

Implications of immune dysfunction on endometriosis associated infertility

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ABSTRACT

Endometriosis is a complex, inflammatory disease that affects 6-10% of reproductive-aged women. Almost half of the women with endometriosis experience infertility. Despite the excessive prevalence, the pathogenesis of endometriosis and its associated infertility is unknown and a cure is not available. While many theories have been suggested to link endometriosis and infertility, a consensus among investigators has not emerged. In this extensive review of the literature as well as research from our laboratory, we provide potential insights into the role of immune dysfunction in endometriosis associated infertility. We discuss the implication of the peritoneal inflammatory microenvironment on various factors that contribute to infertility such as hormonal imbalance, oxidative stress and how these could further lead to poor oocyte, sperm and embryo quality, impaired receptivity of the endometrium and implantation failure.

INTRODUCTION

Endometriosis is a chronic, inflammatory, estrogen-dependent disease that is characterized by the growth of endometrial tissue outside of the uterine cavity. Endometriosis affects 6-10% of reproductive-aged women. Despite its staggering economic impact (\$1.8 Billion/year in Canada [1]; \$22 Billion/year in the USA [2]), endometriosis remains misdiagnosed, misunderstood and ineffectively treated. The cause of this enigmatic disease is unknown; however, the theory of retrograde menstruation is widely accepted. This theory suggests that endometrial tissue, sloughed off during menstruation, is refluxed into the fallopian tubes and peritoneal cavity during menstrual uterine contractions. However, 76-90% of women have been shown to experience retrograde menstruation [3]. This has prompted researchers to question why only 6-10% of women develop endometriosis if so many women experience retrograde menstruation. To date, it has been suggested that the women who develop endometriosis have genetic, biochemical, or immunological dysfunction that prevents the removal of the tissue from the peritoneal cavity and rather facilitates tissue adhesion to peritoneal structures [4].

It has been well established that the immune system of women with endometriosis is dysfunctional. A multitude

of immune cell types, including neutrophils, macrophages, dendritic cells, natural killer cells, T helper cells and B cells have been shown to be dysregulated in women with endometriosis [5-10]. Additionally, cytokines and chemokines involved in inflammation, angiogenesis and tissue growth are increased in the plasma and peritoneal fluid (PF) of women with endometriosis [11, 12]. This local and systemic inflammatory *milieu* is suspected to stimulate symptoms commonly presented including pain and infertility [13, 14]. It has been reported that 35-50% of endometriosis patients experience infertility and 25-50% of infertile women have endometriosis [15]. The monthly fecundity rate in healthy couples, which is a couple's probability of conceiving in one month, is 15-20% [16]. In contrast, women with endometriosis have a monthly fecundity rate of 2-10% [17]. This suggests that women with endometriosis have a significantly lower likelihood of conceiving each month. While the use of medical therapeutics including contraceptive steroids, progestins, aromatase inhibitors, agonists of gonadotropin-releasing hormone and non-steroidal anti-inflammatory agents are helpful to relieve pain, the use or ceasing use of these agents rarely improves fertility [18-20]. Additionally, the side effects of long term use can be detrimental; for example, the use of oral contraceptives has been shown to affect endometrial thickness and growth [21]. Due to

improved fecundity observed during randomized control trial studies [22, 23], surgical removal of an endometriotic lesion and assisted reproductive technologies are used to treat endometriosis associated infertility. However, why surgical excision of endometriotic lesions improves fertility is unknown. It has been suggested that surgical removal of endometriotic lesions decreases peritoneal inflammation and results in improved fertility. Previous reports from our group have demonstrated that plasma and PF concentrations of inflammatory cytokines such as GM-CSF, IL-2, IL-8, IL-10 and IL-17 significantly decreased following lesion excision, suggesting inflammation is prompted by the lesion [24, 25]. These findings indicate the probable impact of endometriosis associated inflammation towards infertility.

Not only is the pathogenesis of endometriosis complex and unknown, the pathogenesis of infertility associated with endometriosis remains elusive. Furthermore, why some women with endometriosis display infertility and others do not is unknown. Infertility is not uniformly associated with disease state or size of the lesion, which prompts researchers to seek other conclusions to explain how infertility manifests in these patients. Immune system dysfunction is speculated to play a large role in endometriosis associated infertility; however, its precise role is rarely discussed in depth. In this review, we compile and analyze the current literature, including our own work, surrounding the role of the inflammation and immune system dysfunction in women with endometriosis associated infertility.

INFLAMMATION, IMMUNE DYSFUNCTION AND INFERTILITY

The pathogenesis of infertility associated with endometriosis is complicated by the involvement of biochemical, endocrine, immune and genetic factors. Whether immune dysfunction initiates the pathogenesis of endometriosis or is a product of the disease has not been identified. After decades of research, there appears to be a consensus that the immune system of women with endometriosis and women with endometriosis associated infertility is different from healthy, fertile controls. Both cellular and secreted immune mediators are aberrantly expressed in the PF and plasma of endometriosis patients. Pro-inflammatory cytokines such as TNF- α , IL-1 β , IL-6, IL-8, IL-10, IL-17, IL-33, IP-10, MCP-1 MIF and RANTES are aberrantly expressed in the PF of women with endometriosis [25-32]. Additionally, women with endometriosis associated infertility have an altered intraperitoneal immune cell status compared to women with unexplained infertility [33]. However, most of these published studies are observational and do not provide mechanistic pathways.

In the pursuit of understanding infertility in endometriosis, molecular profiling has been used

to identify differential gene expression in infertile, endometriotic patients. Using Nanostring immune transcriptomic profiling of 539 immune-inflammation related genes, our group found 91 genes that were aberrantly expressed in the eutopic endometrium of infertile, endometriosis patients. The differentially expressed genes were predominantly involved in cellular adhesion, cytokine-cytokine interaction, apoptosis and decidualization [34]. Specifically, pro-inflammatory cytokines, chemokines and receptors including *CXCL1*, *CX3CL1*, *CXCL9*, *CXCL10*, *IL-32*, *CXCR2*, *IL-7R* and adhesion molecules including *ICAM3* and *SELL* had a higher expression in the eutopic endometrium of infertile, endometriosis patients compared to fertile controls [34]. Furthermore, a decreased expression of *NOTCH1* and *NOTCH2* was observed, which aligns with previous studies associating disturbed NOTCH pathway signaling with impaired decidualization in the endometrium of women with endometriosis [34, 35]. In another study, a number of proteins important for inflammation and oxidative stress, including sphingosine kinase A, Hypoxia inducible factors, heat shock cognate proteins and superoxide dismutase, were found to be dysregulated in the eutopic endometrium from infertile women with ovarian, stage IV endometriosis [36]. The evidence shown here indicates inherent differences in the eutopic endometrium from infertile women with endometriosis. It is plausible that chronic, peritoneal inflammation is involved in shaping the transcriptome of eutopic endometrium in patients.

Cellular components of the immune system are dysregulated in endometriosis patients and specifically infertile endometriosis patients. Uterine natural killer (uNK) cells are a subset of NK cells residing in the uterus and play important roles in pregnancy and the development of the placenta [37]. Uterine NK cells have been linked to spiral artery remodeling and produce pro-inflammatory cytokines such as GM-CSF, CSF-1, TNF- α , IFN- γ , TGF- β , LIF and IL-2 and these cytokines have been associated with invasion of trophoblasts into the uterine wall [38, 39]. Aberrant numbers of uNK cells are linked to pregnancy pathologies including pre-eclampsia and fetal growth restriction [40]. Increased infiltration of CD16+ cytotoxic uNK cells, are found in the eutopic endometrium of women with endometriosis associated infertility [41]. Additionally, healthy, fertile controls have more mature uNK cells; however, immature uNK cell populations exist in infertile women with endometriosis [42]. Further studies are required to understand exactly why uNK cells are not maturing in infertile endometriosis patients and how this affects fertility as a whole.

T regulatory (Treg) cells are altered in endometriosis patients and have been suggested to play a role in the pathogenesis of endometriosis and its associated infertility [43]. Lower numbers of Treg cells have been detected in the eutopic endometrium of a non-human primate

endometriosis model [44]. Interestingly, in humans, the expression of the transcription factor forkhead P3 (Foxp3), a distinctive surface marker for Treg cells, is upregulated in the endometrium of women with endometriosis [45]. Additionally, higher expression of FoxP3 mRNA has been detected in the endometrium of infertile, advanced stage endometriosis women; that said, the Foxp3 protein was not significantly higher in these patients compared to fertile controls [46]. Furthermore, TGF- β has been shown to promote differentiation and induction of Treg cells [47]. TGF- β has been shown to be upregulated in the PF of women with endometriosis [48], which indicates that aberrant expression of Treg cells could be due to aberrant concentrations of TGF- β in the PF of these women. In contrast, women with unexplained infertility have shown reduced expression of Foxp 3 in the endometrium of compared to healthy fertile controls [49]. While the precise mechanism linking Treg cells and infertility requires further investigation, previous reports studying the pre-implantation period in murine pregnancy indicate that Treg cells play a critical role in implantation [50]. The presented evidence suggests that dysregulation of Treg cells may contribute to implantation failure observed in endometriosis patients.

There are other important immune cell types including macrophages and dendritic cells associated with inflammation in endometriosis. While it is likely that these cell types are involved in the endometriosis associated infertility, studies directly associating them with infertility in patients are scarce within the literature. Nevertheless, more studies are required to delineate coordinated interactions between various immune cells in the promotion and/or resolution of inflammatory cascade and its impact on infertility in endometriosis patients.

IMPLICATION OF IMMUNE DYSFUNCTION ON THE ENDOCRINE/IMMUNE PATHWAY

A specific and delicate balance between inflammatory factors and sex-hormones are required for the initiation and propagation of normal female reproductive events including folliculogenesis, ovulation, menstruation, embryo implantation and pregnancy. Disruptions in the endocrine/immune pathway, such as inflammation from endometriosis, create deleterious effects on reproductive function.

It is now well established that the growth of endometriotic lesions is dependent on estrogen. It has been suggested that estrogens are delivered to the ectopic lesions in an endocrine fashion; however, new evidence suggests that endometriotic lesions produce estrogen themselves [51]. This feed forward loop creates myriad of cell signaling cascades in the peritoneal microenvironment. Aromatase P450 (aromP450) is important for the production of estrogen as this enzyme catalyzes the reaction to produce estrone from

androsteneione and 17 β -hydroxysteroid dehydrogenase type 1 which is catalyzed into the biologically active form of estrogen, estradiol (E2) [14]. Aromatase P450 is significantly upregulated in the eutopic endometrium of women with endometriosis compared to healthy controls and aberrantly high expression is consistent regardless of disease state [52]. This suggests that in endometriosis patients, both the eutopic and ectopic endometrial tissue are involved in excess production of estrogen further. Additionally, in the presence of growth factors and pro-inflammatory cytokines including IL-1 β , TNF- α , IFN- γ and IL-17, cyclooxygenase-2 (COX-2) is induced and COX-2 catalyzes the synthesis of prostaglandin E₂ (PGE₂). Production of prostaglandins and cytokines have been suggested to facilitate infertility in women with endometriosis [19]. Increased expression of COX-2 mRNA is observed in the ectopic and eutopic endometrium of women with endometriosis compared to healthy controls [52]. E2 has been shown to stimulate COX-2 expression, which suggests that a positive feedback loop exists [53]. Together, aberrant expressions of COX-2 and aromP450 produce a local, continuous stream of E2 and PGE₂ in endometriosis patients [52, 54] creating a state of estrogen dominance [14]. Estrogen, in particular, plays a critical role in female reproductive events including oocyte maturation, fertilization, ovulation and implantation. Estrogen initiates proliferation and differentiation of granulosa cells and facilitates the actions of luteinizing hormone and follicle stimulating hormone, which are important for folliculogenesis and ovulation [55]. Estrogen also stimulates growth of the uterine lining, which is important for uterine receptivity and implantation of a fertilized embryo at specific time periods. As such, continual high expression of estrogen likely interferes with the transition from proliferative to secretory phase and other important reproductive events. A state of estrogen dominance is also detrimental as estrogen has been shown to be an inhibitor of $\alpha v \beta 3$ integrin, a critical marker for endometrial receptivity, in the uterine lining [56]. When analyzing fertility, the receptivity of the endometrium is a critical factor for successful attachment, implantation and pregnancy [57] and therefore, inhibition of integrins and adhesion molecules that facilitate receptivity likely disturbs fertility.

In addition to aberrant production of sex-hormones, the aberrant expression and signalling of the progesterone receptor (PR) has been demonstrated in women with endometriosis [14]. A high expression of the decoy receptor, progesterone A receptors (PRA), relative to progesterone B receptors (PRB) has been found in the eutopic endometrium of women with endometriosis [58]. Reduced expression of PRB prevents appropriate progesterone signalling and this progesterone resistance has been categorized as a “hallmark for implantation failure” since progesterone facilitates decidualization [14]. Emerging evidence suggests that expression of

both estrogen receptors (ER) and PR are altered by inflammation. Specifically, aberrant expression of ER and PR has been associated with an overexpression of IL-1, IL-6 and TNF- α [59]. Grandi et al demonstrated that TNF- α and IL-1 β (upregulated in the PF of women with endometriosis) cause a decrease in the expression of PRA and PRB mRNA in endometrial stromal cells isolated from women with endometriosis [60]. Additionally, Heublein et al demonstrated that TNF- α down regulates the expression of G-protein coupled estrogen receptor (GPER) in endometrial stromal cells isolated from women with endometriosis and the presence of GPER has been suggested to act as a selector that is important for folliculogenesis and follicle maturation [61]. The presented evidence demonstrates not only that aberrant expression of PR and ER are correlated with an increase in inflammatory mediators but that inflammation can directly alter the expression of both PR and GPER.

While treating endometriosis patients using hormonal intervention was previously emphasized, the regulation of inflammation and hormones appear to be interconnected and complex. Therefore, a comprehensive understanding of immune dysfunction in endometriosis patients is of the utmost importance to treat endometriosis and endometriosis associated infertility.

IMPLICATION OF IMMUNE DYSFUNCTION ON OOCYTE QUALITY, SPERM MOTILITY AND EMBRYOTOXICITY

Retrospective analyse of *in vitro* fertilization (IVF) and oocyte donation programs consistently find that women with endometriosis have significantly reduced pregnancy rates per cycle and per transfer as well as reduced implantation rates [62-64]. They also find that healthy ovum donation to endometriosis patients produces the same rate of implantation and pregnancy compared to controls [62-64]. Additionally, retrospective and prospective clinical trials using IVF have shown decreased oocyte and embryo quality and low ovarian reserves in women with endometriosis compared to controls [65, 66]. Collectively, human studies indicate poor oocyte and embryo quality and lower pregnancy rates in women with endometriosis.

Because the female reproductive organs, including the uterus, ovaries and fallopian tubes, are bathed in PF, pro-inflammatory cytokines and chemokines interact with the oocyte and embryo, which can impact growth and can inflict damage to the oocyte and embryo. A damaging inflammatory *milieu* in endometriosis patients offers a plausible theory to explain why the quality of oocytes and embryos is lower. Intra-follicular levels of IL-8, IL-12 and adrenomedullin are elevated in women with endometriosis undergoing IVF and are indicators of impaired embryo and oocyte quality [67]. In retrospective IVF studies, poor oocyte quality

was observed and measured by diminished blastomere cleavage rates, increased numbers of arrested embryos and impaired cytosolic events [68-71]. Sperm, travelling through the uterus and fallopian tubes, also interact with inflammatory cytokines in the PF and similarly encounter damage. Inflammatory cytokines including TNF- α and oxidative stress has been shown to directly hinder sperm motility [72]. Similarly, murine embryos incubated in the PF from women with endometriosis have shown diminished growth rates of embryos, increased rates of apoptosis, DNA fragmentation and increased number of embryos arrested in development [73-76]. Dexamethasone reduced the observed embryotoxic effect of the PF from women with endometriosis associated infertility [77]. Dexamethasone is a glucocorticoid that has been shown to reduce the expression of prostaglandins and other inflammatory mediators dysregulated in endometriosis [78]. Additionally, inhibiting TNF- α reduces embryotoxic effect on mouse embryos incubated with PF from infertile women with endometriosis [79]. Collectively, these studies link inflammation in the PF, specifically TNF- α , with embryo toxicity. Studying the toxicity of PF from women with endometriosis is limited by ethical constraints as interfering with human embryos violates moral and ethical considerations. However, this murine model provides a convincing argument to suggest the PF from women with endometriosis produces a damaging effect on the embryo.

IMPLICATION OF IMMUNE DYSFUNCTION ON OXIDATIVE STRESS

An imbalance between oxidants and antioxidants creates oxidative stress [80]. This ratio can be altered by an increase in the expression of oxidants such as reactive oxygen species (ROS) and reactive nitrogen species or by a decrease in the expression of antioxidants such as superoxide dismutase, glutathione or vitamin E [81]. Many inflammatory diseases have been correlated with high oxidative stress including rheumatoid arthritis, cardiovascular disease, fibrosis and pre-eclampsia [82-85]. An increase in pro-inflammatory cytokines generates ROS and induces apoptosis or necrosis by activating transcription factors such as AP-1, p53 and NF- κ B [81]. In addition, estrogen and metabolites of estrogen have been shown to generate ROS, which have been shown to contribute to breast cancer [86, 87].

Higher levels of oxidative stress markers have been observed in the PF of women with endometriosis specifically, high concentrations of malondialdehyde (MDA) and oxidized low density lipoprotein [88]. Elevated concentrations of a lipid peroxidation metabolite, 8-iso-prostaglandin F₂- α , has also been detected in the PF and urine of women with endometriosis and has been speculated to play a role in infertility associated with endometriosis [88]. In another study, nine infertile

women with endometriosis were shown to have an increased concentration and increased activity of nitric oxide synthase (NOS), an enzyme found in peritoneal macrophages, compared to fertile controls [89]. Other investigators have failed to find increased oxidative stress markers in the PF of women with endometriosis [90]. Why we observe such differences is unknown. Similarly, high concentrations of oxidative stress molecules and low levels of antioxidative molecules have been found in the follicular fluid from women with endometriosis. Singh et al detected higher levels of ROS, NOS and MDA in the follicular fluid of women with endometriosis and another study detected an increased concentration of 8-hydroxy-2'-deoxyguanosine, a metabolite of DNA oxidative damage, in the follicular fluid of specifically infertile women with endometriosis [91, 92]. In addition, the concentrations of critical antioxidants are aberrantly expressed in endometriosis patients. Levels of glutathione are diminished in the follicular fluid of women with endometriosis [93] and low levels of other antioxidants including vitamin A, C and E were found to be significantly reduced in the intra-follicular fluid of women with endometriosis associated infertility compared to women with tubal infertility [92]. As expected, higher levels of glutathione have been correlated with higher quality embryos [93]. Correspondingly, high concentrations of oxidative stress metabolites and lower concentrations of antioxidant molecules are associated with less successful IVF treatments [92].

Oxidative stress offers a plausible mechanism to link inflammation and infertility as IL-1 β and TNF- α are able to activate apoptotic mechanisms [81] and oocytes from women with endometriosis have exhibited increased rates of apoptosis in the cumulus cells [94]. Ovarian cell

apoptosis is used as an indicator for low oocyte quality [95] and therefore suggests that immune induced oxidative stress may be directly deteriorating oocyte quality.

THERAPEUTIC INTERVENTION: TARGETING INFLAMMATION AND IMMUNE DYSFUNCTION

After twenty years of research, unfortunately, no new treatments have come to the market to relieve the symptoms of endometriosis. While inhibiting inflammatory cytokines, specifically TNF- α had shown promise, anti-TNF- α treatment did not improve fertility in a non-human primate model and a clinical trial in human patients inhibiting TNF- α did not provide pelvic pain relief compared to a placebo [96, 97]. More recently, inhibition of the mitogen activated protein kinase (MAPK) cascade has emerged as a potential therapeutic for endometriosis, as it is activated by pro-inflammatory cytokines and oxidative stress and causes recruitment of immune cells to amplify the inflammation [54]. Sorafenib and Vemurafenib are different inhibitors of the MAPK cascade and have shown therapeutic promise both *in vitro* and in a mouse model by targeting endometriosis related inflammation [98, 99]. Human clinical trials of Sorafenib have been conducted and are underway for treatment of cancers including hepatocellular carcinoma, bladder cancer, acute myeloid leukemia (www.clinicaltrials.gov). Vermurefenib was used in human clinical trials for the treatment of advanced thyroid cancer and melanoma (www.clinicaltrials.gov). However, to our knowledge, human clinical trials using Sorafenib or Vemurafenib for the treatment of endometriosis have not been attempted. Many other inhibitors exist for a variety of targets in the MAPK cascade with the intention to reduce inflammation,

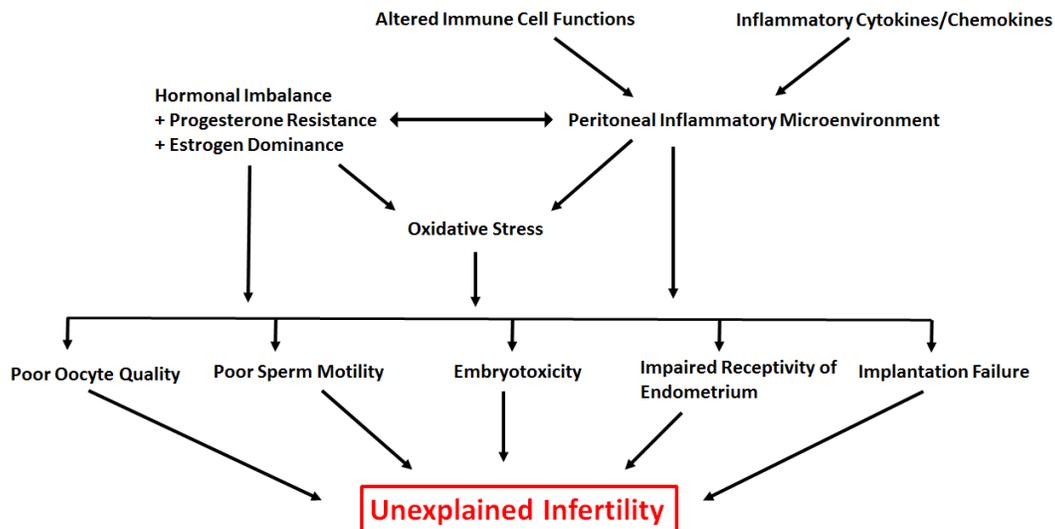


Figure 1: Overview of the underlying contributors to unexplained infertility associated with endometriosis. Hierarchical arrows indicate a potential link between various factors contributing to unexplained infertility associated with endometriosis.

proliferation and angiogenesis through cytokines and chemokines; however, these seem, to our knowledge, to be in the early stages of research and development [54]. Overall, the upstream regulatory factors of inflammatory cascades are emerging as the next therapeutic targets for endometriosis and endometriosis associated infertility.

CONCLUSIONS AND FUTURE DIRECTIONS

Here we provided evidence from the literature and from our own work to understand how the immune system is potentially dysfunctional in endometriosis patients with infertility issues. We also provided insights into the implications of this dysfunction (summarized in Figure 1). There is substantial evidence to suggest that aberrant immune mechanisms in endometriosis are associated with a number of factors that have the ability to deteriorate fecundity including folliculogenesis, oocyte and embryo quality and eutopic receptivity/implantation failure. However, it remains unclear how immune dysregulation is contributing towards pathogenesis of this enigmatic disease. At this time, it is also unclear if a disruption of one, some or all factors lead to infertility. As stated earlier, there has been limited progress in terms of new treatment strategies to manage endometriosis related infertility. Further understanding of the mechanisms and complex interplay between immune-endocrine axis may help explain why we observe heterogeneity of the symptoms in patients and may stimulate the process of developing a more comprehensive and accurate classification system.

Previous therapies for endometriosis have targeted downstream, end products such as levels of estrogen, TNF- α and COX-2. Unfortunately, these therapeutics fail to uniformly reduce symptoms, specifically infertility, and fail to cure the disease. Targeting upstream, regulatory mediators seems rational for future action. Many studies correlate endometriosis and its associated infertility with an up or down regulation of immune products; however, few analytical and explanatory studies exist with the objective to understand why we observe immune dysfunction. Are the increased numbers of infiltrating cell types (if any) in the peritoneal cavity the problem or their functional incompetence for example activation status or cytotoxicity causing dysfunction? Additionally, the complex connection between inflammation and the hormonal imbalance observed is poorly understood. Investigators studying endometriosis seem to have exhausted intuitive explanations to understand the pathogenesis of endometriosis and its associated infertility. Therefore, there is a pressing need at this time in endometriosis research, similar to other chronic inflammatory conditions including cancer, to shift the focus of investigation to understand the complex immunological pathways leading to the disease in order to treat and cure endometriosis along with its infertility.

CONFLICTS OF INTERESTS

The authors declare no conflict of interests.

REFERENCES

1. Levy AR, Osenenko KM, Lozano-Ortega G, Sambrook R, Jeddi M, Bélisle S, Reid RL. Economic burden of surgically confirmed endometriosis in Canada. *J Obstet Gynaecol Can.* 2011; 33: 830-7.
2. Simoens S, Dunselman G, Dirksen C, Hummelshoj L, Bokor A, Brandes I, Brodzky V, Canis M, Colombo GL, DeLeire T, Falcone T, Graham B, Halis G, et al. The burden of endometriosis: costs and quality of life of women with endometriosis and treated in referral centres. *Hum Reprod.* Oxford University Press; 2012; 27: 1292-9. doi: 10.1093/humrep/des073.
3. Halme J, Hammond MG, Hulka JF, Raj SG, Talbert LM. Retrograde menstruation in healthy women and in patients with endometriosis. *Obstet.* 1984; 64: 151-4. doi: 10.1016/0261-5614(91)90061-G.
4. Ahn SH, Monsanto SP, Miller C, Singh SS, Thomas R, Tayade C. Pathophysiology and immune dysfunction in endometriosis. *BioMed Research International.* 2015. doi: 10.1155/2015/795976.
5. Kwak J-Y, Park S-W, Kim K-H, Na Y-J, Lee K-S. Modulation of neutrophil apoptosis by plasma and peritoneal fluid from patients with advanced endometriosis. *Hum Reprod.* 2002; 17: 595-600. doi: 10.1016/S0015-0282(01)03048-5.
6. Beste MT, Pfäffle-Doyle N, Prentice EA, Morris SN, Lauffenburger DA, Isaacson KB, Griffith LG, Eskenazi B, Warner ML, Giudice LC, Herington JL, Bruner-Tran KL, Lucas JA, et al. Molecular network analysis of endometriosis reveals a role for c-Jun-regulated macrophage activation. *Sci Transl Med.* 2014; 6: 222ra16. doi: 10.1126/scitranslmed.3007988.
7. Jeung I, Cheon K, Kim M-R, Jeung I, Cheon K, Kim M-R. Decreased Cytotoxicity of Peripheral and Peritoneal Natural Killer Cell in Endometriosis. *Biomed Res Int.* 2016; 2016: 1-6. doi: 10.1155/2016/2916070.
8. Schulke L, Berbic M, Manconi F, Tokushige N, Markham R, Fraser IS. Dendritic cell populations in the eutopic and ectopic endometrium of women with endometriosis. *Hum Reprod.* 2009; 24: 1695-703. doi: 10.1093/humrep/dep071.
9. Hever A, Roth RB, Hevezi P, Marin ME, Acosta JA, Acosta H, Rojas J, Herrera R, Grigoriadis D, White E, Conlon PJ, Maki RA, Zlotnik A. Human endometriosis is associated with plasma cells and overexpression of B lymphocyte stimulator. *Proc Natl Acad Sci.* 2007; 104: 12451-6. doi: 10.1073/pnas.0703451104.
10. Ho HN, Wu MY, Chao KH, Chen CD, Chen SU, Yang YS. Peritoneal interleukin-10 increases with decrease in activated CD4+ T lymphocytes in women with

- endometriosis. *Hum Reprod.* 1997; 12: 2528-33.
11. Taylor RN, Lebovic DI, Mueller MD. Angiogenic Factors in Endometriosis. *Ann N Y Acad Sci.* Blackwell Publishing Ltd; 2002; 955: 89-100. doi: 10.1111/j.1749-6632.2002.tb02769.x.
 12. Harada T, Iwabe T, Terakawa N. Role of cytokines in endometriosis. *Fertility and Sterility.* 2001. p. 1-10. doi: 10.1016/S0015-0282(01)01816-7.
 13. McKinnon BD, Bertschi D, Bersinger NA, Mueller MD. Inflammation and nerve fiber interaction in endometriotic pain. *Trends Endocrinol Metab.* 2015; 26: 1-10. doi: 10.1016/j.tem.2014.10.003.
 14. Fox C, Morin S, Jeong J-W, Scott RT, Lessey BA. Local and systemic factors and implantation: what is the evidence? *Fertil Steril.* 2016; 105: 873-84. doi: 10.1016/j.fertnstert.2016.02.018.
 15. Sensky TE, Liu DT. Endometriosis: associations with menorrhagia, infertility and oral contraceptives. *Int J Gynaecol Obstet.* 1980; 17: 573-6.
 16. Schwartz D, Mayaux MJ, Mayaux MJ. Female Fecundity as a Function of Age. *N Engl J Med.* 1982; 306: 404-6. doi: 10.1056/NEJM198202183060706.
 17. Hughes EG, Fedorkow DM, Collins JA. A quantitative overview of controlled trials in endometriosis-associated infertility. *Fertil Steril.* 1993; 59: 963-70.
 18. de Ziegler D, Borghese B, Chapron C. Endometriosis and infertility: pathophysiology and management. *Lancet.* 2010; 376: 730-8. doi: 10.1016/S0140-6736(10)60490-4.
 19. Bulun SE. Endometriosis. *N Engl J Med.* 2009; 360: 268-79. doi: 10.1056/NEJMra0804690.
 20. Mousa NA, Bedaiwy MA, Casper RF. Aromatase inhibitors in the treatment of severe endometriosis. *Obstet Gynecol.* 2007; 109: 1421-3. doi: 10.1097/01.AOG.0000265807.19397.6d.
 21. Talukdar N, Bentov Y, Chang PT, Esfandiari N, Nazemian Z, Casper RF. Effect of long-term combined oral contraceptive pill use on endometrial thickness. *Obstet Gynecol.* 2012; 120: 348-54. doi: 10.1097/AOG.0b013e31825ec2ee.
 22. Marcoux S, Maheux R, Bérubé S. Endometriosis the CCG on. Laparoscopic Surgery in Infertile Women with Minimal or Mild Endometriosis. *N Engl J Med.* 1997; 337: 217-22. doi: 10.1056/NEJM199707243370401.
 23. Jacobson TZ, Duffy JM, Barlow DH, Farquhar C, Koninckx PR, Olive D. Laparoscopic surgery for subfertility associated with endometriosis. In: Jacobson TZ, editor. *Cochrane Database of Systematic Reviews.* 2010. doi: 10.1002/14651858.CD001398.pub2.
 24. Monsanto SP, Edwards AK, Zhou J, Nagarkatti P, Nagarkatti M, Young SL, Lessey BA, Tayade C. Surgical removal of endometriotic lesions alters local and systemic proinflammatory cytokines in endometriosis patients. *Fertil Steril.* 2016; 105: 968-77.e5. doi: 10.1016/j.fertnstert.2015.11.047.
 25. Ahn SH, Edwards AK, Singh SS, Young SL, Lessey BA, Tayade C. IL-17A Contributes to the Pathogenesis of Endometriosis by Triggering Proinflammatory Cytokines and Angiogenic Growth Factors. *J Immunol.* 2015; 195: 2591-600. doi: 10.4049/jimmunol.1501138.
 26. Eisermann J, Gast MJ, Pineda J, Odem RR, Collins JL. Tumor necrosis factor in peritoneal fluid of women undergoing laparoscopic surgery. *Fertil Steril.* 1988; 50: 573-9.
 27. Sikora J, Mielczarek-Palacz A, Kondera-Anasz Z. Imbalance in Cytokines from Interleukin-1 Family - Role in Pathogenesis of Endometriosis. *Am J Reprod Immunol.* 2012; 68: 138-45. doi: 10.1111/j.1600-0897.2012.01147.x.
 28. Harada T, Yoshioka H, Yoshida S, Iwabe T, Onohara Y, Tanikawa M, Terakawa N. Increased interleukin-6 levels in peritoneal fluid of infertile patients with active endometriosis. *Am J Obstet Gynecol.* 1997; 176: 593-7. doi: 10.1016/S0002-9378(97)70553-2.
 29. Punnonen J, Teisala K, Ranta H, Bennett B, Punnonen R. Increased levels of interleukin-6 and interleukin-10 in the peritoneal fluid of patients with endometriosis. *Am J Obstet Gynecol.* 1996; 174: 1522-6. doi: 10.1016/S0002-9378(96)70600-2.
 30. Bersinger NA, von Roten S, Wunder DM, Raio L, Dreher E, Mueller MD. PAPP-A and osteoprotegerin, together with interleukin-8 and RANTES, are elevated in the peritoneal fluid of women with endometriosis. *Am J Obstet Gynecol.* 2006; 195: 103-8. doi: 10.1016/j.ajog.2005.12.010.
 31. Yoshino O, Osuga Y, Koga K, Hirota Y, Tsutsumi O, Yano T, Morita Y, Momoeda M, Fujiwara T, Kugu K, Taketani Y. Concentrations of Interferon-gamma-Induced Protein-10 (IP-10), an Antiangiogenic Substance, are Decreased in Peritoneal Fluid of Women with Advanced Endometriosis. *Am J Reprod Immunol.* 2003; 50: 60-5. doi: 10.1034/j.1600-0897.2003.00037.x.
 32. Kats R, Collette T, Metz CN, Akoum A. Marked elevation of macrophage migration inhibitory factor in the peritoneal fluid of women with endometriosis. *Fertil Steril.* 2002; 78: 69-76. doi: 10.1016/S0015-0282(02)03189-8.
 33. Tariverdian N, Siedentopf F, Rucke M, Blois SM, Klapp BF, Kentenich H, Arck PC. Intraperitoneal immune cell status in infertile women with and without endometriosis. *J Reprod Immunol.* 2009; 80: 80-90. doi: 10.1016/j.jri.2008.12.005.
 34. Ahn SH, Khalaj K, Young SL, Lessey BA, Koti M, Tayade C. Immune-inflammation gene signatures in endometriosis patients. *Fertil Steril.* 2016; . doi: 10.1016/j.fertnstert.2016.07.005.
 35. Su R-W, Strug MR, Joshi NR, Jeong J-W, Miele L, Lessey BA, Young SL, Fazleabas AT. Decreased Notch Pathway Signaling in the Endometrium of Women With Endometriosis Impairs Decidualization. *J Clin Endocrinol Metab.* 2015; 100: E433-42. doi: 10.1210/jc.2014-3720.
 36. Ghosh D, Nagpal S, Bhat MA, Anupa G, Srivastava

- A, Sharma JB, Sengupta J. Gel-free proteomics reveals neoplastic potential in endometrium of infertile patients with stage IV ovarian endometriosis. *J Reprod Heal Med.* 2015; 1: 83-95. doi: 10.1016/j.jrhm.2015.06.003.
37. Acar N, Ustunel I, Demir R. Uterine natural killer (uNK) cells and their missions during pregnancy: A review. *Acta Histochem.* 2011; 113: 82-91. doi: 10.1016/j.acthis.2009.12.001.
 38. Jokhi PP, King a, Sharkey a M, Smith SK, Loke YW. Screening for cytokine messenger ribonucleic acids in purified human decidual lymphocyte populations by the reverse-transcriptase polymerase chain reaction. *J Immunol.* 1994; 153: 4427-35.
 39. Dosiou C, Giudice LC. Natural Killer Cells in Pregnancy and Recurrent Pregnancy Loss: Endocrine and Immunologic Perspectives. *Endocr Rev. Endocrine Society;* 2005; 26: 44-62. doi: 10.1210/er.2003-0021.
 40. Williams PJ, Bulmer JN, Searle RF, Innes BA, Robson SC. Altered decidual leucocyte populations in the placental bed in pre-eclampsia and foetal growth restriction: a comparison with late normal pregnancy. *Reproduction. Society for Reproduction and Fertility;* 2009; 138: 177-84. doi: 10.1530/REP-09-0007.
 41. Giuliani E, Parkin KL, Lessey BA, Young SL, Fazleabas AT. Characterization of Uterine NK Cells in Women with Infertility or Recurrent Pregnancy Loss and Associated Endometriosis. *Am J Reprod Immunol.* 2014; 72: 262-9. doi: 10.1111/aji.12259.
 42. Thiruchelvam U, Wingfield M, O'Farrelly C. Increased uNK Progenitor Cells in Women With Endometriosis and Infertility are Associated With Low Levels of Endometrial Stem Cell Factor. *Am J Reprod Immunol.* 2016; 75: 493-502. doi: 10.1111/aji.12486.
 43. Berbic M, Hey-Cunningham AJ, Ng C, Tokushige N, Ganewatta S, Markham R, Russell P, Fraser IS. The role of Foxp3⁺ regulatory T-cells in endometriosis: a potential controlling mechanism for a complex, chronic immunological condition. *Hum Reprod.* 2010; 25: 900-7. doi: 10.1093/humrep/deq020.
 44. Braundmeier A, Jackson K, Hastings J, Koehler J, Nowak R, Fazleabas A. Induction of endometriosis alters the peripheral and endometrial regulatory T cell population in the non-human primate. *Hum Reprod.* 2012; 27: 1712-22. doi: 10.1093/humrep/des083.
 45. Guerin LR, Prins JR, Robertson SA. Regulatory T-cells and immune tolerance in pregnancy: A new target for infertility treatment? *Human Reproduction Update.* 2009. p. 517-35. doi: 10.1093/humupd/dmp004.
 46. Chen S, Zhang J, Huang C, Lu W, Liang Y, Wan X, Giudice L, Kao L, Ulukus M, Cakmak H, Arici A, Simo'n C, Gutie'rrez A, et al. Expression of the T regulatory cell transcription factor FoxP3 in peri-implantation phase endometrium in infertile women with endometriosis. *Reprod Biol Endocrinol.* 2012; 10: 34. doi: 10.1186/1477-7827-10-34.
 47. Bommireddy R, Doetschman T. TGF-beta, T-cell tolerance and anti-CD3 therapy. *Trends Mol Med. NIH Public Access;* 2004; 10: 3-9.
 48. Young VJ, Brown JK, Saunders PTK, Duncan WC, Horne AW, Giudice L, Simoens S, Dunselman G, Dirksen C, Hummelshoj L, Bokor A, Candiani B, Fedele L, et al. The Peritoneum Is Both a Source and Target of TGF- β in Women with Endometriosis. Dettman R, editor. *PLoS One.* 2014; 9: e106773. doi: 10.1371/journal.pone.0106773.
 49. Jasper MJ, Tremellen KP, Robertson SA. Primary unexplained infertility is associated with reduced expression of the T-regulatory cell transcription factor Foxp3 in endometrial tissue. *Mol Hum Reprod.* 2006; 12: 301-8. doi: 10.1093/molehr/gal032.
 50. Zenclussen AC, Gerlof K, Zenclussen ML, Sollwedel A, Bertoja AZ, Ritter T, Kotsch K, Leber J, Volk H-D. Abnormal T-cell reactivity against paternal antigens in spontaneous abortion: adoptive transfer of pregnancy-induced CD4⁺CD25⁺ T regulatory cells prevents fetal rejection in a murine abortion model. *Am J Pathol.* 2005; 166: 811-22. doi: 10.1016/S0002-9440(10)62302-4.
 51. Bulun SE, Imir G, Utsunomiya H, Thung S, Gurates B, Tamura M, Lin Z. Aromatase in endometriosis and uterine leiomyomata. *J Steroid Biochem Mol Biol.* 2005; 95: 57-62. doi: 10.1016/j.jsbmb.2005.04.012.
 52. Bukulmez O, Hardy DB, Carr BR, Word RA, Mendelson CR. Inflammatory status influences aromatase and steroid receptor expression in endometriosis. *Endocrinology.* 2008; 149: 1190-204. doi: 10.1210/en.2007-0665.
 53. Tamura M, Deb S, Sebastian S, Okamura K, Bulun SE. Estrogen up-regulates cyclooxygenase-2 *via* estrogen receptor in human uterine microvascular endothelial cells. *Fertil Steril.* 2004; 81: 1351-6. doi: 10.1016/j.fertnstert.2003.09.076.
 54. Santulli P, Marcellin L, Tosti C, Chouzenoux S, Cerles O, Borghese B, Batteux F, Chapron C. MAP kinases and the inflammatory signaling cascade as targets for the treatment of endometriosis? *Expert Opin Ther Targets. Informa Healthcare;* 2015; 19: 1465-83. doi: 10.1517/14728222.2015.1090974.
 55. Drummond AE, Findlay JK. The role of estrogen in folliculogenesis. *Mol Cell Endocrinol.* 1999; 151: 57-64. doi: 10.1016/S0303-7207(99)00038-6.
 56. Somkuti SG, Yuan L, Fritz MA, Lessey BA. Epidermal growth factor and sex steroids dynamically regulate a marker of endometrial receptivity in Ishikawa cells. *J Clin Endocrinol Metab.* 1997; 82: 2192-7. doi: 10.1210/jcem.82.7.4102.
 57. Casper RF. It's time to pay attention to the endometrium. *Fertil Steril.* 2011; 96: 519-21. doi: 10.1016/j.fertnstert.2011.07.1096.
 58. Attia GR, Zeitoun K, Edwards D, Johns A, Carr BR, Bulun SE. Progesterone receptor isoform A but not B is expressed in endometriosis. *J Clin Endocrinol Metab.* 2000; 85: 2897-

902. doi: 10.1210/jcem.85.8.6739.
59. Florio P, Pinzauti S, Altomare A, Luisi S, Litta P, Petraglia F. Expression and role of peptides, proteins and growth factors in the pathogenesis of endometriosis. *J Endometr.* 2009; 1: 79-93. doi: 10.5301/JE.2009.2293.
 60. Grandi G, Mueller MD, Papadia A, Kocbek V, Bersinger NA, Petraglia F, Cagnacci A, McKinnon B. Inflammation influences steroid hormone receptors targeted by progestins in endometrial stromal cells from women with endometriosis. *J Reprod Immunol.* 2016; 117: 30-8. doi: 10.1016/j.jri.2016.06.004.
 61. Heublein S, Vrekoussis T, Kuhn C, Friese K, Makrigiannakis A, Mayr D, Lenhard M, Jeschke U. Inducers of G-protein coupled estrogen receptor (GPER) in endometriosis: potential implications for macrophages and follicle maturation. *J Reprod Immunol.* 2013; 97: 95-103. doi: 10.1016/j.jri.2012.10.013.
 62. Simón C, Gutiérrez A, Vidal A, de los Santos MJ, Tarín JJ, Remohí J, Pellicer A. Outcome of patients with endometriosis in assisted reproduction: results from in-vitro fertilization and oocyte donation. *Hum Reprod.* 1994; 9: 725-9.
 63. Díaz I, Navarro J, Blasco L, Simón C, Pellicer A, Remohí J. Impact of stage III-IV endometriosis on recipients of sibling oocytes: matched case-control study. *Fertil Steril.* 2000; 74: 31-4.
 64. Budak E, Garrido N, Soares SR, Melo MAB, Meseguer M, Pellicer A, Remohí J. Improvements achieved in an oocyte donation program over a 10-year period: sequential increase in implantation and pregnancy rates and decrease in high-order multiple pregnancies. *Fertil Steril.* 2007; 88: 342-9. doi: 10.1016/j.fertnstert.2006.11.118.
 65. Garrido N, Navarro J, Remohí J, Simón C, Pellicer A. Follicular hormonal environment and embryo quality in women with endometriosis. *Hum Reprod Update.* 6: 67-74.
 66. Kuroda M, Kuroda K, Arakawa A, Fukumura Y, Kitade M, Kikuchi I, Kumakiri J, Matsuo S, Brosens IA, Brosens JJ, Takeda S, Yao T. Histological assessment of impact of ovarian endometrioma and laparoscopic cystectomy on ovarian reserve. *J Obstet Gynaecol Res.* 2012; 38: 1187-93. doi: 10.1111/j.1447-0756.2012.01845.x.
 67. Singh AK, Dutta M, Chattopadhyay R, Chakravarty B, Chaudhury K. Intrafollicular interleukin-8, interleukin-12, and adrenomedullin are the promising prognostic markers of oocyte and embryo quality in women with endometriosis. *J Assist Reprod Genet.* 2016; . doi: 10.1007/s10815-016-0782-5.
 68. Brizek CL, Schlaff S, Pellegrini VA, Frank JB, Worriow KC. Increased incidence of aberrant morphological phenotypes in human embryogenesis—an association with endometriosis. *J Assist Reprod Genet.* 1995; 12: 106-12.
 69. Yanushpolsky EH, Best CL, Jackson K V, Clarke RN, Barbieri RL, Hornstein MD. Effects of endometriomas on oocyte quality, embryo quality, and pregnancy rates in *in vitro* fertilization cycles: a prospective, case-controlled study. *J Assist Reprod Genet.* 1998; 15: 193-7.
 70. Pellicer A, Oliveira N, Ruiz A, Remohí J, Simón C. Exploring the mechanism(s) of endometriosis-related infertility: an analysis of embryo development and implantation in assisted reproduction. *Hum Reprod.* 1995; 10 Suppl 2: 91-7.
 71. Tanbo T, Omland A, Dale PO, Abyholm T. *In vitro* fertilization/embryo transfer in unexplained infertility and minimal peritoneal endometriosis. *Acta Obstet Gynecol Scand.* 1995; 74: 539-43.
 72. Eisermann J, Register KB, Strickler RC, Collins JL. The Effect of Tumor Necrosis Factor on Human Sperm Motility *In Vitro.* *J Androl.* 1989 [cited 2016 Aug 22]; 10: 270-4. doi: 10.1002/j.1939-4640.1989.tb00100.x.
 73. Ding G-L, Chen X-J, Luo Q, Dong M-Y, Wang N, Huang H-F. Attenuated oocyte fertilization and embryo development associated with altered growth factor/signal transduction induced by endometriotic peritoneal fluid. *Fertil Steril.* 2010; 93: 2538-44. doi: 10.1016/j.fertnstert.2009.11.011.
 74. Rajani S, Chattopadhyay R, Goswami SK, Ghosh S, Sharma S, Chakravarty B. Assessment of oocyte quality in polycystic ovarian syndrome and endometriosis by spindle imaging and reactive oxygen species levels in follicular fluid and its relationship with IVF-ET outcome. *J Hum Reprod Sci.* 2012; 5: 187-93. doi: 10.4103/0974-1208.101020.
 75. Morcos RN, Gibbons WE, Findley WE. Effect of peritoneal fluid on *in vitro* cleavage of 2-cell mouse embryos: possible role in infertility associated with endometriosis. *Fertil Steril.* 1985; 44: 678-83.
 76. Esfandiari N, Falcone T, Goldberg JM, Agarwal A, Sharma RK. Effects of peritoneal fluid on preimplantation mouse embryo development and apoptosis *in vitro.* *Reprod Biomed Online.* 2005; 11: 615-9.
 77. Heitmann RJ, Tobler KJ, Gillette L, Tercero J, Burney RO. Dexamethasone attenuates the embryotoxic effect of endometriotic peritoneal fluid in a murine model. *J Assist Reprod Genet.* 2015; 32: 1317-23. doi: 10.1007/s10815-015-0516-0.
 78. Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids—new mechanisms for old drugs. *N Engl J Med.* 2005; 353: 1711-23. doi: 10.1056/NEJMra050541.
 79. Chen Y-J, Wu H-H, Liao W-T, Tsai C-Y, Tsai H-W, Chao K-C, Sung Y-J, Li H-Y. A tumor necrosis factor- α inhibitor reduces the embryotoxic effects of endometriotic peritoneal fluid. *Fertil Steril.* 2013; 100: 1476-85. doi: 10.1016/j.fertnstert.2013.07.1985.
 80. Al-Gubory KH, Fowler PA, Garrel C. The roles of cellular reactive oxygen species, oxidative stress and antioxidants in pregnancy outcomes. *Int J Biochem Cell Biol.* 2010; 42: 1634-50. doi: 10.1016/j.biocel.2010.06.001.
 81. Agarwal A, Aponte-Mellado A, Premkumar BJ, Shaman

- A, Gupta S. The effects of oxidative stress on female reproduction: a review. *Reprod Biol Endocrinol.* 2012; 10: 49. doi: 10.1186/1477-7827-10-49.
82. Lemarechal H, Anract P, Beaudoux J-L, Bonnefont-Rousselot D, Ekindjian OG, Borderie D. Impairment of thioredoxin reductase activity by oxidative stress in human rheumatoid synoviocytes. *Free Radic Res.* 2007; 41: 688-98. doi: 10.1080/10715760701294468.
 83. Schisterman EF, Faraggi D, Browne R, Freudenheim J, Dorn J, Muti P, Armstrong D, Reiser B, Trevisan M. TBARS and cardiovascular disease in a population-based sample. *J Cardiovasc Risk.* 2001; 8: 219-25.
 84. Huang QT, Zhang M, Zhong M, Yu YH, Liang WZ, Hang LL, Gao YF, Huang LP, Wang ZJ. Advanced glycation end products as an upstream molecule triggers ROS-induced sFlt-1 production in extravillous trophoblasts: a novel bridge between oxidative stress and preeclampsia. *Placenta.* 2013; 34: 1177-82. doi: 10.1016/j.placenta.2013.09.017.
 85. Kaviani N, Marut W, Servettaz A, Nicco C, Chéreau C, Lemaréchal H, Borderie D, Dupin N, Weill B, Batteux F. Reactive oxygen species-mediated killing of activated fibroblasts by arsenic trioxide ameliorates fibrosis in a murine model of systemic sclerosis. *Arthritis Rheum.* 2012; 64: 3430-40. doi: 10.1002/art.34534.
 86. Bolton JL. Quinoids, quinoid radicals, and phenoxyl radicals formed from estrogens and antiestrogens. *Toxicology.* 2002; 177: 55-65.
 87. Okoh V, Deoraj A, Roy D. Estrogen-induced reactive oxygen species-mediated signalings contribute to breast cancer. *Biochim Biophys Acta.* 2011; 1815: 115-33. doi: 10.1016/j.bbcan.2010.10.005.
 88. Sharma I, Dhaliwal LK, Saha SC, Sangwan S, Dhawan V. Role of 8-iso-prostaglandin F2alpha and 25-hydroxycholesterol in the pathophysiology of endometriosis. *Fertil Steril.* 2010; 94: 63-70. doi: 10.1016/j.fertnstert.2009.01.141.
 89. Osborn BH, Haney AF, Misukonis MA, Weinberg JB. Inducible nitric oxide synthase expression by peritoneal macrophages in endometriosis-associated infertility. *Fertil Steril.* 2002; 77: 46-51.
 90. Wang Y, Sharma RK, Falcone T, Goldberg J, Agarwal A. Importance portance of reactive oxygen species in the peritoneal fluid of women with endometriosis or idiopathic infertility. *Fertil Steril.* Elsevier; 1997; 68: 826-30. doi: 10.1016/S0015-0282(97)00343-9.
 91. Da Broi MG, de Albuquerque FO, de Andrade AZ, Cardoso RL, Jordão Junior AA, Navarro PA. Increased concentration of 8-hydroxy-2'-deoxyguanosine in follicular fluid of infertile women with endometriosis. *Cell Tissue Res.* 2016; . doi: 10.1007/s00441-016-2428-4.
 92. Singh AK, Chattopadhyay R, Chakravarty B, Chaudhury K. Markers of oxidative stress in follicular fluid of women with endometriosis and tubal infertility undergoing IVF. *Reprod Toxicol.* 2013; 42: 116-24. doi: 10.1016/j.reprotox.2013.08.005.
 93. Choi YS, Cho S, Seo SK, Park JH, Kim SH, Lee BS. Alteration in the intrafollicular thiol-redox system in infertile women with endometriosis. *Reproduction.* 2015; 149: 155-62. doi: 10.1530/REP-14-0438.
 94. Díaz-Fontdevila M, Pommer R, Smith R. Cumulus cell apoptosis changes with exposure to spermatozoa and pathologies involved in infertility. *Fertil Steril.* 2009; 91: 2061-8. doi: 10.1016/j.fertnstert.2008.05.073.
 95. Nakahara K, Saito H, Saito T, Ito M, Ohta N, Takahashi T, Hiroi M. The incidence of apoptotic bodies in membrana granulosa can predict prognosis of ova from patients participating in *in vitro* fertilization programs. *Fertil Steril.* 1997; 68: 312-7.
 96. Falconer H, Mwenda JM, Chai DC, Song X-YR, Cornillie FJ, Bergqvist A, Fried G, D'Hooghe TM. Effects of anti-TNF-mAb treatment on pregnancy in baboons with induced endometriosis. *Fertil Steril.* 2008; 89: 1537-45. doi: 10.1016/j.fertnstert.2007.05.062.
 97. Lu D, Song H, Shi G. Anti-TNF- α treatment for pelvic pain associated with endometriosis. *Cochrane database Syst Rev.* 2013; : CD008088. doi: 10.1002/14651858.CD008088.pub3.
 98. Ngô C, Nicco C, Leconte M, Chéreau C, Arkwright S, Vacher-Lavenu M-C, Weill B, Chapron C, Batteux F. Protein kinase inhibitors can control the progression of endometriosis *in vitro* and *in vivo*. *J Pathol.* 2010; 222: 148-57. doi: 10.1002/path.2756.
 99. Leconte M, Santulli P, Chouzenoux S, Marcellin L, Cerles O, Chapron C, Dousset B, Batteux F. Inhibition of MAPK and VEGFR by Sorafenib Controls the Progression of Endometriosis. *Reprod Sci.* 2015; 22: 1171-80. doi: 10.1177/1933719115592708.