

Haemonchus contortus excretory and secretory proteins (HcESPs) suppress functions of goat PBMCs *in vitro*

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ABSTRACT

Excretory and secretory products (ESPs) of nematode contain various proteins which are capable of inducing the instigation or depression of the host immune response and are involved in the pathogenesis of the worms. In the present study, *Haemonchus contortus* excretory and secretory products (HcESPs) were collected from the adult worms. Binding of HcESPs to goat peripheral blood mononuclear cells (PBMCs) was confirmed by immune-fluorescence assay. Effects of the HcESPs on cytokine production, cell proliferation, cell migration and nitric oxide (NO) production of PBMCs were checked by co-incubation of HcESPs with goat PBMCs. The results indicated that the production of IL-4 and IFN- γ were significantly decreased by HcESPs in dose dependent manner. On the contrary, the production of IL-10 and IL-17 were increased. Cell migration was significantly enhanced by HcESPs, whereas, HcESPs treatment significantly suppressed the cell proliferation and NO production. These results indicated that the HcESPs played important suppressive regulatory roles on PBMCs and provided highlights to the understanding of the host-parasite interactions.

INTRODUCTION

Gastrointestinal nematodes generally develop chronic infections and survive themselves in the host for a longer duration. The survival in the host reflects the ability of parasites to evade the host immune responses from the early stages of infection [1, 2]. Excretory and secretory products (ESPs) are produced and released by the parasites during *in vitro* cultivation [3] and *in vivo* [4]. ESPs contain various proteins and are capable to induce the depression or instigation of the host immune response and are related to the pathogenesis of the parasites [5-7]. Binding to the host cell is a prerequisite for ESP function [3, 8-11]. Some ESP molecules react to the molecules on the surface of the host cell to form receptor-ligand complexes, similar to many other receptor-ligand systems. For instance, galectin binds to β -galactoside sugars in a metal-independent manner [12, 13].

Host immune responses are usually inhibited by nematode ESPs through various mechanisms, such as interfering with antigen processing, suppression of macrophage and antigen-presenting cell function,

interference with cytokine signaling, or induction of immune-regulatory cell [14, 15]. It was reported that helminth ESPs induced the regulatory T cell (Treg) function in activated CD4⁺ T lymphocytes *in vitro* [15]. Klesius et al [16] showed that *Ostertagia ostertagi* could suppress the lymphocyte activation and lymphocyte proliferation during the pre-patent period of infection, whilst, Gomez et al [17] found that ESPs collected from *O. ostertagi* L4 were capable to inhibit the mitogen-induced bovine lymphocyte proliferation.

H. contortus excretory and secretory products (HcESPs) contain many proteins [18] that can perform various functions including the tissue penetration and host protein degradation [6]. A 55kDa secretory glycoprotein was identified with the ability to inhibit host neutrophils [8]. The purified 66-kDa adult *H. contortus* excretory/secretory (E/S) antigen was confirmed to suppress monocyte function *in vitro* by decreasing the production of hydrogen peroxide and nitric oxide in the culture medium of the cells [3]. Recently, it was reported that recombinant *H. contortus* galectin (rHco-gal-m) inhibited the expression of MHC II molecules, decreased the T cell

activation and proliferation, induced apoptosis of T cells and effected several signaling cascades *in vitro* [10]. *In vitro* studies also showed that ESPs had direct effects on cultured cells or tissues, such as inhibiting acid secretion [19] and inducing the vacuolation and detachment of HeLa cells [20, 21]. These findings indicated that ESPs might play multiple functions *in vivo*.

Up to now, some ESP molecules have been recognized and functionally characterized. However, in the natural infection of the nematode, the effects of the ESPs on the host cells and the final roles of the ESPs in the interactions of host and helminths might be dependent on the total ESPs, but not one or several molecules. In the present study, we evaluated the immune-modulatory effects of HcESPs on the goat PBMCs.

RESULTS

Confirmation of binding of HcESPs to goat PBMCs

Goat PBMCs were incubated with HcESPs and the binding was investigated by IFA. The emissions from

the Cy3-labeled HcESPs were red and the DAPI-labeled nuclei were blue. Some cells treated with HcESPs showed red fluorescence in the periphery of the blue nucleus. And no fluorescence was observed in the non-treated group. It indicated that HcESPs could bind to the PBMCs (Figure 1).

Effects of the HcESPs on the production of cytokines of PBMCs

Effects of HcESPs on the cytokine production were analyzed by ELISA and the results showed that HcESPs could modulate the cytokine production (Figure 2). Production of IL-4 and IFN- γ were significantly decreased in PBMCs incubated with different concentrations of HcESPs. The productions of IL-4 of 5 $\mu\text{g/ml}$ and 10 $\mu\text{g/ml}$ HcESPs treated groups were significantly lower than that of control group. No significant differences were observed between the control and the PBMCs treated with HcESPs at the dose of 20 $\mu\text{g/ml}$ and 40 $\mu\text{g/ml}$. Various concentrations of HcESPs decreased the IFN- γ levels. Contrary to that, the secretions of cytokine IL-10 and IL-17 were significantly increased by the all concentrations of HcESPs.

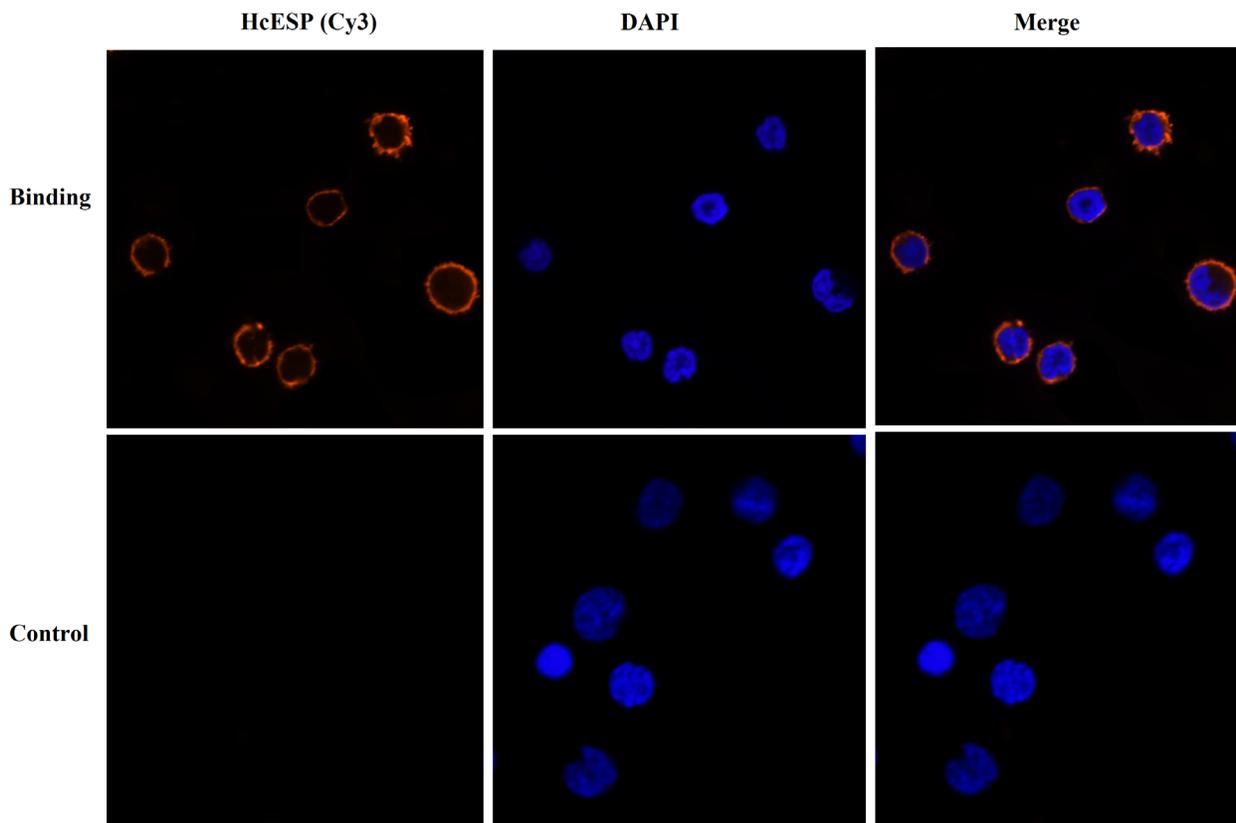


Figure 1: Confirmation of binding of HcESPs to goat PBMCs by IFA. The nuclei of the corresponding cells were visualized by DAPI (blue) staining. Staining of the target proteins (red) were visualized by Cy3-conjugated secondary antibody. Merge, overlap of red and blue channels. No red fluorescence was observed in control group.

Cell migration assay

The effect of the HcESPs on the cell migration was evaluated by a Transwell system (Corning, USA). The results showed that 10 μ g/ml and 20 μ g/ml of HcESPs significantly increased the cell migration compared to the control, but the 5 μ g/ml and 40 μ g/ml not.

Nitric oxide (NO) production

Nitric oxide (NO) production by PBMCs treated with different concentration of HcESPs was measured by using the total nitric oxide assay kit. The results revealed that HcESPs significantly suppressed the NO production by PBMCs in a dose dependent manner (Figure 4).

Cell proliferation

Cell counting kit (CCK8) was used to evaluate the effects of the HcESPs on the PBMC proliferation. Cell Proliferations were significantly suppressed by all concentrations of HcESPs compared with control group (Figure 5).

DISCUSSION

Nematode ESPs are molecules secreted by the worms *in vivo* or *in vitro* and played complex functions in the interactions between the parasites and the hosts. However, in the natural infection, the effects of the ESPs on the host and the final roles of the ESPs in the interactions of host and helminths might be dependent on the total ESPs, but not one or several molecules. In the present research, the effects of HcESPs on some functions of goat PBMCs were firstly investigated. The results

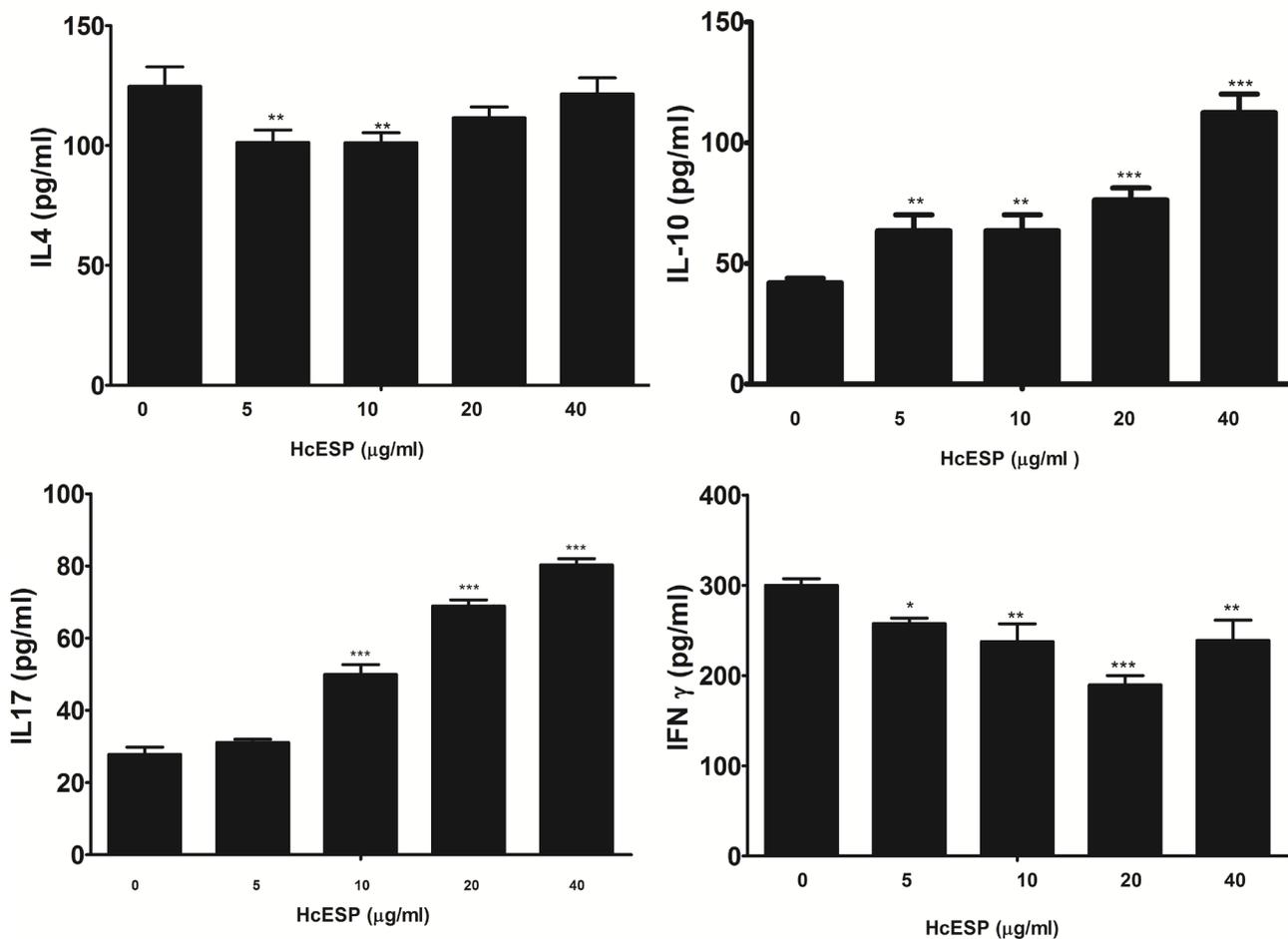


Figure 2: Analysis of the level of multiple cytokine production by PBMCs *in vitro*. PBMCs were stimulated with ConA (10 μ g/ml) for 24h in the presence or absence of various concentrations of HcESPs. Cytokine secretion in the supernatant of cell cultures was quantified by ELISA. The data are representative of three independent experiments (* p < 0.01, ** p < 0.001, ns non significant).

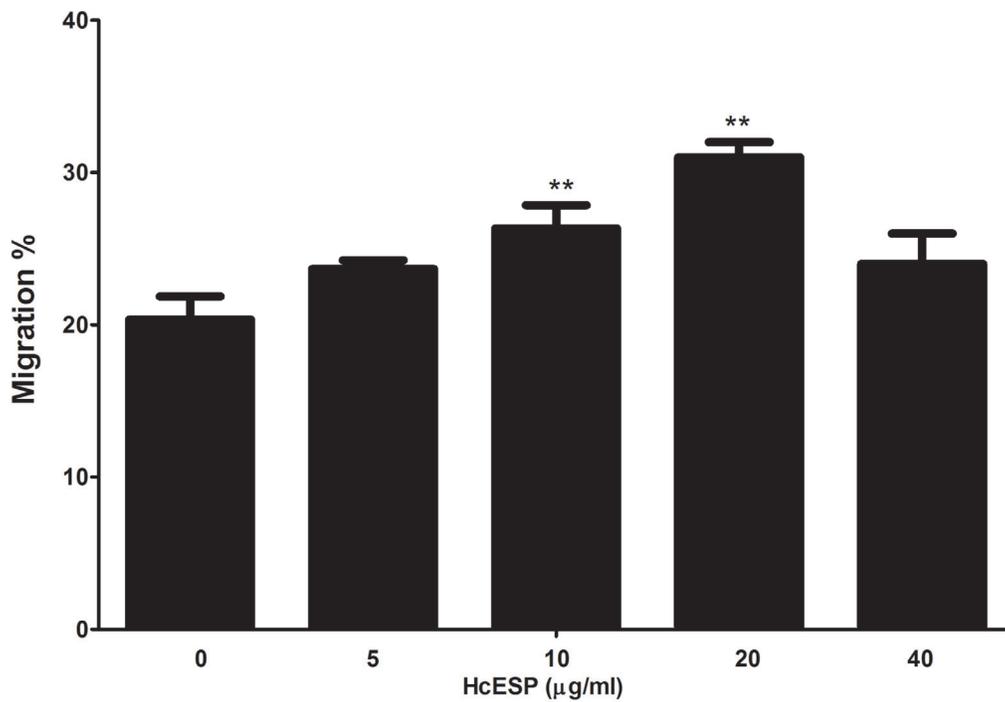


Figure 3: Effects of HcESPs on PBMC migration. PBMC were treated with control buffer and various concentrations of HcESPs, Then the random migration was determined. The difference between the mean values was calculated using ANOVA. Data are representative of 3 independent experiments (** $p < 0.001$ and *** $p < 0.0001$).

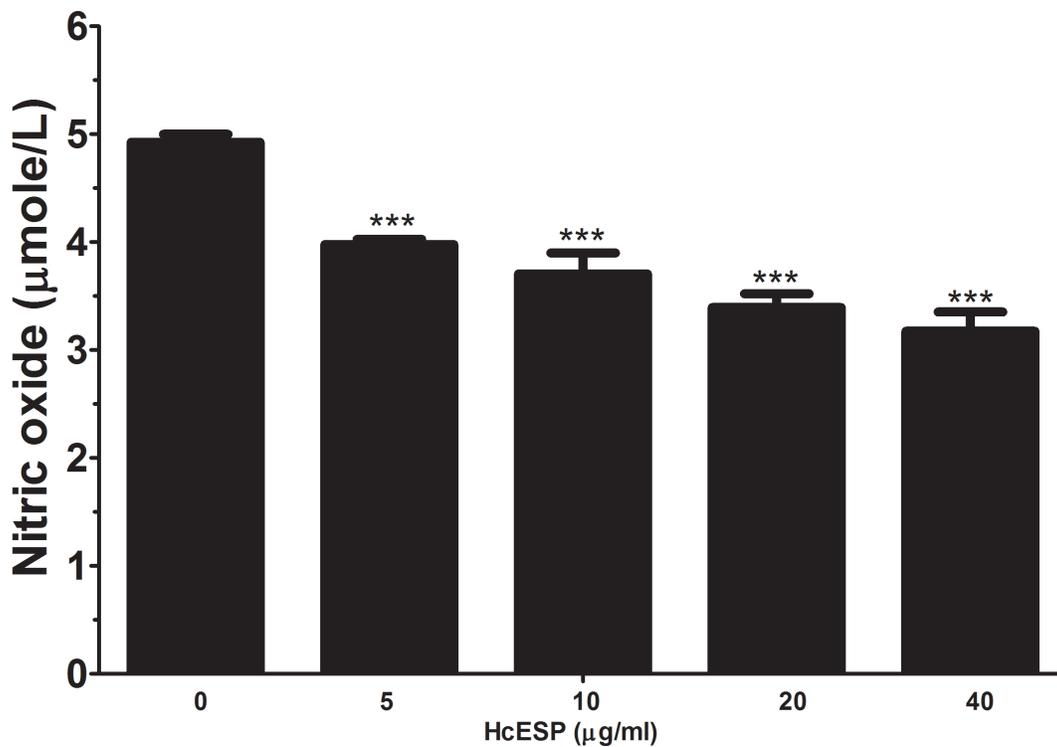


Figure 4: Effects of HcESPs on nitric oxide production by PBMCs *in vitro*. Cells were activated with ConA and incubated at the same time with serial concentrations of HcESPs at 37°C and 5% CO₂. The nitrite concentration in the PBMCs was measured by using the Griess assay and used as an indicator of nitric oxide production by the PBMCs. The data were representative of three independent experiments (*** $p < 0.0001$).

showed that HcESPs presented suppressive potential on the PBMCs. It showed highlight to the understanding of the interactions of the host and the worms.

It was reported that Type 2 (Th2) and Type 1 (Th1) immune responses and tissue inflammation played important roles in the resistance to the nematodes infections including *H. contortus* [22-24]. Jacobs et al [25] demonstrated that early IL-4 expression was associated with resistance to *Haemonchus contortus* in the resistance sheep breeds. The immune and inflammatory responses were strongly mediated by some cell lines and cytokines they produced. Type 1 and Type 2 responses were positively mediated by Th1 and Th2 cells and their typical cytokines, IFN- γ and IL-4, respectively. Another cell line, T regulatory cells (TReg), and its typical cytokine IL-10 usually played suppressive functions in the immune responses. It inhibited the development of Th2 cell responses [26-28] and decreased the production of IFN- γ [29, 30]. In this study, HcESPs significantly inhibited the productions of IL-4 and IFN- γ , increased the level of IL-10. These results suggested that HcESPs could suppress Type 2 and Type 1 immune responses simultaneously and strengthened the suppression by increasing the production of IL-10. This might be one of the immune evasion mechanisms of *H. contortus*.

Th17 cells and their typical cytokine, IL-17, were functionally characterized as tissue inflammatory

modulator [31]. It was demonstrated that neutrophil activation and tissue damage were reduced in IL-17 deficient animals and the increased level of IL-17 was related to the pathogenesis of various parasites [32-36]. In the present research, the IL-17 secretion was significantly increased by HcESPs. This finding suggested that HcESPs could induce the Th17 cells. The enhanced IL-17 level might favour the survival of the worm in host. However, the real relationship of IL-17 and pathogenesis need to be further investigated.

NO was produced by macrophages activated by IFN- γ and TNF- α and usually played killing functions on the helminths [37, 38]. In the present study, the production of NO was significantly suppressed by HcESPs. This indicated that the HcESPs could inhibit the production of NO through decreasing IFN- γ level or other ways, and thus alleviated the harmful effects of some chemical factors produced by the host cells on the helminths.

Cell proliferation and migration are very important to the developments of immune responses. Cell proliferation increased the numbers of the effector cells and cell migration recruited the effector cells to the sites of infection. It was reported that helminths actively stimulated eosinophils and other lymphocyte to migrate to the sites of infection and resulted in tissue damage [39, 40]. In this study, it was identified that HcESPs significantly suppressed the goat PBMCs proliferation

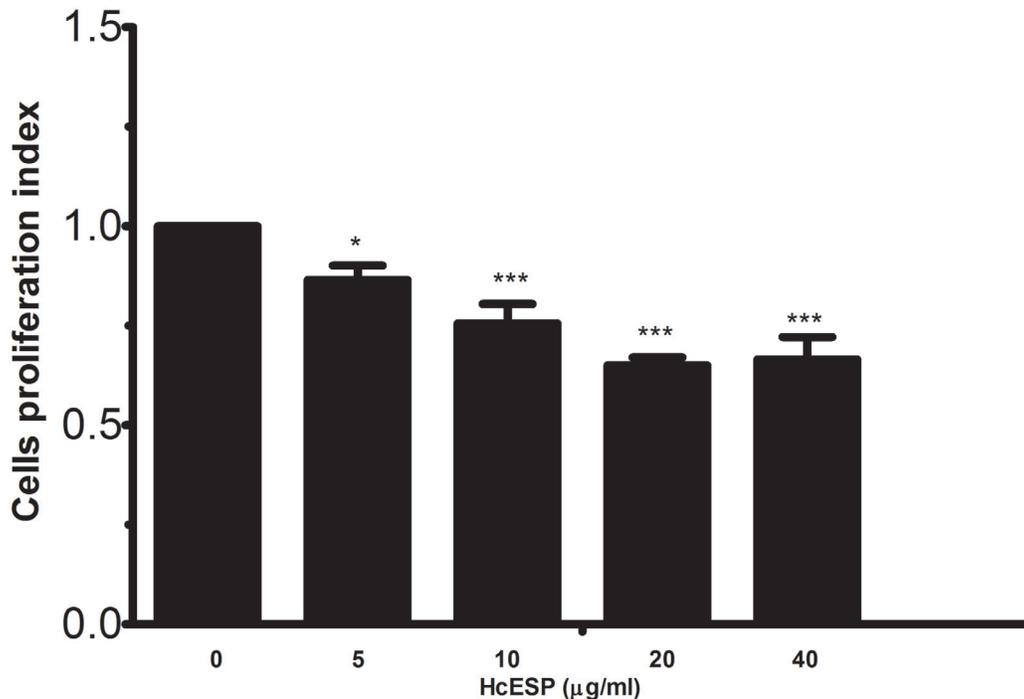


Figure 5: Effects of HcESPs on PBMCs proliferation. Cells were activated with ConA and incubated at the same time with serial concentrations of HcESPs at 37°C and 5% CO₂. The proliferation was measured by CCK-8 incorporation after 72 h. Cell proliferation index was calculated considering the OD₄₅₀ values in controls as 100%. The data were representative of three independent experiments (* p < 0.01 and *** p < 0.0001).

and significantly enhanced the cell migration. These results showed that HcESPs also inhibited the Type 2 and Type 1 responses through decreasing the numbers of the effector cells. However, the real roles of the increased cell migration in the immune responses should be investigated.

In our previous studies, we found that recombinant galectin of male and female adult *H. contortus* displayed different effects on the T cells and monocytes [10, 41]. Thus, it was suspected that the ESPs from males and females also had different functions. However, in the natural infection of this worm the ESPs from males and females were secreted and functioned simultaneously. So in this research, we used the mixed ESPs from males and females. Thus, differentiating the different effects of ESPs from males and females worms on PBMCs is worth of further investigating. PBMCs are consisted of lymphocytes (T and B), monocytes, DCs and other cells. Perez et al [42] reported that T cells, particularly CD4⁺ and $\gamma\delta^+$ lymphocytes, in the abomasal mucosa of goats infected with *H. contortus* were mildly increased at 3 and 6 days post infection (dpi) and marked increased at 10 and 13 dpi. B cells and IgG+ plasma cells also showed a marked increase in the abomasal mucosa at 10 and 13 dpi. Whereas the increases in CD8⁺ cells were less pronounced. These results indicated that the infection of *H. contortus* could change the cell populations *in vivo*. In this research, it was demonstrated that HcESPs could effect the functions of the PBMCs of goat. However, the effects of HcESPs on the populations of PBMCs were not studied. Thus, the cell populations that govern the functional changes of PBMCs treated with HcESPs and the protein or proteins that induce the changes *in vitro*, as well as *in vivo*, need to be further probed.

Conclusively, this study identified that the HcESPs displayed suppressive potential on the goat PBMCs *in vitro*. It inhibited the productions of IL-4, IFN- γ , increased the suppressive cytokine IL-10, enhanced the inflammatory modulator IL-17, suppressed the production of chemical factor NO, decreased the cell proliferation and activated the cell migration. These results are favorable to the profoundly understanding of the immune invasion and the host-parasite interactions of *H. contortus* and other gastrointestinal nematode. However, for complex life cycle of *H. contortus*, the real effects of the HcESPs on the PBMCs *in vivo* need to be further investigated. Although the functions of some molecules of HcESPs had been characterized, the molecule or the molecules that governed the suppressive functions on the PBMCs also need to be further studied for the huge numbers of the molecules of HcESPs.

MATERIALS AND METHODS

Ethics statement

Animal experiments were conducted following the guidelines of the Animal Ethics Committee, Nanjing Agricultural University, China. All experimental protocols were approved by the Science and Technology Agency of Jiangsu Province. The approval ID is SYXK (SU) 2010-0005.

Collection of adult *H. contortus* worms and *in vitro* production of excretory and secretory products (HcESPs)

Two local male goats (02 years old) were raised under nematode free condition and dewormed twice at 15 days interval by anthelmintic drug. Both goats were orally infected with 10,000 infective stage larvae (L₃) of *H. contortus*. For the confirmation of the infection, faecal sample was collected and checked after weekly intervals for the presence of *H. contortus* eggs. After confirmation of infection, goats were euthanized and killed at 27th day post infection. The abomasum was tied off at both ends and detached from the remaining digestive tract. Mixed adult worms (male and female) were collected from the abomasums, washed several times in PBS, and kept in RPMI 1640 medium (100 worms/ml) containing 100 IU of penicillin and 0.1 mg/ml streptomycin (Pen strep, gibco, Life Technologies) at 37 °C under 5% CO₂. The parasites were initially incubated for 4 hours, after than the medium was collected and new medium containing 2% glucose was added and incubated for overnight. At the end of incubation period, the cultured media was centrifuged and the supernatant was collected. Collected supernatant was filter-sterilized by using the 0.2 μ m pore size membrane filter. HcESP from the collected supernatant was concentrated and desalted (10 mM Tris, NaCl pH7.4) by using the 3-kDa filters (Centriprep YM-3, Millipore). The protein concentration was checked by Bradford method (Bradford, 1976).

Production polyclonal IgG against HcESP (IgGHcESP)

SD Rats were used for the generation of polyclonal antibodies against *H. contortus* excretory and secretory proteins (IgGHcESP). HcESP protein of 400 μ g was mixed with Freund's complete adjuvant (1:1) and injected into SD rats subcutaneously [39, 43]. The booster doses were injected four times at 2-week intervals with the same dose to the first immunization and the HcESP was mixed with Freund's incomplete adjuvant at 1:1. One week after

the last injection the rats were bled and the sera containing specific anti-HcESP antibodies were collected and stored at -20°C. Blank sera were collected before starting the immunization and kept as negative control.

Isolation of goat PBMCs and confirmation of HcESP binding to PBMCs

Heparinized blood was collected by vein puncture from dewormed healthy goats. PBMCs were separated with the standard Ficoll-hypaque (GE Healthcare, USA) gradient centrifugation method [44] and washed twice in Ca²⁺/Mg²⁺- free PBS pH 7.4. Cell viability assessed by means of the trypan blue exclusion test was consistently >95%. The PBMC were re-suspended to a final density of 1×10⁵ cells/ml in RPMI 1640 medium containing 10% heat inactivated fetal calf serum (FCS), 100 U/ml penicillin and 100 mg/ml streptomycin Pen Strep (Penicillin and Streptomycin) (gibco, Life Technology). PBMCs were incubated in the presence and absence of HcESPs (5µg/ml) for 1 h at 37°C.

Confirmation of binding was determined by an immunofluorescence assay (IFA) as described by Yuan et al [45]. Briefly, washed cells (10⁵ / ml) were fixed with 4% paraformaldehyde on a poly-L-lysine-coated glass slide. The cells were then treated with blocking solution (4% BSA in PBS) for 30 min to minimize background staining. Then cells were incubated with rat anti-HcESP IgG (1:100) as a primary antibody for 2 h. After being washed with PBS for 3 times (5 min each), the cells were incubated with second antibody (1:300) coupled to the fluorescent dye Cy3 (Beyotime, Jiangsu, China) for 1h at room temperature. The nuclear staining was performed with 2-(4-amidinophenyl)-6-indole carbamidinedihydrochloride (DAPI, 1.5 µM; Sigma, MO, USA) for 6 min. finally the protein localization was determined by checking the staining patterns with a 100× oil objective lens on a laser scanning confocal microscope (L SM710, Zeiss, Jena, Germany). Digital images were captured using the Zeiss microscope software package ZEN 2012 (Zeiss, Jena, Germany).

Detection of the cytokine levels by ELISA

Enzyme linked immunosorbent assay (ELISA) was used for the detection of cytokine levels. Briefly, the freshly isolated PBMCs were re-suspended to a final density of 5 × 10⁶ in complete medium (RPMI 1640 supplemented with 100 U/ml penicillin, 100 µg/ml streptomycin, 2 mM L-glutamine, 10% FCS). The cells were activated with ConA (10 µg/ml) and treated at the same time with a serial concentrations of HcESPs (5, 10, 20, and 40 µg/ml) and an equal volume of PBS as control. Then, the cells were seeded into 24-well plates (1ml/well) and cultured for 24h in 5% CO₂ atmosphere at 37 °C.

Next, the plates were then centrifuged at 200 × g for 15 min and the supernatants were collected. The levels of IL-4, IL-10, IL-17 and IFN-γ in supernatants were determined using commercially available goat ELISA kits (Jian chen, China). Three individual experiments were performed.

Cell migration assay

The cells were placed on the upper layer of a cell permeable membrane and a solution containing the test agent is placed below the cell permeable membrane. Following an incubation period (3-18 hours), the cells that have migrated through the membrane are stained and counted. The membrane is coated with some extracellular matrix component (e.g. collagen) which facilitates both adherence and migration.

The cell migration assay was performed using a Transwell system (Corning, USA), which allows cells to migrate throughout an 8 µm pore size polycarbonate membrane [27]. The treatment group was incubated with different concentrations of HcESPs (5, 10, 20, and 40µg/ml) and the control group was treated with an equal volume of PBS. The cells were placed on the upper chamber of cell permeable membrane and culture media was placed below the membrane. Following the incubation period of 12 hours, the cells migrated to lower chambers were collected and then the random migration was determined. The difference between the mean values was calculated using ANOVA. Each experiment was performed in triplicate.

Nitric oxide production assay

The goat PBMCs were harvested and washed twice with PBS. Then, 100 µl of cells (1 × 10⁶ cells/ml) were incubated either with PBS and a serial concentrations of HcESPs (5, 10, 20, and 40µg/ml) in 96-well plates in DMEM medium. Production of NO by PBMCs was determined by measurement of intracellular nitrite in the PBMC by using the Griess assay [46] according to the instructions of Total Nitric Oxide Assay Kit (Beyotime Biotechnology, China). Absorbance of the colored solution at 540 nm (OD540) in each well was measured using a plate reader (Bio-Rad Laboratories , USA). Absorbance values were converted to micromoles per liter (µmol/L) using a standard curve that was generated by addition of 0 to 80 µmol/L sodium nitrite to fresh culture media. Three individual experiments were performed.

Cell proliferation assay

Cell proliferation assay was performed as described previously [10]. Briefly, PBMCs (1 × 10⁶ cells/ml) were activated with ConA (10 µg/ml) and incubated at the

same time with a serial concentrations of HcESPs (5, 10, 20, and 40 μ g/ml) and an equal volume of PBS as control at 37°C in 5% CO₂ incubator for 72 h. CCK-8 solutions (Beyotime Biotechnology, China) were added to each well of the plates 4 h before harvesting the cells and the absorbance values at 450 nm (OD₄₅₀) were measured using a microplate reader (Thermo Scientific, USA). Cells exposed to ConA with control buffer served as controls and the OD₄₅₀ in controls were set as 100%. Cell proliferation index was calculated by the formula: OD₄₅₀ rHco-gal-m /OD₄₅₀ control. Each experiment was performed in triplicate.

CONFLICTS OF INTERESTS

The authors declare that they have no competing financial interests.

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