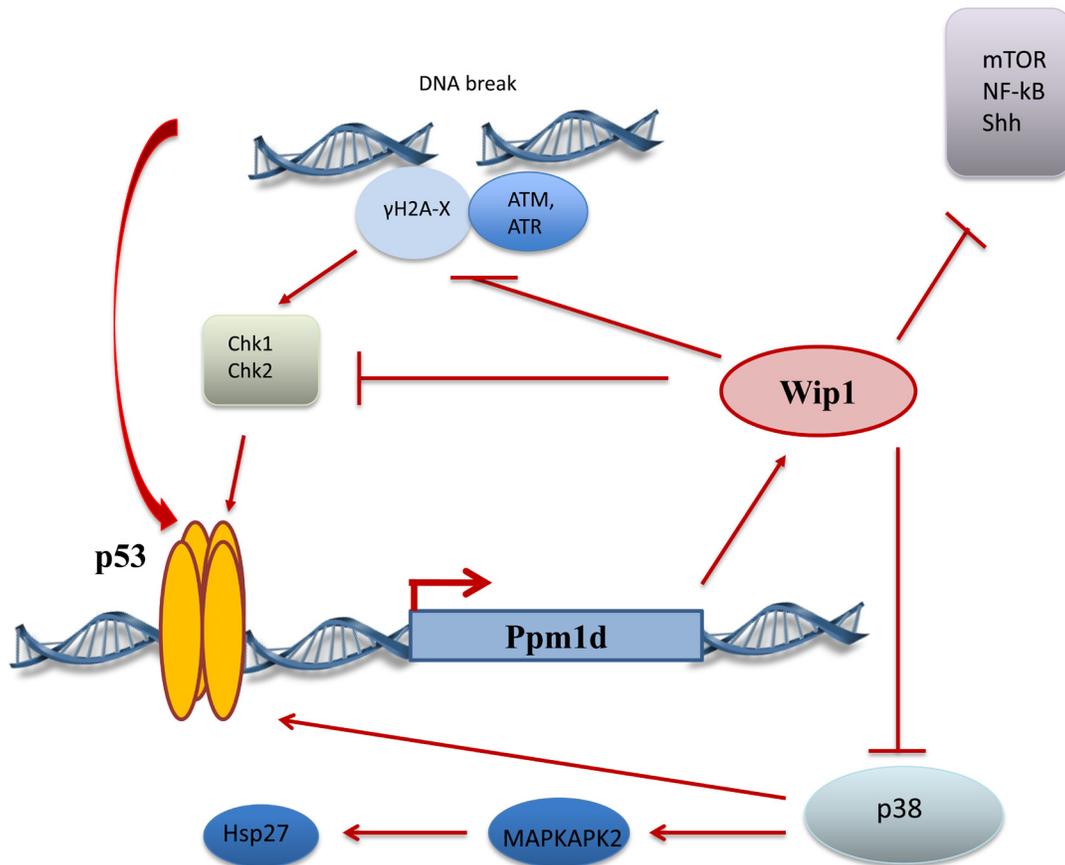


Figure 1: Wip1 regulation and targets.

In the breast cancer animal model Wip1 overexpression significantly accelerated mammary gland tumor formation MMTV- Erbb2/MMTV-Wip1 mice. The oncogenic effect was not dependent on p53 status. No differences was observed in MMTV- Erbb2/MMTV-Wip1/p53^{-/-} mice. On the contrary, introduction of an active copy of the MKK6 (MMTV-MKK6*) into the mammary gland epithelium minimized the pro-oncogenic effect of Wip1 overexpression [39].

Interestingly, that the initial observation by Fiscella et al 1998 that Wip1 overexpression could have some tumorsuppressive effect was confirmed later [41], but it is true only for tumors with negative p53 status.

For some cell lines, e.g. MCF7, with with amplification of the PPM1D chromosomal locus or some patient derived tumor cell explants with overexpressed Wip1, the uncontrolled cell cycling and extended resistance to apoptosis was supported by the fact of negative influence of elevated Wip1 levels on activity of p53 pathway [40].

The opposite effect was observed in tumor cell lines with p53 negative status.

p53 is one of the most important tumor suppressors regulating initial steps of oncogenic transformation, tumor growth and response to anti-cancer treatments. It is also one of the most frequently mutated genes in cancer. In some cancers e.g. large intestine, p53 mutations occur in majority of the cases according to Wellcome Trust Sanger Institute catalogue of somatic mutations in

cancer. The consequences of different types of mutations in p53 gene can vary. They could either mildly affect p53 functions or culminate in lost-of- function and/or gain of alternative functions. Chromosomal deletion or mutations, compromising p53 DNA binding, abrogate p53 transcriptional functions. Hence, the initiation of apoptotic program in response to DNA damage inducing agents can be abolished due to insufficient increase in expression of pro-apoptotic genes and lack of suppression of anti-apoptotic proteins. The unchanged in response to therapeutic stress pro-/ anti-apoptotic ratio can be responsible for drug resistance in tumors. For example, in our model osteosarcoma cell line Saos2 mutation in p53 gene allows cells to tolerate well high doses of chemotherapeutic drugs e.g. cisplatin, that are usually toxic for cells with preserved p53 functions. The absence of p53 nullify positive anti-tumor effect of Wip1 depletion/inhibitor observed in cells with p53 functions are still intact [39].

On the contrary, after disappearance of p53 as the main direct and indirect substrate for Wip1, new functions of Wip1 and its new substrates can be more easily revealed and become more important for cellular physiology. We found that inducible overexpression of Wip1 in Saos2 compensated tumor cells response to chemotherapy for the loss of p53. Tumor cells became more sensitive to cisplatin and underwent caspase-3 dependent apoptosis due to restored shift of Bax/Bcl-XL ratio towards pro-apoptotic protein. In the absence of p53 Wip1 regulated

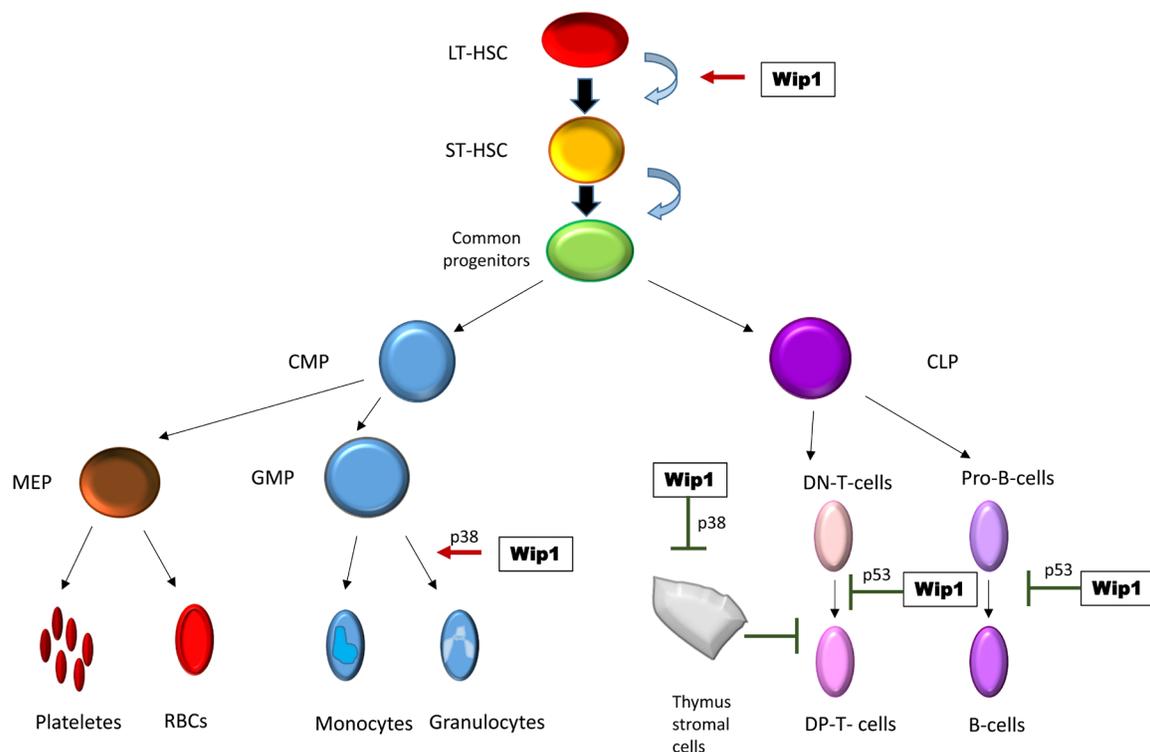


Figure 2: Wip1 in regulation of hematopoiesis.

transcriptional factor Runx2 binding to specific RUNT elements in Bax promoter and activation of Bax transcription by dephosphorylation of critical inhibitory sites in Runx2 molecule [41]. Furthermore, Wip1 suppressed anti-apoptotic BCL-XL expression by interfering with NFkB pathway [42].

The peculiar fact is that to increase efficiency of DNA-damaging drug, such as cisplatin, in p53 negative cells you do not need the increase in DDR. Normally, double strand breaks in DNA induce cascade of signaling events including phosphorylation of histone γ H2AX on Ser-139. This phosphorylation is a marker an early initiation point of DDR resulted in cell cycle arrest or cell death. The DNA damage induced γ H2AX phosphorylation was reduced in Wip1 overexpressing cells as Wip1 targets the p-Ser 139 site. At the same time these cells with lower phospho- γ H2AX levels were more responsive to cisplatin than control Saos2 cells with high phospho- γ H2AX levels. This observation accentuate the fact that phospho- γ H2AX levels as a marker of DDR, does not always correlate with toxicity of chemotherapeutic drug.

This phenomena is important so as in normal tissues hyper phosphorylation of γ H2AX could induced extensive signaling in p53 pathway resulted in cell death and damage to normal tissues. Wip1 is capable to decrease the toxic DNA damage signaling from chemotherapeutic agents to p53 in normal tissues, so as p53 functions preserved in all other organs and tissues except tumors with p53 mutations. Prevention of cell death in highly sensitive to DNA damage tissues with high rate of proliferation such as intestinal epithelium, testes, and hematopoietic cells could reduce side effects of anti-cancer therapy that together with increased sensitivity of p53-negative tumor cells will widen therapeutic window and allow selection of optimal dosage for anti-cancer drug.

Currently, Wip1 is established and well confirmed by significant body of publications as a regulator of two main pathways, DDR-p53 and MAPK-kinases. Recently announced involvement of Wip1 phosphatase in regulation of other pathways, such as Stats, NFkB [43], mTOR, Sonic Hedgehog [44] and Notch signaling requires additional clarification so as non-specific Wip1 inhibitor (CCT007093) was used in many studies and these observation could be purely correlative.

WIP1, MTOR AND AUTOPHAGY

Recent consideration of Wip1 as a potential oncogene allows to develop hypotheses about links between Wip1 activity and several processes, crucial for carcinogenesis, including autophagy. Autophagy (macroautophagy) is an evolutionarily conserved degradation process, during which proteins or damaged organelles (cargo) are isolated into double-membrane vesicles and digested as a result of fusion with lysosomes [45, 46]. Autophagy is an essential process that occurs

throughout all cell types, supporting normal homeostasis as well as helping to cope with stress conditions and carcinogenesis [47, 48]. Role of autophagy for cancer cells is dual, depending on stage of transformation and cell type. Autophagy is inhibited by multiple oncogene products, including PI3K, AKT, BCL-2 and mutant p53, and inhibition of autophagy is a potentially oncogenic event [49]. However whereas at the stage of solid tumor autophagy can serve as rescue pathway for cancer cells that face lack of oxygen and nutrients, thus inhibition of autophagy at this stage can lead to elimination of malignant cells. Being an important process for cancer cells, autophagy is studied as a potential target for cancer therapy.

A suggestion can be made that Wip1 phosphatase is somehow involved in autophagy regulation. One may suppose that the links are in DNA damage response regulation. Autophagic process seems to be in a tight interplay with DDR [50, 51], some DDR participants being shown to contribute to autophagy regulation: ATM [52], PARP-1 [53] p53 [54, 55]. Autophagy, in turn, was shown to contribute to maintaining genome stability [56, 57]. These authors have shown that loss of core autophagic protein Beclin 1 functions increase genome abnormalities. Autophagic protein FIP200 deficiency turned out to lead to lag in DNA repair [58]. However, despite the facts of significance of autophagy for genome stability, connecting Wip1 with autophagy through Wip1 interactions with DDR participants still needs experimental proves.

A more promising candidate for a link between Wip1 and autophagy is mTOR kinase. mTOR is known as a key regulator of cellular senescence [59, 60, 61, 62], stress response and cell death [63, 64, 65]. mTOR exists in two complexes - mTORC1 and mTORC2, mTORC1 being a negative regulator of autophagy [66, 67]. First, Wip1 was linked to autophagy in the study conducted by Le Guezennec et al [68]. They have shown involvement Wip1 in regulation of autophagy in mice during conversion of macrophages into foam cells in atherosclerosis. Authors suggested involvement of ATM and mTOR in this process. Previously, links between Wip1 and mTOR signaling were only to be assumed, built on the fact that Wip1 dephosphorylates several proteins that regulate mTOR activity directly or indirectly. These include p53, which affects mTOR activity and autophagy [69, 70]. p53 binds to the promoters of genes, the protein products of which regulate autophagy, including upstream regulators, such as the β 1, β 2 and γ -subunits of AMP-activated protein kinase (AMPK), several regulators of the AMPK and mTOR1 complexes, sestrin 1 and sestrin 2 proteins, connect genotoxic stress and mTOR signaling [71]. Recently catalogue of autophagy-relevant genes induced by p53 was published [72]. In addition to *Ulk1*, authors identified a set of autophagy genes, including *Atg2b*, *Atg4a*, *Atg4c*, *Atg7*, *Atg10*, *Tmem49/Vmp1*, *Ulk2*, and *Uvrag*, as p53-responsive. Furthermore, it was as previously found,

lysosomal protein also bound by p53 [73].

Then, another Wip1 target p38 was shown to participate in mTOR signaling activity regulation, for example, through interactions with mTORC1 component Raptor [74] or with Rag GTPases that regulate mTORC1 in response to amino acid modulation and stress [75, 76]. Also, p38 is involved into the regulation of autophagy. It was shown that p38 suppressed starvation-induced autophagy by regulating autophagic protein mAtg9 [77]. In the other hand, modulation of p38 activity was shown to affect LC3 conversion from LC3 I to LC3 II, which is a necessary event for the occurrence of autophagy [78, 79]. On the whole, it appears that influence of p38 on autophagy depends on cell type and conditions. However, the affect of p38 on autophagy through mTOR signaling is still to investigate.

Importantly, recently it was shown that Wip1 can directly dephosphorylate mTOR [80]. Authors have found that Wip1^{-/-} mice exhibit enhanced liver regeneration after partial hepatectomy, characterized by increased levels of phosphorylated mTOR (Ser2448, Ser2481, Ser2159), as well as it's phosphorylated targets p70S6K and S6. Enhanced liver regeneration and mTOR activity were linked with increased proliferation. Nevertheless, mTOR kinase is not only involved in proliferation, but also in cellular senescence, stress response, cell death, negative autophagy regulation. Inhibition or enhancement of these processes is an important event for carcinogenesis, these facts allow to suggest a new signaling network between Wip1 and mTOR, that can be considered as a potential target for modulation in anticancer therapy.

CONCLUSIONS AND PERSPECTIVES

1. Wip1 is a potential oncotarget and strategies directed to manipulation with its levels/activity could benefit modern day anti-cancer therapeutics. All current and future data on Wip1 polymorphism/mutations in tumors or manipulations with Wip1 activity/levels should take into account p53 status, so as p53 gene is one most frequently mutated genes in tumors and its status affects Wip1 expression and actions. All previously accumulated data should be re-analyzed carefully taking this note under consideration.

2. Wip1 levels/activity could be important diagnostic and prognostic tool in oncology. The clinical development of this tool required more specific anti-Wip1 antibodies, analysis of nuclear versus cytoplasm localization of Wip1, and taking into the account regulators of Wip1 expression: p53, DNA repair genes, MAPK, NFκB status of the tumor.

3. In tumors with preserved wild type p53 pathway Wip1 inhibition increases tumor suppressive effect of p53, but this strategy should take into account that Wip1 inhibition in normal tissues and their stem cell compartment significantly increases sensitivity to current

day anti-cancer therapy. In this type of tumor Wip1 is a better alternative to MDM2 inhibitors, because it does not affect p53 levels directly and does not immediately increase apoptosis in normal tissues per se, but rather mildly affects sensitivity of p53 for upstream signaling during DDR.

4. In more than half of all tumors p53 is mutated and Wip1 inhibition is ineffective. On contrary, temporary Wip1 activation could increase sensitivity to chemotherapeutic drugs and decrease side effects of DDR in normal tissues. The current day oncological practice could benefit from this strategy of Wip1 activation/p53 downregulation synthetic sensitivity to chemotherapeutic drugs.

5. Wip1 is a potent anti-aging, anti-inflammatory and neuromodulator agent, but these type of studies should take into account possible pro-tumorigenic effect of Wip1 activation.

6. The perspective of further studies is understanding of Wip1 interplay with multiple pathways on the cellular level (nuclear and cytoplasm) and whole organism level (secretion and immune system), finding new regulators and targets of Wip1, as potential therapeutic tool.

Taking into account information that was written above the most urgent task in the field is creation of specific anti-Wip1 assays as diagnostic tool and developing new bioavailable inhibitors/activators of Wip1 as promising therapeutics for tumor prevention and anti-cancer treatment that could significantly improve success rate in the treatment of oncological diseases.

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CONFLICTS OF INTEREST

The authors declare no conflict of interest.

REFERENCES

1. Zhu YH and Bulavin DV. Wip1-dependent signaling pathways in health and diseases. *Prog Mol Biol Transl Sci*, 2012; 106: p. 307-25.
2. Dudgeon C, Shreeram S, Tanoue K, Mazur SJ, Sayadi A, Robinson RC, Appella E, Bulavin DV. Genetic variants and mutations of PPM1D control the response to DNA damage. *Cell Cycle*, 2013; 12: 2656-2664.
3. Zaikowicz A, Butkiewicz D, Drosik A, Giglok M, Suwinski

- R, Rusin M. Truncating mutations of PPM1D are found in blood DNA samples of lung cancer patients. *Br J Cancer*, 2015; 112: 1114-20.
4. Le Guezennec X, Bulavin DV. WIP1 phosphatase at the crossroads of cancer and aging. *Trends Biochem Sci*, 2010; 109-114.
 5. Fiscella M, Zhang H, Fan HL, Sakaguchi K, Shen S, Mercer WE, Vande Woude GF, O'Connor PM, Appella E. Wip1, a novel human protein phosphatase that is induced in response to ionizing radiation in a p53-dependent manner. *Proc. Natl. Acad. Sci*, 1997; 94: 6048-6053.
 6. Lowe JM, Cha H, Yang Q, Fornace AJ Jr. Nuclear factor-kappaB (Nf-kappaB) is a novel positive transcriptional regulator of the oncogenic Wip1 phosphatase. *J Biol Chem*, 2010; 285: 5249-57.
 7. Rossi M, Demidov ON, Anderson CW, Appella E, Mazur SJ. Induction of Ppm1d following DNA-damaging treatments through a conserved p53 response element coincides with a shift in the use of transcription initiation sites. *Nucleic Acids Res*, 2008; 36: 7168-80.
 8. Han HS, Yu E, Song JY, Park JY, Jang SJ, Choi J. The estrogen receptor alpha pathway induces oncogenic Wip1 phosphatase expression. *Mol Cancer Res*, 2009; 7: 713-723.
 9. Song JY, Han HS, Sabapathy BM, Lee EYu, Chou J. Expression of a homeostatic regulator, Wip1 (Wild-type p53-induced phosphatase), is temporarily induced by c-Jun and p53 in response to UV irradiation. *J Biol Chem*, 2010; 19: 9067-9076.
 10. Lu X, Nannenga B, Donehower LA. Ppm1d dephosphorylates Chk1 and p53 and abrogates cell cycle checkpoints. *Genes Dev*, 2005; 19: 1162-74.
 11. Shreeram S, Hee WK, Demidov ON, Kek C, Yamaguchi H, Fornace AJ Jr, Anderson CW, Appella E, Bulavin DV. Regulation of ATM/p53-dependent suppression of myc-induced lymphomas by Wip1 phosphatase. *J Exp Med*, 2006. 203: p. 2793-9.
 12. Fujimoto H, Onishi N, Kato N, Takekawa M, Xu XZ, Kosugi A, Kondo T, Imamura M, Oishi I, Yoda A, Minami Y. Regulation of the antioncogenic Chk2 kinase by the oncogenic Wip1 phosphatase. *Cell Death and Differentiation*, 2006; 13: 1170-1180.
 13. Cha H, Lowe JM, Li H, Lee JS, Belova GI, Bulavin DV, Fornace AJ Jr. Wip1 directly dephosphorylates gamma-H2AX and attenuates the DNA damage response. *Cancer Res*. 2010;70:4112-22.
 14. Moon SH, Lin L, Zhang X, Nguyen TA, Darlington Y, Waldman AS, Lu X, Donehower LA. Wild-type p53-induced phosphatase 1 dephosphorylates histone variant gamma-H2AX and suppresses DNA double strand break repair. *J Biol Chem*, 2010; 285: 12935-47.
 15. Oh KS, Bustin M, Mazur SJ, Appella E, Kraemer KH. UV-induced histone H2AX phosphorylation and DNA damage related proteins accumulate and persist in nucleotide excision repair-deficient XP-B cells. *DNA Repair (Amst)*, 2011; 10: 5-15.
 16. Macurek L, Lindqvist A, Voets O, Kool J, Vos HR, Medema RH. Wip1 phosphatase is associated with chromatin and dephosphorylates gammaH2AX to promote checkpoint inhibition. *Oncogene*, 2010; 29: 2281-2291.
 17. Lu X, Ma O, Nguyen TA, Jones SN, Oren M, Donehower LA. The Wip1 Phosphatase acts as a gatekeeper in the p53-Mdm2 autoregulatory loop. *Cancer cell*, 2007; 12: 342-54.
 18. Demidov ON, Timofeev O, Lwin HN, Kek C, Appella E, Bulavin DV. Wip1 phosphatase regulates p53-dependent apoptosis of stem cells and tumorigenesis in the mouse intestine. *Cell Stem Cell*, 2007; 1: 180-90.
 19. Sakai H, Fujigaku H, Mazur SJ, Appella E. Wild-type p53-induced phosphatase 1 (Wip1) forestalls cellular premature senescence at physiological oxygen levels by regulating DNA damage response signaling during DNA replication. *Cell Cycle*, 2014; 13: 1015-1029.
 20. Schito ML, Demidov ON, Saito S, Ashwell JD, Appella E. Wip1 phosphatase-deficient mice exhibit defective T cell maturation due to sustained p53 activation. *J Immunol*, 2006. 176: p. 4818-25.
 21. Harrison M, Degenhardt Y, Hoey T, Powers S. Wip1-deficient mice are resistant to common cancer genes. *Trends Mol Med*, 2004; 10: 359-361.
 22. Bulavin DV, Philips C, Nannenga B, Timofeev O, Donehower LA, Anderson, CW, Appella E, Fornace AJ Jr. Inactivation of the Wip1 phosphatase inhibits mammary tumorigenesis through p38 MAPK-mediated activation of the p16(Ink4a)-p19(Arf) pathway. *Nat Genet*, 2004. 36: p. 343-50.
 23. Takekawa M, Adachi M, Nakahata A, Nakayama I, Itoh F, Tsukuda H, Taya Y, Imai K. p53-inducible wip1 phosphatase mediates a negative feedback regulation of p38 MAPK-p53 signaling in response to UV radiation. *EMBO J*, 2000; 19: 6517-26.
 24. Emelyanov A, Bulavin DV. Wip1 phosphatase in breast cancer. *Oncogene*, 2015; 34: 4429-4438.
 25. Yi W, Hu X, Chen Z, Liu L, Tian Y, Chen H, Conq YS, Yang F, Zhang L, Rudolph, KL, Zhang Z, Zhao Y, Ju Z. Phosphatase Wip1 controls antigen-independent B-cell development in a p53-dependent manner. *Blood*, 2015; 126: 620-8.
 26. Liu G, Hu X, Sun B, Yang T, Shi J, Zhang L, Zhao Y. Phosphatase Wip1 negatively regulates neutrophil development through p38 MAPK-STAT1. *Blood*, 2013. 121: p. 519-29.
 27. Sun L, Li H, Luo H, Zhang L, Hu X, Yang T, Sun C, Chen H, Zhang L, Zhao Y. Phosphatase Wip1 is essential for the maturation and homeostasis of medullary thymic epithelial cells in mice. *J Immunol*, 2013. 191: p. 3210-20.
 28. Sun B, Hu X, Liu G, Ma B, Xu Y, Yang T, Shi J, Yang F, Li H, Zhang L, Zhao Y Phosphatase Wip1 negatively regulates neutrophil migration and inflammation. *J Immunol*, 2014. 192: p. 1184-95.

29. Choi J, Nannenga B, Demidov ON, Bulavin DV, Cooney A, Bravton C, Zhang Y, Mbawuiké IN, Bradley A, Appella E, Donehower LA Mice deficient for the wild-type p53-induced phosphatase gene (Wip1) exhibit defects in reproductive organs, immune function, and cell cycle control. *Mol Cell Biol*, 2002; 22: p. 1094-105.
30. Tarulli GA, De Silva D, Ho V, Kunasegaran K, Ghosh K, Tan BC, Bulavin DV, Pietersen AM. Hormone-sensing cells require Wip1 for paracrine stimulation in normal and premalignant mammary epithelium. *Breast Cancer Res*. 2013 Jan 31;15(1):R10. doi: 10.1186/bcr3381. PubMed PMID: 23369183; PubMed Central PMCID: PMC3672744.
31. Demidov ON, Zhu Y, Kek C, Goloudina AR, Motoyama N, Bulavin DV. Role of Gadd45a in Wip1-dependent regulation of intestinal tumorigenesis. *Cell Death Differ*, 2012; 19: 1761-8.
32. Chew J, Biswas S, Shreeram S, Humaidi M, Wong ET, Dhillon MK, Teo H, Hazra A, Fang CC, López-Collazo E, Bulavin DV, Tergaonkar V. WIP1 phosphatase is a negative regulator of NF-kappaB signalling. *Nat Cell Biol*, 2009; 11: 659-666.
33. Goloudina AR, Mazur SJ, Appella E, Garrido C, Demidov ON. Wip1 sensitizes p53-negative tumors to apoptosis by regulating the Bax/Bcl-xL ratio. *Cell Cycle*, 2012; 11: 1883-7.
34. Lee JS, Park JR, Kwon OS, Kim H, Fornace AJ Jr., Cha HJ. Off-target response of a Wip1 chemical inhibitor in skin keratinocytes. *J Dermatol Sci*, 2014; 73: 125-34.
35. Ogasawara S, Kiyota Y, Chuman Y, Kowata A, Yoshimura F, Tanino K, Kamada R, Sakaguchi K. Novel inhibitors targeting PPM1D phosphatase potently suppress cancer cell proliferation. *Bioorg Med Chem*. 2015 Oct 1;23:6246-9.
36. Richter M, Dayaram T, Gilmartin AG, Ganji G, Pemmasani SK, Van Der Key H, Shohet JM, Donehower LA, Kumar R. WIP1 phosphatase as a potential therapeutic target in neuroblastoma. *PLoS One*, 2015; 10: e0115635.
37. Gilmartin AG, Fajtá TH, Richter M, Groy A, Seefeld MA, Darcy MG, Peng X, Federowicz K, Yang J, Zhang SY, Minthorn E, Jaworski JP, Schaber M, et al. Allosteric Wip1 phosphatase inhibition through flap-subdomain interaction. *Nat Chem Biol*, 2014;10:181-7.
38. Wong ES, Le Guezennec X, Demidov ON, Marshall NT, Wang ST, Krishnamurthy J, Sharpless NE, Dunn NR, Bulavin D.V. p38MAPK controls expression of multiple cell cycle inhibitors and islet proliferation with advancing age. *Dev Cell*, 2009.; 17: 142-9.
39. Demidov ON, Kek C, Shreeram S, Timofeev O, Fornace AJ Jr, Appella E, Bulavin DV The role of the MKK6/p38 MAPK pathway in Wip1-dependent regulation of ErbB2-driven mammary gland tumorigenesis. *Oncogene*, 2007; 26: 2502-6.
40. Yu E, Anh YS, Jang SJ, Kim MJ, Yoon HS, Gong G, Choi J. Overexpression of the Wip1 gene abrogates the p38 MAPK/p53/Wip1 pathway and silences p16 expression in human breast cancers. *Breast Cancer Res Treat*, 2007; 101: 269-278.
41. Goloudina AR, Tanoue K, Hammann A, Fourmaux E, Le Guezennec X, Bulavin DV, Mazur SJ, Appella E, Garrido C, Demidov ON. Wip1 promotes RUNX2-dependent apoptosis in p53-negative tumors and protects normal tissues during treatment with anticancer agents. *Proc Natl Acad Sci U S A*. 2012; 109: 68-75.
42. Goloudin, AR, Mazur SJ, Appella E, Garrido C, Demidov ON. Wip1 sensitizes p53-negative tumors to apoptosis by regulating the Bax/Bcl-xL ratio. *Cell Cycle*. 2012;11:1883-1887
43. Salminen A, Kaarniranta K. Control of p53 and NF-kB signaling by WIP1 and MIF: role in cellular senescence and organismal aging. *Cell Signal*, 2011; 23: 747-752.
44. Doucette TA, Yang Y, Pedone C, Kim JY, Dubuc A, Northcott PD, Taylor MD, Fuhs DV, Rao G. WIP1 enhances tumor formation in a sonic hedgehog-dependent model of medulloblastoma. *Neurosurgery*, 2012; 70: 1003-1010.
45. Klionsky DJ. Autophagy. *Curr Biol* 2005; 15: 282-283.
46. Klionsky DJ., Emr S.D. Autophagy as a regulated pathway of cellular degradation. *Science*, 2000; 290: 1717-1721.
47. Kroemer G, Mariño G, Levine B. Autophagy and the integrated stress response. *Mol Cell*, 2010; 40: 280-293.
48. Rubinsztein DC, Codogno P, Levine B. Autophagy modulation as a potential therapeutic target for diverse diseases. *Nat Rev Drug Discov*. 2012; 11: 709-730.
49. Eisenberg-Lerner A, Kimchi A. The paradox of autophagy and its implication in cancer etiology and therapy. *Apoptosis*, 2009; 14: 376-391.
50. Czarny P, Pawlowska E, Bialkowska-Warzecha J, Kaarniranta K, Blasiak J. Autophagy in DNA damage response. *Int. J. Mol. Sci.*, 2015; 16: 2641-2662.
51. Filomeni G, De Zio D, Cecconi F. Oxidative stress and autophagy: the clash between damage and metabolic needs. *Cell Death Differ*, 2015; 22: 377-388.
52. Alexander A; Kim J; Walker CL. ATM engages the TSC2/mTORC1 signaling node to regulate autophagy. *Autophagy* 2010, 6, 672-673.
53. Munoz-Gamez JA, Rodríguez-Vargas JM, Quiles-Pérez R, Aguilar-Quesada R, Martín-Oliva D, de Murcia G, Menissier de Murcia J, Almendros A, Ruiz de Almodóvar M, Oliver FJ. PARP-1 is involved in autophagy induced by DNA damage. *Autophagy* 2009; 5: 61-74.
54. Maiuri MC, Galluzzi L, Morselli E, Kepp O, Malik SA, Kroemer G. Autophagy regulation by p53. *Curr Opin Cell Biol*, 2010; 22: 181-185.
55. Pietrocola F, Izzo V, Niso-Santano M, Vacchelli E, Galluzzi L, Maiuri MC, Kroemer G. Regulation of autophagy by stress-responsive transcription factors. 23: 310-322.
56. Mathew R, Kongara S, Beaudoin B, Karp CM, Bray K, Degenhardt K, Chen G, Jin S, White E. Autophagy suppresses tumor progression by limiting chromosomal

- instability. *Genes Dev.* 2007, 21, 1367-1381.
57. Vessoni AT, Filippi-Chiela EC, Menck GFM, Lenz G. Autophagy and genomic integrity. *Cell Death Differ*, 2013; 20: 1444-1454.
 58. Bae H, Guan J. Suppression of autophagy by FIP200 deletions impairs DNA damage repair and increases cell death upon treatments with anti-cancer agents. *Mol Cancer Res*, 2011; 9: 1232-1241.
 59. Demidenko ZN, Zubova SG, Bukreeva EL, Pospelov VA, Pospelova TV, Blagosklonny MV. Rapamycin decelerates cellular senescence. *Cell Cycle*, 2009; 8: 1886-1895.
 60. Pospelova TV, Leontieva OV, Bykova TV, Zubova SG, Pospelov VA, Blagosklonny MV. Suppression of replicative senescence by rapamycin in rodent embryonic cells. *Cell Cycle*, 2012; 11: 2402-2407.
 61. Korotchikina LG, Leontieva OV, Bukreeva EI, Demidenko ZN, Gudkov AV, Blagosklonny MV. The choice between p53-induced senescence and quiescence is determined in part by the mTOR pathway. *Aging (Albany NY)*, 2010; 2: 344-352. doi: 10.18632/aging.100160.
 62. Sarbassov DD, Ali SM, Sabatini DM. Growing roles for the mTOR pathway. *Curr Opin Cell Biol.* 2005; 17: 596-603.
 63. Blagosklonny MV. Hypoxia, MTOR and autophagy: converging on senescence or quiescence. *Autophagy*, 2013; 9: 260-262.
 64. Reiling JH, Sabatini DM. Stress and mTOR signaling. *Oncogene*, 2006; 25: 6373-6383.
 65. Castedo M, Ferri KF, Kroemer G. Mammalian Target of Rapamycin (mTOR): Pro- and Anti-Apoptotic. *Cell Death Differ*, 2002; 9: 99-100.
 66. Codogo P, Meijer AJ. Regulation and role of autophagy in mammalian cells. *Int J Biochem Cell Biol.* 2004; 36: 2445-2462.
 67. Jung CH, Ro S, Cao J, Otto NM, Kim D. mTOR regulation of autophagy. *FEBS Lett*, 2010; 584: 1287-1295.
 68. Le Guezennec X, Brichtkina A, Huang Y, Kostromina E, Han W, Bulavin DV. Wip1-dependent regulation of autophagy, obesity and atherosclerosis. *Cell Metabolism*, 2012; 16: 68-80.
 69. Feng Z. p53 regulation of the IGF-1/AKT/mTOR pathways and the endosomal compartment. *Cold Spring Harb Perspect Biol*, 2010; 2: a001057.
 70. Tang J, Di J, Cao H, Bai J, Zheng J. p53-mediated autophagic regulation: A prospective strategy for cancer therapy. *Cancer Lett*, 2015; 363: 101-107.
 71. Budanov AV, Karin M. p53 target genes *sestrin1* and *sestrin2* connect genotoxic stress and mTOR signaling. *Cell*, 2008; 134: 451-460.
 72. Kenzelman Broz D, Spano Mello S, Bieging KT, Jiang D, Dusek RL, Brady CA, Sidow A, Attardi LD. Global genomic profiling reveals an extensive p53-regulated autophagy program contributing to key p53 responses. *Genes Dev*, 2013; 27: 1016-1031.
 73. Azzolli CG, Sagar M, Wu A, Lowry D, Hennings H, Morgan DL, Weinberg WC. Cooperation of p53 loss of function and v-Ha-Ras in transformation of mouse keratinocyte cell lines. *Mol Carcinol*, 1998; 21: 50-61.
 74. Wu XN, Wang XK, Wu SQ, Lu J, Zheng M, Wang YH, Zhou H, Zhang H, Han J. Phosphorylation of Raptor by p38beta participates in arsenite-induced mammalian Target of Rapamycin Complex 1 (mTORC1) activation. *J Biol Chem*, 2011; 286: 31501-31511.
 75. Efeyan A, Zoncu R, Sabatini DM. Amino acids and mTOR: from lysosomes to disease. *Trends Mol Med*, 2012; 18: 524-533.
 76. Cully M, Genevet A, Warne P, Treins C, Liu T, Bastien J, Baum B, Tapon N, Leever SJ, Downward J. A Role for p38 Stress-Activated Protein Kinase in Regulation of Cell Growth via TORC1. *Mol Cell Biol*, 2010; 30: 481-495.
 77. Webber JL, Sharon AT. Coordinated regulation of autophagy by p38 MAPK through mAtg9 and p38IP. *The EMBO Journal*, 2010; 29: 27-40.
 78. Li J, Yang Y, Liu Q, Pan S, He Z, Zhang X, Yang T, Chen X, Wang D, Qiu J, Zhou S. The investigational Aurora kinase A inhibitor alisertib (MLN8237) induces cell cycle G2/M arrest, apoptosis, and autophagy via p38 MAPK and Akt/mTOR signaling pathways in human breast cancer cells. *Drug Des Devel Ther* 2015; 9: 1627-1652.
 79. Zhao R, Chen M, Jiang Z, Zhao F, Xi B, Zhang X, Fu H, Zhou K. Platycodin-D induced autophagy in non-small lung cancer cells via PI3K/Akt/mTOR and MAPK signaling pathways. *J Cancer*, 2015. 6: 623-631.
 80. Zhang L, Liu L, He Z, Li G, Song S, Jin H, Rudolph KL, Yang H, Mao Y, Zhang L, Xiao Z, Ju Z. Inhibition of wild-type p53-induced phosphatase 1 promotes liver regeneration in mice by direct activation of mammalian Target of Rapamycin. *Hepatology*, 2015; 61: 2030-2041.