Deletion of the γ -secretase subunits *Aph1B/C* impairs memory and worsens the deficits of knock-in mice modeling the Alzheimer-like familial Danish dementia

Fabrizio Biundo¹, Keita Ishiwari^{1,2}, Dolores Del Prete¹ and Luciano D'Adamio¹

¹ Department of Microbiology and Immunology, Albert Einstein College of Medicine, Bronx, New York, United States of America

² Research Institute on Addictions University at Buffalo, Buffalo, New York, United States of America

Correspondence to: Luciano D'Adamio, email: Luciano.dadamio@einsein.yu.edu Keywords: APP, Itm2b-bri2, Alzheimer, Danish dementia, gamma-secretase, Gerotarget

Received: December 18, 2015 Accepted: February 08, 2016 Published: March 01, 2016

ABSTRACT

Mutations in *BR12/ITM2b* genes cause Familial British and Danish Dementias (FBD and FDD), which are pathogenically similar to Familial Alzheimer Disease (FAD). BR12 inhibits processing of Amyloid precursor protein (APP), a protein involved in FAD pathogenesis. Accumulation of a carboxyl-terminal APP metabolite $-\beta$ -CTF- causes memory deficits in a knock-in mouse model of FDD, called FDD_{k1}. We have investigated further the pathogenic function of β -CTF studying the effect of *Aph1B/C* deletion on FDD_{k1} mice. This strategy is based on the evidence that deletion of Aph1B/C proteins, which are components of the γ -secretase that cleaves β -CTF, results in stabilization of β -CTF and a reduction of A β . We found that both the FDD mutation and the Aph1B/C deficiency mildly interfered with spatial long term memory, spatial working/short-term memory and long-term contextual fear memory. In addition, the Aph1BC deficiency induced deficits in long-term cued fear memory. Moreover, the two mutations have additive adverse effects as they compromise the accuracy of spatial long-term memory and induce spatial memory retention deficits in young mice. Overall, the data are consistent with a role for β -CTF in the genesis of memory deficits.

INTRODUCTION

APP plays a central role in the pathogenesis of both sporadic and familial AD. Indeed, *APP* mutations that alter APP processing either protect from sporadic AD or cause familial AD; additionally, mutations in genes that regulate APP processing -such as *PSENs* and *BRI2/ITM2B*- cause FAD, FBD and FDD [1-12].

As briefly mentioned above, mutations in *BRI2/ ITM2B* cause the AD-like autosomal dominant FBD and FDD [5, 7]. FBD is characterized by the early onset of personality changes, memory and cognitive deficits, spastic rigidity, and ataxia [5]. FDD patients present early onset cataracts, deafness, progressive ataxia and dementia [7]. BRI2 is a type II membrane protein of 266 amino acids that is cleaved at the C terminus into a peptide of 23 amino acids (Bri23) plus a membranebound mature BRI2 (mBRI2) product [13, 14]. In FBD patients, a point mutation at the stop codon of *BRI2* results in a read-through of the 3'-untranslated region and the synthesis of a BRI2 molecule containing 11 extra amino acids at the COOH terminus. Cleavage by convertases generates a normal mBRI2 plus a longer peptide, the ABri peptide. FDD is caused by a10- nucleotide duplication before the stop codon of the BRI2 gene, which leads to the synthesis of a longer (277 amino acids) mutant protein [7, 15]. Convertase-mediated processing of the Danish mutant protein generates a longer C-terminal fragment, called ADan, and a normal mBRI2 polypeptide. Both ABri and ADan are deposited as amyloid fibrils. Of note, ADan deposits together with APP-derived Amyloid (Aβ42) peptides forming wide-spread amyloid angiopathy in the small blood vessels and capillaries of the cerebrum, choroid plexus, cerebellum, spinal cord, and retina [15]. Overall, FBD and FDD patients present cognitive dysfunctions and neuropathology including neurodegeneration, amyloid, and neurofibrillary tangles [7, 15-17], which are similar to those of Alzheimer's patients.

Knock-in mice models of FDD and FBD (FDD_{KI} and FBD_{KI} mice) showed that the mutant BRI2 proteins are mainly targeted for degradation, leading to a loss of mBRI2 function and, consequently, increased APP processing. Of note, loss on mBRI2 and increased APP processing was also detected in brain lysates from FDD and FBD patients. These alterations in APP processing, and not amyloid lesions, mediate memory and synaptic plasticity deficits caused by *BRI2/ITM2B* mutations. In fact, synaptic and memory deficits or FDD_{KI} mice were reduced by inhibition of β -cleavage of APP, which generates the fragments β -CTF and sAPP β [18], while they were worsened by inhibition of γ -secretase, which cleaves β -CTF into A β and AID/AICD [6, 19-27]

These findings suggest that increases in β -CTF can be neurotoxic and predicts that reducing γ -cleavage of APP has pathogenic consequences, while enhancing clearance of β -CTF is therapeutically advantageous. Several data support the first hypothesis: 1) loss of γ -secretase in the mouse brain induces neurodegeneration, memory and synaptic plasticity deficits; 2) Presenilins mutations associated with FAD cause a loss of γ -secretase function [28-36]; 3) sub-chronic administration of GSIs impairs normal cognitive function in APP transgenic mice [37]; 4) the GSI Semagacestat exacerbated cognitive deficits and impaired activities of daily living in human AD patients [38].

The evidence that reduction of γ -secretase activity impairs cognitive functions is consistent with a negative effect of β -CTF. Yet, γ -secretase cleaves other type I trans-membrane proteins, including the APP-like Protein 1 and -2, Neuregulin-1 and Notch [39-44] and cognitive functions deficits prompted by γ -secretase inhibition could be caused by decreased processing of any combination of γ -secretase substrates.

 γ -secretase is a multi-molecular complex comprising the catalytic subunits PSEN1 or PSEN2 and three accessory proteins: Anterior Pharynx-Defective 1 (Aph1), Nicastrin and Presenilin Enhancer Protein 2 (PEN2) [45-47]. Humans have two APH1 genes (APH1A and APH1B) [48, 49]; rodents have three because of a duplication of Aph1B that gave rise to the Aph1C gene [50, 51]. Aph1A-containing γ -secretase complexes are essential for Notch processing, while APP and Neuregulin-1 are preferred substrates of γ -secretase complexes containing either Aph1B or Aph1C; hence, Aph1BC^{-/-} mice show increased β -CTF but decreased A β peptide due to reduced γ -processing of β -CTF [50-53] and allow studying the consequence of inactivating γ -processing of APP limiting the confounding effects of inhibition of processing of other γ -secretase substrates.

Pharmacological evidence suggests a synaptictoxicity of β -CTF. Here, we have tested this hypothesis genetically. If increases in β -CTF prompt learning and memory deficits, *Aph1BC*^{-/-} mice may show deficits similar to those observed in FDD_{KI} mice; additionally, deletion of *Aph1BC* may worsen the defects of FDD_{KI} mice. On the contrary, if Danish mice develop learning and memory defects due to over-production of A β , the *Aph1BC*^{-/-} mutation will ameliorate learning and memory deficits of FDD_{KI} mice.

RESULTS

Young mice carrying the FDD mutation and deletion of Aph1B/C show mild learning and memory deficits

Mice were first tested at four months of age for anxiety-like behavior on the elevated rero maze. We analyzed the percentage of time spent in the open areas of the elevated zero maze during the 5-min testing period. While $\text{FDD}_{\text{KI}}/Aph1BC^{-/}$ mice spent more time in the open areas on average than mice of the other genotypes, oneway ANOVA revealed no significant effect of genotype, F(3, 60) = 1.94, p = 0.1331. Twelve animals (3 WT, 2 $\text{FDD}_{\text{KI}}, 4 \text{ FDD}_{\text{KI}}/Aph1BC^{-/}$, and 3 $Aph1BC^{-/}$) fell off the open areas of the maze during testing and were excluded from the data analysis. In addition, two mice (1 FDD_{KI} and 1 $\text{FDD}_{\text{KI}}/Aph1BC^{-/}$) were excluded due to technical problems with video tracking encountered during testing (Figure 1).

Next, mice were assayed for general locomotor activity levels and anxiety-like behavior in the open field. Four mice (1 FDD_{KI} and 2 FDD_{KI}/Aph1BC^{-/-}) were dropped from the statistical analysis due to persistent tracking errors caused by their light-colored fur. Since the video tracking system was not able to track these mice consistently, they were not included in the subsequent experiments. In addition, one FDD_{KI} mouse was excluded from the analysis due to a one-time tracking error during testing. Analysis of the mean distance traveled during the 10-min testing period by two-way ANOVA found a significant main effect for day, F(2, 138) = 82.23, p <0.0001, indicating habituation to the box over the threeday testing period, and a significant main effect for genotype, F(3, 69) = 3.46, p < 0.05, but no significant interaction between genotype and day, F(6, 138) = 1.15, p = 0.3387. Post-hoc comparisons (Dunnett's) showed that the mean distance traveled by FDD_{KI} animals were significantly greater than that traveled by WT mice on the first (p < 0.01) and second (p < 0.05) days (Figure 2A). Analysis of the time spent traveling at speed greater than 50 mm/s yielded significant main effects for day, F(2, 138) = 136.2, p < 0.0001, and for genotype, F(3, 69) = 3.97, p < 0.05, but no significant day \times genotype interaction, F(6, 138) = 0.95, p = 0.4583, and showed that FDD_{VI} mice spent more time traveling at speed greater than 50 mm/s than WT mice on all three days (p < 0.001, Day 1; p < 0.05, Days 2 and 3, Dunnett's) (Figure 2B). Analysis of the mean time spent in the center of the open field showed a significant main effect for day, F(2, 138) = 7.09, p < 0.01, and a significant day \times genotype interaction, F(6, 138) = 3.41, p < 0.01, while the main effect for genotype was close to significance, F(3, 69) = 2.47, p = 0.0692 (Figure 2C). FDD_{KI} mice spent more time in the arena center than WT mice on the first two days (p < 0.05, Dunnett's). Analysis of the number of entries into the arena center showed significant main effects for day, F(2, 138) = 37.67, p < 0.0001, and genotype, F(3, 69) = 4.20, p < 0.01, and a near significant interaction between genotype and day, F(6, 138) = 2.04, p = 0.0650. Post-hoc comparisons (Dunnett's) revealed that FDD_{KI} mice entered the arena center significantly more than did WT mice on the first (p < 0.001) and second (p < 0.05) days (Figure 2D). Overall these data indicate that $\ensuremath{\mathsf{FDD}_{\mathsf{KI}}}$ mice were generally more active and less anxious than WT, Aph1BC^{-/-} and FDD_{KI}/ Aph1BC^{-/-} mice, while no differences were detected between WT, *Aph1BC*^{-/-} and FDD_{K1}/*Aph1BC*^{-/-} animals.

Following the open field test, mice were tested in the MWM for spatial reference memory. One WT and one FDD_{KI} mouse were dropped from the experiments permanently due to severe bite wounds and persistent tracking errors caused by its light fur color, respectively. In addition, one $FDD_{KI}/Aph1BC^{-/-}$ mouse died before the task was completed. As shown in Figures 3A and 3B, the visible platform task conducted prior to the reference memory task revealed no significant differences among the four genotype groups in path length traveled, F(3,(66) = 1.43, p = 0.2427, or swim speed, F(3, 66) = 0.60,p = 0.6177, indicating that none of the mutant mice had any visual or motor deficits relative to WT control at this age. Figure 3C depicts the mean path length traveled by animals during the acquisition phase of the hidden platform task. Two-way ANOVA revealed a significant main effect for day on path length, F(5, 330) = 68.20, p < 0.0001, indicating animals' acquisition of reference memory for the platform location. There was also a significant interaction between day and genotype, F(15,(330) = 2.18, p < 0.01, while no significant main effect for genotype was found, F(3, 66) = 2.49, p = 0.0676. Tukey's comparisons revealed some differences among the genotypes on the first day, with Aph1BC^{-/-} mice traveling a significantly larger distance than WT mice (p < 0.05) and FDD_{KI} mice (p<0.001), and with $FDD_{KI}/Aph1BC^{-}$ mice than FDD_{KI} mice (p < 0.01). On the probe trial conducted two days after the last acquisition session, the analysis of the percentage of time spent in the four quadrants revealed a significant main effect for quadrant, F(3, 198) = 50.71, p < 0.0001, but no significant main effect for genotype, F(3, $(66) = 0.55, p = 0.6469, \text{ or significant quadrant} \times \text{genotype}$ interaction, F(9, 198) = 1.15, p = 0.3293 (Figure 3D).

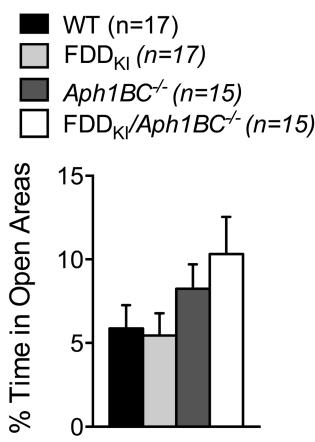


Figure 1: Elevated Zero Maze test on mice at 4 months of age. Data are expressed as means \pm S.E.M. No significant effect of genotype was found.

While the percentage of time spent in the target quadrant is the most popular measure of probe trial performance [54], counting the number of times the animal crosses a small area surrounding the former platform position (counter crossings) provides more information on the spatial accuracy with which the exact location of the platform has been encoded [55]. One-way ANOVA did not reveal a significant effect of genotype on the number of counter crossings in the target quadrant, F(3, 66) = 2.00, p= 0.1233 (Figure 3E). However, a separate unpaired t-test showed a significant difference between WT and FDD_{KI} Aph1BC^{-/-} mice (p = 0.0177, Figure 3F). There were no significant differences in the average proximity to the original platform location among the genotype groups, F(3, 66) = 0.75, p = 0.5282 (Figure 3G). Thus, at 4 months of age, when present together the FDD mutation and the Aph1BC deficiency mildly interfered with spatial longterm memory by compromising its accuracy.

In the reversal learning task, in which the location of the hidden platform was moved to the quadrant opposite to

the original target quadrant, all the genotypes performed the task in a similar manner during the acquisition phase (Figure 3H), with ANOVA showing a significant main effect for day, F(5, 330) = 102.5, p < 0.0001, but no significant main effect for genotype, F(3, 66) = 0.70, p =0.5577, or day \times genotype interaction, F(15, 330) = 0.71, p = 0.7770. On the probe trial given two days after the last reversal learning session, no differences were found among the genotypes in the percentage of time spent in the quadrants (Figure 3I), with a significant quadrant main effect, F(3, 198) = 140.2, p < 0.0001, but no significant main effect for genotype, F(3, 66) = 0.38, p = 0.7681, or quadrant \times genotype interaction, F(9, 198) = 0.55, p = 0.8376. Similarly, no differences were found among the genotypes in the number of counter crossings in the target quadrant (Figure 3J), F(3, 66) = 0.95, p = 0.4230, or the average proximity to the target (Figure 3K), F(3, 66) =0.47, p = 0.7029.

The same cohorts of mice were again assessed for spatial reference memory in the Morris water maze

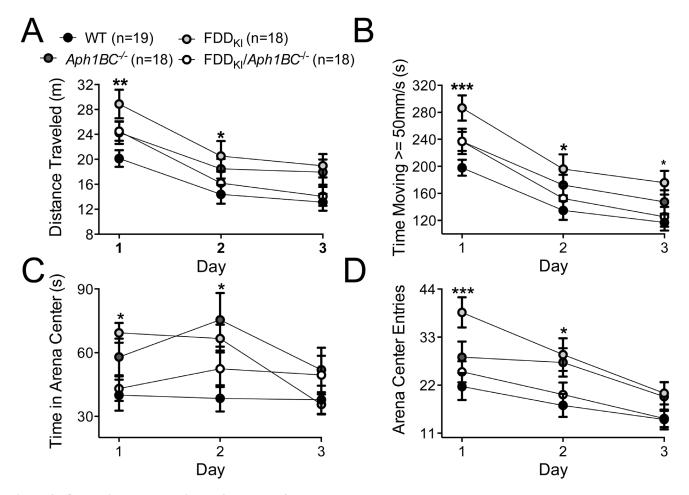


Figure 2: Open Field test on mice at 4 months of age. Data are expressed as means (\pm S.E.M.) during the 10-min testing period over 3 days. **A.** Total distance traveled. **B.** Amount of time in which the animal ambulated at speed greater than 50 mm/s. FDD_{K1} mice were more active than WT mice, especially on Day 1. **C.** Amount of time the animal spent in the center of the arena (20 cm × 20 cm). **D.** Total number of entries into the arena center. FDD_{K1} mice entered and spent more time in the arena center on Days 1 and 2 than WT mice. * p < 0.05, ** p < 0.01, *** p < 0.001, WT vs. FDD_{K1}.

at 7-8 months of age. To determine whether genetic manipulations affect learning and memory in an aging dependent manner, it is customary to use different cohorts of animal to perform a task (such as the Morris water maze) at different ages. In this manner, the results are not influenced by learning and/or habituation to the tasks that may occur when the same cohort of mice are subjected to the same task more than once. However, a longitudinal analysis is ethically and economically preferable since it reduces the number of experimental animals. In addition, it eliminates possible confounding effects due to genetic variability between different cohorts of mice. Finally, testing if mice forget, during aging, tasks learned as young adults, mirrors what happens in AD patients. In this new Morris water maze test, the number of daily trials was reduced to three to alter the difficulty of the task. One WT mouse was dropped since it had become too weak to swim. The visible platform task revealed no significant differences among the genotypes in path length traveled, F(3, 65) = 1.36, p = 0.2638 (Figure 4A), or swim speed, F(3, 65) = 0.27, p = 0.8488 (Figure 4B). In the reference memory task, as shown in Figure 4C, no significant differences among the genotypes were found during acquisition, with two-way ANOVA showing a significant main effect for day, F(4, 260) = 12.41, p < 0.0001, but no significant main effect for genotype, F(3, 65) = 0.81, p =0.4932, or day \times genotype interaction, F(12, 260) = 0.95, p = 0.4957. The probe trial conducted two days after the last acquisition session did not reveal any differences among the genotypes in the percentage of time spent in the four quadrants (Figure 4D), with a significant quadrant main effect, F(3, 195) = 115.4, p < 0.0001, but no significant main effect for genotype, F(3, 65) = 1.18, p = 0.3257, or quadrant \times genotype interaction, F(9, 195) = 1.25, p = 0.2670. There was no significant effect of genotype on the number of counter crossings in the target quadrant, F(3, 65) = 1.67, p = 0.1828 (Figure 4E), or the average proximity to the target, F(3, 65) = 2.18, p = 0.0990 (Figure 4F). Thus, no significant differences were found among the genotypes in the acquisition or retention of spatial reference memory at 7-8 months of age, and the small deficits in counter crossing observed in 4 month-old $FDD_{y_1}/Aph1BC^{-/-}$ mice was not seen again at 7-8 months of age, possibly due to learning/habituation to the task. Following the completion of the probe trial at 7-8 months, mice were subjected to a working memory task in

rollowing the completion of the probe trial at 7-8 months, mice were subjected to a working memory task in the eight-arm radial arm water maze. Mice were initially given four consecutive daily acquisition trials with a 15 seconds inter-trial interval. However, it became apparent after six days of training with this procedure that most mice lacked the physical strength required for maintaining their performance in the maze during the four consecutive acquisition trials, and that their performance was not improving after six days (data not shown). Accordingly, the procedure was modified such that mice would be given a 6-min inter-trial interval in the holding cage, and that three, instead of four, acquisition trials would be given before the retention trial. After a break on the seventh day, mice were tested for five more days with this new procedure and the results pertaining to these last five days are shown in Figure 5. Repeated measures oneway ANOVA (RM one-way ANOVA) found a significant main effect for trial in WT [F (2.065, 33.04) = 18.37, p < 18.37(0.0001)], FDD_{K1} [F (2.320, 37.12) = 19.88, p < 0.0001)] and $Aph1BC^{-1}$ [F (2.312, 39.31) = 22.14, p < 0.0001)], but not in $FDD_{yy}/Aph1BC^{-2}$ mice [F (2.033, 28.46) = 3.218, p=0.0542)]. A post hoc multiple comparison of the mean of each trial to the mean of every other trial (Tukey's multiple comparisons test) indicated that performance by WT and FDD_{KI} mice improved significantly between Trials 1 and 2 (p < 0.05 for WT and p < 0.01 for FDD_{K1}), Trials 1 and 3 (p < 0.001 for WT and p < 0.01 for FDD_{v1}) and Trials 1 and R (p < 0.0001 for both genotypes). The performance of Aph1BC^{-/-} mice improved significantly between Trials 1 and 3, 1 and R (p < 0.0001) as well as 2 and 3 (p < 0.05), and 2 and R (p < 0.01). On the other hand, this post hoc multiple comparison analysis was not performed for the FDD_{KI}/Aph1BC--- mice since the RM one-way ANOVA analysis did not find a significant main effect for trial in these mice. Overall, these observations indicated that the magnitude of improvement in performance was the smallest in FDD_{KI}/Aph1BC-/mice, compared to WT, FDD_{KI}, or Aph1BC^{-/-} mice. This difference may reflect working memory impairment in $FDD_{KI}/Aph1BC^{-}$ mice, or might have resulted from the fact that FDD_{KI}/Aph1BC^{-/-} mice actually made fewer errors than mice of the other genotypes on the first trial, thereby making the gain smaller.

The FDD and *Aph1BC*^{-/-} mutations induce mild deficits in spatial long-term memory in middle age

At 12 months of age, another reference memory task was given. For this task, the number of daily trials was further reduced to two per day, and the length of the probe trial was reduced to 30 s. In addition, the visible platform task was given after the completion of the probe trials. One WT mouse that displayed extensive and persistent thigmotaxis during the reference memory task was excluded from the data analysis. Also, one Aph1BC^{-/-} mouse was eliminated because it had developed rectal prolapse. Again, the visible platform task did not find any differences among the genotypes in path length traveled, F(3, 63) = 0.81, p = 0.4911 (Figure 6A), or swim speed, F(3, 63) = 0.81, p = 0.4908 (Figure 6B). In the reference memory task, mice of all the genotype learned the location of the hidden platform in a similar manner (Figure 6C). ANOVA found a significant main effect for day, F(7, 441) = 11.84, p < 0.0001, but no significant genotype main effect, F(3, 63) = 0.10, p = 0.9576, or day × genotype interaction, F(21, 441) = 0.75, p = 0.7773. On the probe

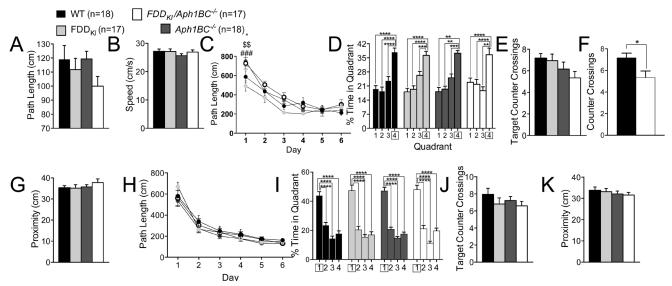


Figure 3: The Morris Water Maze task shows a mild deficit in accuracy of spatial long-term memory 4 month-old FDD_{KI}/*Aph1BC*^{-/-} mice. Data are expressed as means ± S.E.M. A.-B. Performance on the visible platform task. The visible platform task indicates that there were no significant differences among the genotypes in path length traveled (A) or swim speed (B). C. Acquisition of spatial reference memory in the hidden platform task. Mean path lengths across 6 daily trials are shown. Significant differences among the genotypes were found only on the first day: * p < 0.05, ** p < 0.01, *** p < 0.001. D.-F. Performance on the 60-s probe trial given 2 days after the last acquisition session. (D-F) Performance on the 60-s probe trial given 2 days after the last acquisition session. (D) Percentage of time spent in the four quadrants. ** p < 0.01, *** p < 0.0001 (E) Number of counter crossings in the target quadrant (Quadrant 4). * p < 0.05 (F) A separate unpaired t-test showed a significant difference between WT and FDD_{KI}/*Aph1BC*^{-/-} mice (* p = 0.0177). G. Average proximity to the former platform location. H. Performance in the reversal learning task with a new platform location. Mean path length across 6 daily trials are shown. I.-K. Performance on the 60-s probe trial given 2 days after the last reversal learning session. (I) Percentage of time spent in the four quadrants. **** p < 0.001 (J) Number of counter crossings in the target quadrant (Quadrant 1). (K) Average proximity to the former platform location.

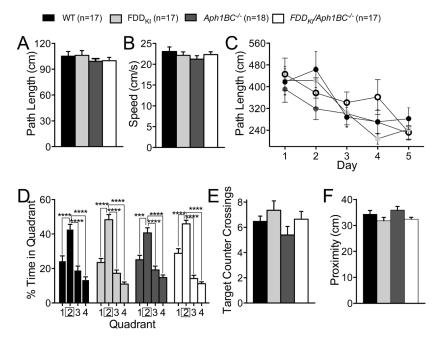


Figure 4: Normal spatial memory in 7-8 month-old mice. Data are expressed as means \pm S.E.M. **A.-B.** Performance on the visible platform task. There were no significant differences among the genotypes in path length traveled (A) or swim speed (B). **C.** Acquisition of spatial reference memory in the hidden platform task. Mean path lengths across 3 daily trials are shown. No significant differences were found among the genotypes. **D.-F.** Performance on the 60-s probe trial given 2 days after acquisition. (D) Percentage of time spent in the four quadrants. *** p < 0.001, **** p < 0.0001. (E) Number of counter crossings in the target quadrant (Quadrant 2). (F) Average proximity to the former platform location. No significant differences were found among the genotypes in any of the measures.

trial given two days after the last acquisition session, no significant differences were found among the genotypes in the analysis of the percentage of time spent in the quadrants (Figure 6D), with a significant main effect for quadrant, F(3, 189) = 35.30, p < 0.0001, but no significant genotype main effect, F(3, 63) = 1.21, p = 0.3123, or quadrant × genotype interaction, F(9, 189) = 0.65, p = 0.7571. However, the analysis of the number of counter

crossings in the target quadrant revealed a significant effect of genotype, F(3, 63) = 5.80, p < 0.01 (Figure 6E). *Post-hoc* comparisons (uncorrected Fisher's LSD) revealed that WT mice crossed the counter in the target quadrant significantly more often than FDD_{K1} mice (p < 0.001), *Aph1BC^{-/-}* (p < 0.01) and FDD_{K1}/*Aph1BC^{-/-}* mice (p < 0.05). There was no significant effect of genotype on the average proximity to the former platform location, F(3,

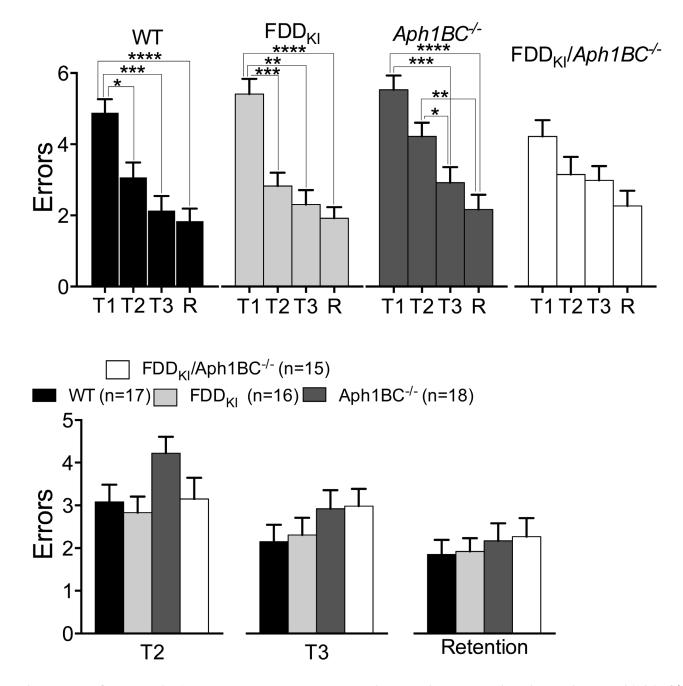


Figure 5: The 8-arm Radial Arm Water Maze task detects possible working memory impairment in FDD_{KI}/Aph1BC^{-/-} mice at 7-8 months of age. Data are expressed as means \pm S.E.M. A. Number of errors on 3 daily acquisition trials averaged across the last 3 test days. The performance by WT, FDD_{KI}, and Aph1BC^{-/-} mice improved across trials to a greater degree, compared to FDD_{KI}/Aph1BC^{-/-} mice. *p < 0.05, **p < 0.01, ****p < 0.001. B. Number of errors on trials 2, 3 and retention (R) given 30 min after the third acquisition trial. No significant differences were found among the genotypes.

63) = 0.79, p = 0.5065 (Figure 6F). Thus, at 12 months of age, the FDD mutation and the *Aph1BC* deficiency both mildly interfered with spatial long-term memory by compromising its accuracy.

Mice were tested for spatial working memory in Morris water maze at 15 months of age. As shown in Figures 7A and 7B, the visible platform task given at this age found no significant effect of genotype on path length traveled, F(3, 64) = 0.37, p = 0.7743, or swim speed, F(3, 64) = 0.66, p = 0.5797. In the working memory task, in which the location of the hidden platform was changed daily, we measured the path-length to target (Figure 7C). RM one-way ANOVA found a significant main effect for trial in WT [F (1.984, 29.76) = 5.004, p =0.0136]], but not in FDD_{KI} [F (1.948, 31.17) = 1.822, p =0.1793)], $Aph1BC^{-/-}$ [F (1.943, 31.09) = 2.749, p = 0.0809] and FDD_{KI}/ $Aph1BC^{-/-}$ mice [F (1.655, 26.48) = 2.180, p=0.14)]. A post hoc multiple comparison of the mean of each trial to the mean of every other trial (Tukey's multiple comparisons test) indicated that only the performance of WT mice improved significantly between Trials 1 and 3 (p < 0.05). This test suggests that, at 15 months of age, the FDD mutation and the *Aph1BC* deficiency both mildly interfered with working memory.

Learning and memory deficits are caused by the Aph1BC deficiency and the FDD mutation at old age (18-19 months)

At 18-19 months of age, mice were given a series of tests. The first test, conducted at 18-19 months of age, evaluated mice in the two-trial Y-maze test task for spatial recognition memory. One $\text{FDD}_{\text{KI}}/Aph1BC^{/-}$ mouse had died before reaching this age. Figure 8A depicts the mean

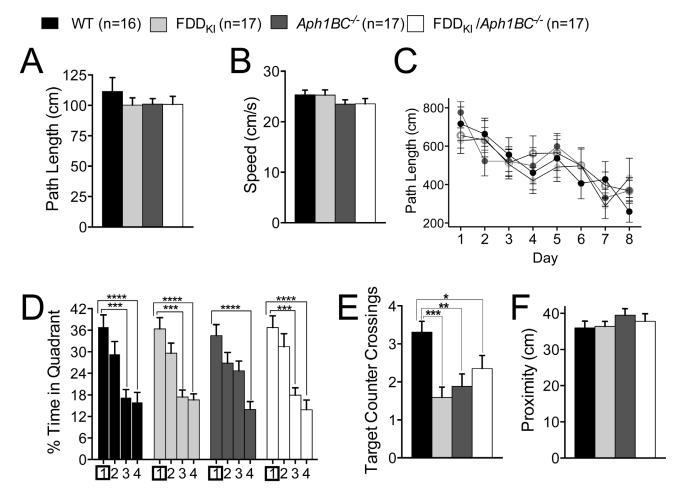
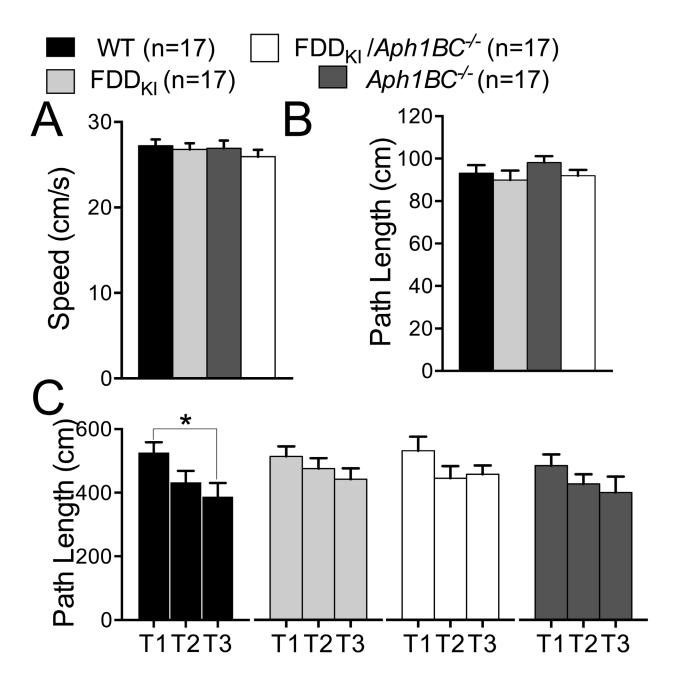
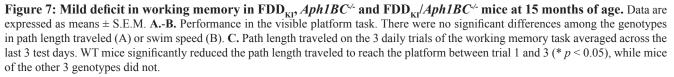


Figure 6: Mild deficit in accuracy of spatial long-term memory in 15 month-old FDD_{KI}, *Aph1BC*^{-/-} and FDD_{KI}/*Aph1BC*^{-/-} mice. Data are expressed as means \pm S.E.M. A.-B. Performance on the visible platform task. There were no significant differences among the genotypes in path length traveled (A) or swim speed (B). C. Acquisition of spatial reference memory in the hidden platform task. Mean path lengths across 2 daily trials are shown. No significant differences were found among the genotypes. D.-F. Performance on the 30-s probe trial given 2 days after acquisition. D. Percentage of time spent in the four quadrants. *** p < 0.001, **** p < 0.0001. E. Number of counter crossings in the target quadrant (Quadrant 1). WT mice crossed the target counter significantly more than FDD_{KI} or *Aph1BC*^{-/-} mice. *** p < 0.01, WT *vs*. FDD_{KI}, ** p < 0.01, WT *vs*. Aph1BC^{-/-}; * p < 0.05, WT *vs*. FDD_{KI}/Aph1BC^{-/-} F. Average proximity to the former platform location.

number of arm entries during the 5-min test trial, which is an index for animals' total activity levels. ANOVA found a small but significant effect of genotype, F(3, 63) = 2.77, p = 0.0490 < 0.05, and Fisher's LSD comparison test showed that FDD_{KI} mice were more mobile than WT and $FDD_{KI}/Aph1BC^{J-}$ mice (p < 0.05). Fisher's test was used here because, despite the significant overall genotype effect, no significant differences among the genotypes were detected by corrected comparison tests. Given the significant genotype effect on the number of total arm entries, the percentage of entries into each arm, instead of raw numbers of arm entries, was used to analyze animals' preference for the novel arm vs. the known arm. As shown in Figure 8B, the analysis of the percentage of entries into the novel and known arms found a significant main effect for arm, F(1, 63) = 43.34, p < 0.0001, but no significant genotype main effect, F(3, 63) = 0.78, p = 0.5076, or arm × genotype interaction, F(3, 63) = 1.96, p = 0.1295. Sidak's





multiple comparisons between the arms revealed that the novel arm was entered significantly more than the known arm by WT mice and FDD_{KI} mice, but not by FDD_{KI} Aph1BC^{-/-} mice or Aph1BC^{-/-} mice, and that the level of statistical significance was much higher for WT mice (p <0.0001) than for FDD_{KI} mice (p < 0.01). As can be seen in Figure 8C, the analysis of the time spent in the novel and known arms found a significant main effect for arm, F(1,(63) = 29.63, p < 0.0001, as well as a significant interaction between arm and genotype, F(3, 63) = 2.82, p < 0.05, while showing no significant main effect for genotype, F(3, 63) = 0.43, p = 0.7345. Post-hoc comparisons across the genotypes (Dunnett's) revealed that WT mice spent significantly more time in the novel arm than did FDD_{yy} Aph1BC^{-/-} mice (p < 0.05). In addition, comparisons between the arms (Sidak's) showed that WT mice spent significantly more time in the novel arm than in the known arm (p < 0.0001). FDD_{KI} mice also spent significantly more time in the novel arm than in the known arm, but to a much smaller degree (p < 0.05) than WT mice. By contrast, the difference in time spent between the novel and known arms were not statistically significant for FDD_{KI}/Aph1BC^{-/-} or Aph1BC^{-/-} mice. In sum, the results of the two-trial Y-maze task demonstrated that WT mice and, to a lesser extent, $\mathrm{FDD}_{\mathrm{KI}}$ mice distinguished between the novel and familiar arms better than FDD_{KI}/Aph1BC^{-/-} or Aph1BC^{-/-} mice after a 1-h retention interval, indicating that the Aph1BC deficiency, and to a lesser extent the FDD mutation, leads to impairment of short-term spatial recognition memory.

Next we analyzed the general locomotor activity levels in the open field at 18-19 months. There was a main effect for day in the total distance traveled, F(2,126 = 85.01, p < 0.0001, but no significant main effect for genotype, F(3, 63) = 2.03, p = 0.1186, or interaction between day and genotype, F(6, 126) = 1.83, p = 0.0977(Figure 9A). There was a main effect for day in the amount of time in which the animal ambulated at speed greater than 50 mm/s, F(2, 126) = 94.25, p < 0.0001, but no significant genotype main effect, F(3, 63) = 2.24, p = 0.0919, or day \times genotype interaction, F(6, 126) = 1.62, p = 0.1466 (Figure 9B). We detected a significant main effect for day, F(2, 126) = 10.66, p < 0.0001, but no significant main effect for genotype, F(3, 63) = 0.09, p =0.9667, or day \times genotype interaction, F(6, 126) = 0.66, p = 0.6770 in the mean time spent in the center of the arena (Figure 9C). The total number of entries into the arena center showed significant main effect for day, F(2, 126) =35.27, p < 0.0001, but no significant genotype main effect, F(3, 63) = 1.51, p = 0.2212, or day \times genotype interaction, F(6, 126) = 0.45, p = 0.8419 (Figure 9D).

Following the open field test, the spontaneous alternation test was conducted in the Y-maze to assess animals' spatial working memory. Analysis of the total number of arm entries during the 5-min testing period showed no significant effect of genotype was detected, F(3, 62) = 0.36, p = 0.7811 (Figure 10A). Since the minimum number of arm entries needed for a complete alternation is three, six mice that had less than three arm entries in total (2 WT, 1 FDD_{K1}, 1 FDD_{K1}/*Aph1BC*^{-/-}, and

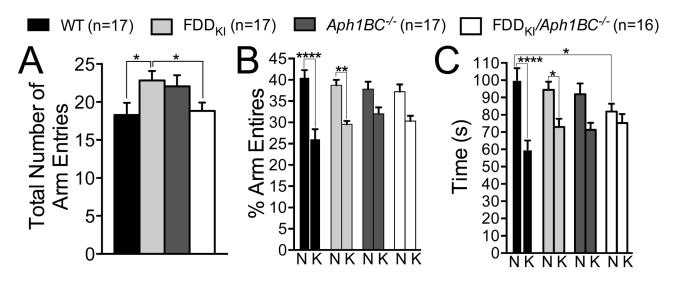


Figure 8: The two-trial Y-maze test showed mild deficit of short-term spatial recognition memory in FDD_{Kl} , $Aph1BC^{-/-}$ and $FDD_{Kl}/Aph1BC^{-/-}$ mice at 18-19 months of age. Data are expressed as means ± S.E.M. A. Total number of arm entries. FDD_{Kl} mice made significantly more arm entries than WT or $FDD_{K}/Aph1BC^{-/-}$ mice (*p < 0.05). B. Percentage of entries into the novel (N) and known (K) arms. WT and, to a smaller degree, FDD_{Kl} mice entered the novel arm significantly more than the known arm, while $FDD_{Kl}/Aph1BC^{-/-}$ or $Aph1BC^{-/-}$ mice did not (*p < 0.01; **** p < 0.0001). C. Time spent in the novel and known arms. WT mice and, to a smaller degree, FDD_{Kl} mice spent significantly more time in the novel arm than in the known arm, while $FDD_{Kl}/Aph1BC^{-/-}$ or $Aph1BC^{-/-}$ or $Aph1BC^{-/-}$ mice did not (*p < 0.05; **** p < 0.0001). In addition, there was a significant difference in time spent in the novel arm between WT and $FDD_{Kl}/Aph1BC^{-/-}$ (*p < 0.05).

2 *Aph1BC*^{-/-}) were not included in the analysis of the percentage of alternations, which showed no significant effect of genotype, F(3, 57) = 1.26, p = 0.2969 (Figure 10B).

Next, we used the elevated zero maze to test for anxiety-like behavior. As shown in Figure 11, while $Aph1BC^{-/-}$ mice spent more time in open areas than mice of the other genotypes on average, the genotype effect did not reach statistical significance, F(3, 60) = 1.396, p = 0.2527. Three animals (2 FDD_{KI}/Aph1BC^{-/-} and 1 Aph1BC^{-/-}) fell off the open areas of the maze during testing and were excluded from the data analysis.

Mice were then evaluated for spatial working memory in the 6-arm radial arm water maze. RM oneway ANOVA found a significant main effect for trial in WT [F (2.025, 32.40) = 28.12, p < 0.0001)], FDD_{KI} [F (2.082, 33.32) = 13.32, p < 0.0001)], $Aph1BC^{-/-}$ [F (2.437, 38.99) = 4.542, p = 0.012)] and FDD_{KI}/Aph1BC^{-/-} mice [F (2.738, 41.07) = 4.680, p=0.0081)] (Figure 12A). A *post hoc* multiple comparison of the mean of each trial to the mean of every other trial (Tukey's multiple comparisons) test) indicated that performance by WT mice improved significantly between Trials 1 and 2 (p < 0.01), 1 and 3 (p < 0.001), 1 and R (p < 0.0001) as well as between Trials 2 and R (p < 0.01). The FDD_{KI} mice improved between Trials 1 and 3 and 1 and R as well as between Trials 2 and 3 and 2 and R (p < 0.01). As for FDD_{KI}/*Aph1BC*^{-/-} and *Aph1BC*^{-/-} mice, their performance improved significantly between Trials 1 and R (p < 0.05). However, it is worth noting that *Aph1BC*^{-/-} mice actually made fewer errors than mice of the other genotypes on the first trial, thereby making the gain smaller. On the other hand, the performance by FDD_{KI}/*Aph1BC*^{-/-} mice significantly improved between Trials 1 and 2,1 and R, but the degree of significance was much smaller (p < 0.05) as compared to WT and FDD_{KI} mice.

We also compared the number of errors made by mice of the four genotypes at Trials 2, 3 and R (Figure 12B). Ordinary one-way ANOVA found significant effect of genotype at Trials 3 [F (3, 63) = 2.751, p = 0.05)] and R [F (3, 63) = 3.616, p = 0.00178)], but not 2 [F (3, 63) = 0.9047, p = 0.4439)]. A *post hoc* multiple comparison

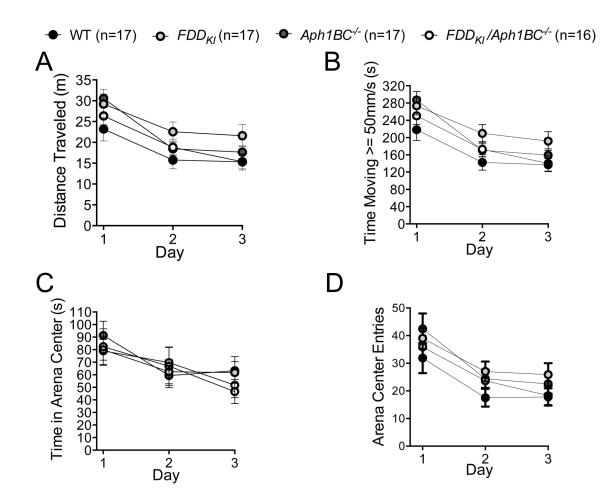
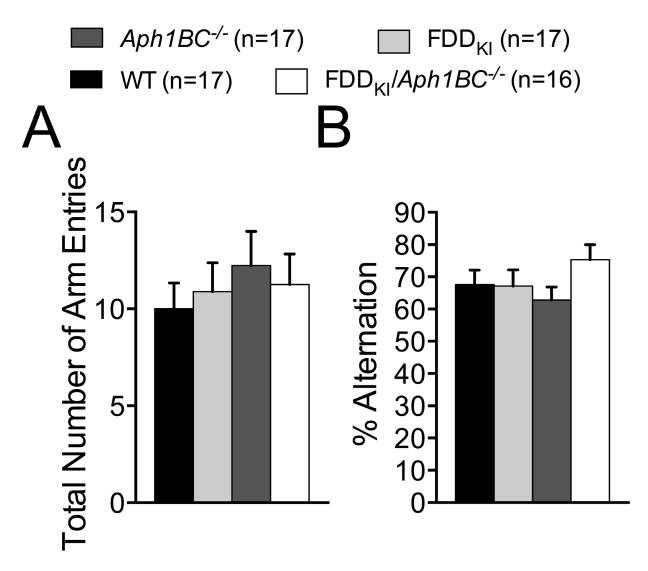


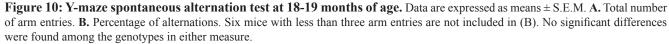
Figure 9: Open Field test on mice at 18-19 months of age. Data are expressed as means (\pm S.E.M.) during the 10-min testing period over 3 days. **A.** Total distance traveled. **B.** Amount of time in which the animal ambulated at speed greater than 50 mm/s. **C.** Amount of time the animal spent in the center of the arena (20 cm \times 20 cm). **D.** Total number of entries into the arena center. No significant differences were found among the genotypes in any of the measures.

of the mean of each genotype to the mean of every other genotype (Tukey's multiple comparisons test) indicated that WT mice committed significantly fewer errors than $FDD_{KI}/Aph1BC^{-/-}$ mice both at Trials 3 (p < 0.05) and R (p < 0.01). The differences between WT vs. FDD_{KP} WT vs. Aph1BC^{-/-}, FDD_{KI}/Aph1BC^{-/-} vs. FDD_{KI} and FDD_{KI}/ Aph1BC^{-/-} vs. Aph1BC^{-/-} were not statistically significant. Altogether, these observations indicated that the degree of improvement in performance across trials during acquisition and retention was the greatest for WT mice, followed by FDD_{KI}, Aph1BC^{-/-} and FDD_{KI}/Aph1BC^{-/-} mice in the order named. These results further indicate that the FDD mutation and deletion of Aph1BC induces spatial working memory deficits. These data are also in accordance with the results of a previous study showing that the Aph1BC^{-/-} mutation induces spatial working memory deficits in mice of the F2 generation of the C57BL/6J–129/Ola hybrids [53].

After the radial arm water maze test, mice were assessed for possible visual or motor deficits in the visible platform task. Ten animals (2 WT, 1 FDD_{KI}, 5 FDD_{KI}' *Aph1BC*^{-/-}, and 2 *Aph1BC*^{-/-}) were mistakenly sacrificed prematurely after the radial arm water maze and before the visual platform task. As shown in Figures 12C and 12D, the visible platform task did not reveal any visual or motor deficits at this age. There was no significant effect of genotype on path length traveled, F(3, 53) = 1.13, *p* = 0.3467, or swim speed, F(3, 53) = 2.19, *p* = 0.0999.

Finally, mice were tested for contextual and cued fear memory using the fear conditioning paradigm. As is indicated in Figures 13A and 13B, in the test for contextual fear memory conducted 24 h after conditioning,





WT mice exhibited freezing behavior significantly more than mice in the other genotype groups. ANOVA revealed a significant effect of genotype on the mean percentage of freezing during the last 3 min of the test session, F(3,(53) = 5.45, p < 0.01, and post-hoc comparisons (Tukey's)indicated that WT mice froze significantly more than FDD_{KI} (p < 0.01), $Aph1BC^{-}$ (p < 0.01), and FDD_{KI} AphIBC^{-/-} (p < 0.05) mice (Figure 13A). As can be seen in Figure 13B, the analysis of the time course of the percentage of freezing in 1-min bins during the contextual test also showed a significant main effect for genotype, F(3, 53) = 4.75, p < 0.01, in addition to a significant main effect for time, F(4, 212) = 20.55, p < 0.0001, while finding no significant time \times genotype interaction, F(12, (212) = 0.89, p = 0.5557. Tukey's multiple comparison test further indicated that WT mice froze significantly more than FDD_{KI} mice in the last three minutes (p < 0.01 for the fourth min; p < 0.05 for the third and fifth min), more than Aph1BC^{-/-} mice in the first minute (p < 0.05) and the last three minutes (p < 0.01 for the third and fourth min; p < 0.05 for the fifth min), and more than FDD_{v1}/ *Aph1BC*^{-/-} mice in the third minute (p < 0.05). In the test for cued fear memory conducted 24 h after the contextual test, no significant effect of genotype was found on the percentage of freezing during the presentation of the tone CS, F(3, 53) = 2.27, p = 0.0910 (Figure 13C). However, as can be seen in Figure 13D, which shows the time course of freezing during tone presentation, while WT mice remained frozen throughout the 3-min presentation of the tone, FDD_{KI} mice, which were initially as frozen as WT mice, became more mobile towards the end of this period,

and FDD_{K1}/Aph1BC^{-/-} and Aph1BC^{-/-} mice froze less than WT mice for the entire period. Yet, the analysis of the time course showed a significant main effect for time, F(2,106) = 6.97, p < 0.01, but no significant main effect for genotype, F(3, 53) = 2.27, p = 0.0910, or time \times genotype interaction, F(6, 106) = 2.09, p = 0.0603. Nevertheless, the time \times genotype interaction approached significance, and when one-way ANOVA was performed on the percentage of freezing during the first, second, and third 1-min time periods separately, while no significant genotype effect was detected in the first minute, F(3, 53) = 2.47, p = 0.0720, or the second minute, F(3, 53) = 1.34, p =0.2712, there was a significant genotype effect in the third minute, F(3, 53) = 3.25, p < 0.05. Dunnett's comparison test further revealed a significant difference between WT and $\text{FDD}_{\text{KI}}/Aph1BC^{-/-}$ mice (p < 0.05), as well as between WT and *Aph1BC*^{-/-} mice (p < 0.05), in the last minute of tone presentation. Lastly, a day after the completion of the test for cued fear memory, all the experimental mice were evaluated for shock sensitivity. As shown in Figure 13E, mice of all the genotypes reacted to electric shock at the four intensity levels in a similar manner, with ANOVA showing a significant main effect for shock level, F(3,(159) = 97.70, p < 0.0001, but no significant main effect for genotype, F(3, 53) = 1.60, p = 0.2010, or shock level × genotype interaction, F(159, 159) = 1.45, p = 0.1709. While the amygdala is required and sufficient for fear conditioning to discrete sensory cues such as a tone, rapid formation of conditioned fear responses to environmental context is additionally mediated by the hippocampus [56-59]. Thus, our results indicate that both the FDD and

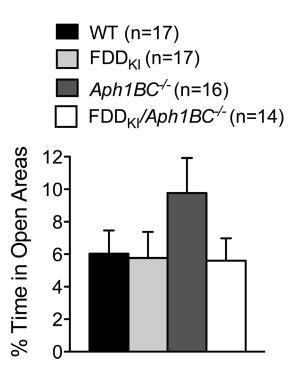


Figure 11: Elevated Zero maze test on mice at 18/-9 months of age. Mean (\pm S.E.M.) percentage of time spent in the open areas of the elevated zero maze. There were no significant differences among the genotypes.

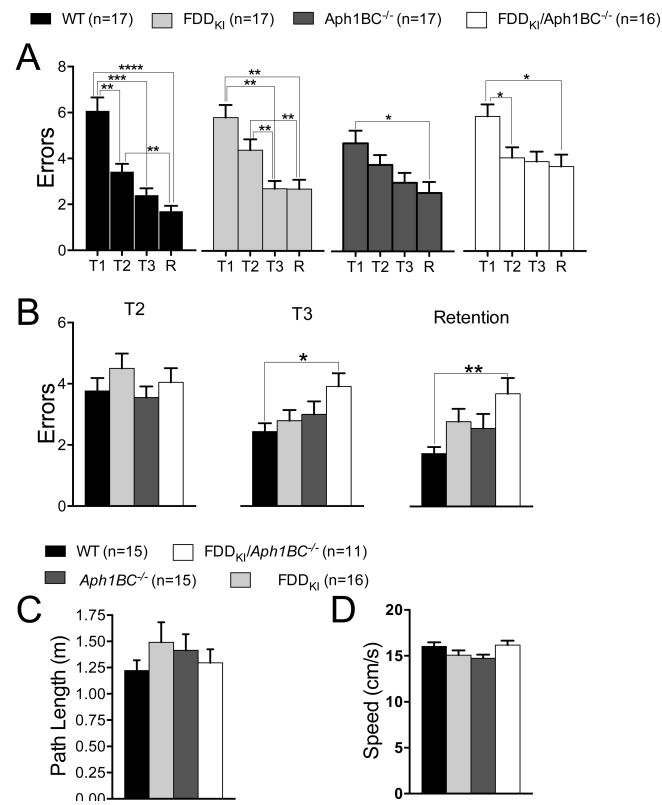


Figure 12: Mild spatial working memory deficits in 18-19 month-old FDD_{Kl} , *Aph1BC*^{-/-} and FDD_{Kl} /*Aph1BC*^{-/-} mice. Data are expressed as means ± S.E.M. (A) Number of errors on 3 daily acquisition trials averaged across days 6-9 of testing. (B) Number of errors on trials 2, 3 and retention (R) given 30 min after the third acquisition trial. The visible platform task showed no significant differences among the genotypes in path length traveled (C) or swim speed (D). *p < 0.05, **p < 0.01, ***p < 0.001, **** p < 0.0001.

Aph1BC^{-/-} mutations interfere with mnemonic information processing at the level of the hippocampus for the formation of long-term contextual fear memory at this age, while the *Aph1BC*^{-/-} mutation additionally has adverse effects on fear memory formation in the amygdala.

DISCUSSION

The clinical symptoms of AD include progressive loss of memory, thinking and language skills, as well as other behavioral changes. Short-term memory is the first to fail in AD patients. These clinical symptoms are accompanied by neuronal degeneration. Although it is widely believed that the disease is precipitated by the insurgence of brain lesions, such as A β amyloid plaques and neurofibrillary tau tangles, whether these alterations of tau and A β metabolism cause the debilitating clinical symptoms of AD and FDD is still unclear.

Because of the uncertainty concerning the pathogenic biochemical mechanisms of AD and related dementias, and considering that to improve the quality of life of AD patients -and caregivers- we need to either reverse or slow down the progression of the clinical symptomatology, we focused our analysis on learning and memory in our animal model of disease. In this study, the same cohort of mice of WT, FDD_{KI} , $FDD_{KI}/Aph1BC^{-/-}$, and $Aph1BC^{-/-}$ mice was assessed longitudinally for possible

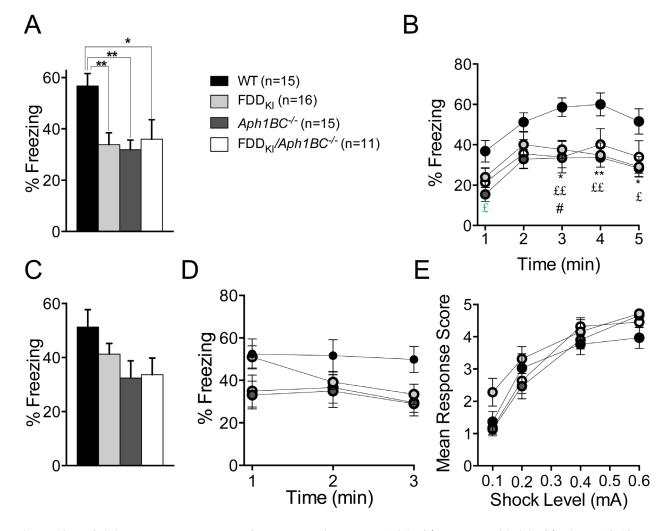


Figure 13: Deficit in long-term contextual fear memory in FDD_{KI}, *Aph1BC^{-/-}* and FDD_{KI}/*Aph1BC^{-/-}* mice at 18-19 months of age. Data are expressed as means \pm S.E.M. A. Percentage of freezing during the last 3-m period of the contextual test. WT mice froze significantly more than mice of the other genotypes. *p < 0.05, WT vs. FDD_{KI}/*Aph1BC^{-/-}*; **p < 0.01 WT vs. FDD_{KI} and WT vs. *Aph1BC^{-/-}*. B. Time course of freezing behavior during the contextual test in 1-m time bins. There was a significant genotype main effect (p < 0.01), with WT freezing more than the other genotypes. *p < 0.05, **p < 0.01, WT vs. FDD_{KI}: $\pm p < 0.05$, $\pm p < 0.01$, WT vs. *Aph1BC^{-/-}*. C. Percentage of freezing during the 3-m tone presentation in the altered context in the cued test. WT mice froze more than mice of the other genotypes, although not significantly. D. Time course of freezing behavior during tone presentation in the cued test. Only WT mice remained frozen for the entire period. E. Sensitivity to varying intensity levels of foot shock. Mice of all the genotypes reacted to shock at the four intensity levels in a similar manner.

deficits in learning and memory from ~4 to ~19 months of age. FDD_{k1}/Aph1BC^{-/-}mice presented mildly compromise accuracy of spatial long-term memory and working shortterm memory impairments at 4 and 8 months of age, respectively. Deficits in spatial and working memory were, at later ages, observed also in single mutant FDD_{KI} and Aph1BC^{-/-} mice. Analysis of older animals showed that spatial recognition memory and long-term contextual fear memory were impaired in FDD_{KI}, FDD_{KI}/Aph1BC^{-/-}, and Aph1BC^{-/-} mice. Overall, this study shows that; 1) the Aph1BC^{-/-} mutation does not rescue memory deficits in $FDD_{\kappa I}$ mice, as would have been expected if A β played a pathogenic role in FDD_{KI} mice; 2) in contrast, the FDD mutation and the $AphIBC^{-}$ deletion cause similar behavioral deficits in spatial memory and appear to have a small additive negative effect, at least on spatial longterm memory and working short-term memory, that is more pronounced at younger ages. Performing the twotrial Y-maze and the fear conditioning tasks in younger mice could unveil whether the FDD mutation and the Aph1BC deletion have also a negative additive effect on spatial recognition memory and long-term contextual fear memory. These observations are consistent with a pathological role of β -CTF, which is augmented in FDD_{KI} mice due to increased production and in Aph1BC^{-/-} mice due to reduced turnover.

We have previously shown that inhibition of $\beta\text{-}processing$ of APP rescues memory and synaptic impairments of FDD_{KI} mice acutely and transiently, suggesting that increased β -cleavage of APP, perhaps in the synaptic cleft, during synaptic events leading to LTP and memory acquisition, leads to memory/synaptic deficits in FDD_{KI} mice [25, 26]. This evidence, together with the data showing memory deficits in Aph1BC^{-/-} mice, suggest that either increasing the rate of production (like in FDD_{KI} mice) or decreasing the rate of clearance (as in Aph1BC mice) of β -CTF during synaptic transmission might lead to cognitive impairments. Future experiments will have to test whether these hypotheses are correct and whether the additive adverse effects on spatial long-term memory in young mice of the FDD_{KI} and the *Aph1BC^{-/-}* mutations are due to a synergistic effect on β -CTF levels transiently produced during synaptic transmission.

FDD_{KI} and *Aph1BC^{-/-}* mice show a few distinct phenotypes. In fear conditioning tests, mice carrying the *Aph1BC* deletion (FDD_{KI}/*Aph1BC^{-/-}* and *Aph1BC^{-/-}* mice) showed a mild impairment of cued fear memory, a task completely dependent on the functional integrity of the amygdala, as compared to FDD_{KI} and WT animals. This result is not surprising since several differences exist between FDD_{KI} and *Aph1BC^{-/-}* mice. First, the Aph1B/C deficiency causes a reduced clearance of β-CTF that leads to accumulation of β-CTF and a concomitant reduction in the β-CTF metabolites AID/AICD and Aβ. On the contrary, in FDD_{KI} mice the loss of mBRI2 leads to an increased production of all these APP metabolites. Second, in *Aph1BC*^{-/-} mice processing of another γ -secretase substrate, Neuregulin-1 [53] is reduced. Thus, behavioral deficits caused by the deletion of *Aph1B* and *Aph1C*, but not by the FDD mutation, could be attributed to reduction in AID/A β and/or reduction of processing of Neuregulin-1.

MATERIALS AND METHODS

Subjects

All behavioral experiments were conducted by using male littermates of the F2 generation of the C57BL/6J-129 hybrid mice as subjects. Mice were generated and maintained at the Animal facility of the Albert Einstein College of Medicine. Four genotypes, Aph1BC^{-/-}, FDD_{KP} $FDD_{\nu}/Aph1BC^{-}$, and wild type (WT), of F2 mice (n=11-19 per genotype) were evaluated for behavior. Aph1BC^{-/-} mice have been previously described [51]. FDD_{KI} mice carried one mutant and one wild type BRI2/ITM2b allele [19]. Upon weaning, all mice were implanted with electronic chips (PharmaSeq, Monmouth Junction, NJ) subcutaneously on the tail for identification purposes, and their identity was regularly checked during testing periods. Animals were group-housed in plastic cages with ad libitum access to food and water in a temperatureand humidity-controlled animal care facility with a 12-h light/12-h dark cycle. All experimental procedures were in accordance with the National Institutes of Health guidelines and approved by the Institutional Animal Care and Use Committee (IACUC) at the Albert Einstein College of Medicine in animal protocol number 20130509.

Behavioral experimental procedures

All mice were extensively handled prior to the start of behavioral testing. All behavioral testing was conducted during the light cycle. On each testing day, animals were transported to a behavioral testing suite in their home cages and allowed to acclimate for at least 30 min prior to the start of testing. The experimenter was not blind to the genotypes of the animals in tests conducted at 4, 7-8, 12 and 15 months of age but was made blind in tests conducted at 18-19 months. All measurements were taken automatically by video tracking software.

Elevated zero maze

Mice were assessed for anxiety-like behavior on the elevated zero maze initially at 4 months of age and again at 18-19 months. The zero maze (Stoelting, Wood Dale, IL) consisted of an annular platform (inner diameter 50 cm, width 5 cm) elevated to 50 cm above the ground level, divided equally into four quadrants. Two opposite quadrants were enclosed by walls (15 cm high) on both the inner and outer edges of the platform (closed areas), while the remaining two opposite quadrants were open without walls (open areas). Light levels over the maze were kept constant at approximately 50 lx in the open areas and 30 lx in the closed areas. Mice were placed individually in a closed quadrant and allowed to explore the maze freely for 5 min. The behavior of mice was monitored using a video camera, and their movements were analyzed with a video tracking system (ANY-maze, Stoelting). The percentage of time spent in the open and closed areas was used as measures of anxiety-like behavior, with larger time in the open arms indicating lower levels of anxiety.

Open field

The open field test was conducted to assess animals' general locomotor activity, exploratory behavior, and anxiety-like behavior at 4 and 18-19 months of age. The open field apparatus (Stoelting) consisted of a square open field (40 cm \times 40 cm) surrounded by opaque walls (35 cm high) and was dimly lit with a single light bulb directly above the apparatus, which illuminated the arena at approximately 5 lx in the center and 9 lx in corners. Each mouse was placed in the center of the open field box and allowed to explore the box freely for 10 min. The total distance traveled and the number of entries into, and the time spent in, the center of the arena (20 cm \times 20 cm) were recorded with the ANY-maze video tracking system. This was repeated for three consecutive days to assess how animals would habituate to the increasingly familiar environment.

Morris water maze

Mice were tested in the Morris water maze for spatial reference memory at 4, 7-8, and 12 months and for spatial working memory at 15 months of age. The water maze consisted of a circular tank (120 cm in diameter) filled with water made opaque with nontoxic white paint and maintained slightly above the room temperature (25 \pm 2 °C). All water maze tasks involved the animal finding a circular platform (10 cm in diameter) submerged in the water in order to escape from the water. On each trial, the mouse was released into the water facing the wall of the pool and allowed to swim freely in the pool to find the platform for the maximum of 60 s. Once the animal located the platform, it was allowed to stay on it for 15 s. Mice that did not locate the platform within 60 s were guided to the platform and allowed to stay on it for 15 s. After 15 s on the platform, the animals were removed from the pool, gently dried with paper towels, and placed in a single holding cage under a heat lamp until the next trial. A video tracking system (HVS 2020 and 2014; HVS Image, Mountain View, CA) was used to measure parameters such as the distance traveled, escape latency, swimming speed, the percentage of time spent in the quadrants, the number of counter crossings, and the average proximity to the platform location. The experimenter's position was maintained at the southeast (SE) corner of the room far from the tank for all the water maze tasks conducted at different ages. The experimenter was visible to the animal but remained stationary.

Visible platform task

At each age that mice were tested in the water maze, either before (4 and 7-8 months) or after (12, 15 and 18-19 months) the memory task, a visible platform task, in which the platform was made visible by attaching a small flag $(7 \text{ cm} \times 5 \text{ cm})$ to it, was conducted to examine whether mice had any visual, motor, or motivational deficits at that particular age. At 4, 7-8, 12, and 15 months of age, two or three daily sessions, with three or four trials a day, were given, in which both the platform location and the starting position were changed in a semi-random manner between trials to ensure that the animal was using the proximal cue (i.e., flag) to locate the platform. At 18-19 months, two three-trial sessions were given in a single day. The distance traveled to the platform (path length) and swimming speed were measured by the HVS video tracking system. The data collected from the last session were used for data analysis.

Reference memory task

The Morris water maze hidden platform task was performed to assess spatial reference memory at 4, 7-8, and 12 months of age. In this task, the platform was hidden 1 cm below the water level, and distal visual cues were placed on the walls surrounding the pool. The location of the hidden platform remained constant across the acquisition sessions, while the starting position was varied in a pseudo-random manner between trials within each session. The distance traveled by the mouse to reach the platform was recorded by the HVS video tracking system. At 4 months, mice received two daily sessions of three trials each with an inter-trial interval of 6-10 min and an inter-session interval of 3 h for six consecutive days. The platform was located at the center of the fourth quadrant between the center and the northwest (NW). Following a probe trial given two days after the last acquisition session of this initial reference memory task, mice received a reversal learning task for another six days, in which the location of the hidden platform was moved to the quadrant opposite to the original target quadrant (i.e., the first quadrant). At 7-8 months, the number of daily trials was reduced to three, with an intertrial interval of 6-10 min, and the task was run for five consecutive days. The platform was placed at the center of the second quadrant between the center and the southeast (SE). Mice were tested again at 12 months and received two trials per day with an inter-trial interval of 6-10 min for eight consecutive days. The platform position was at the center of the first quadrant between the center and the northeast (NE). Two days after the last acquisition session, a single probe trial was given, during which the platform was removed from the pool, and each mouse was released from the quadrant opposite to the trained platform location and allowed to search the pool for 60 s (4 and 7-8 months) or 30 s (12 months). The time spent in the target quadrant, where the platform had been located prior to its removal, the number of crossings of a circular area encompassing the original platform location (counter: 2 \times platform diameter), and the average proximity to the former platform location were measured to assess the animal's spatial reference memory for the location of the hidden platform.

Working memory task

Mice were tested in the Morris water maze for working memory at 15 months of age. In the working memory task, the position of the hidden platform varied from day to day but remained in the same place throughout the trials of a given day. The starting position was pseudorandomly changed from trial to trial within a given day. On each day, four trials (1 cued trial+ 3 test trials) were given. On the cued trial, each mouse was placed on the platform for 20s, after which it was removed from the platform to a single holding cage, where it remained for 5 min. After 5 min, three test trials were given, with an intertrial interval of 6-8 min. On each test trial, the animal was allowed to swim freely to find the hidden platform for the maximum of 60 s. Any mouse not locating the platform within 60 s was guided to the platform and allowed to stay on it for 15 s. Mice were tested for 10 days, with a one-day break after the seventh day. The distance traveled for each mouse was averaged over the last three days of testing and used for statistical analysis.

Radial arm water maze

Mice were tested in the radial arm water maze for spatial working memory at 7-8 and 18-19 months of age. An eight-arm radial arm water maze (each arm 8 cm in width and 38 cm in length) was used at 7-8 months. A six-armed radial arm water maze (each arm 20 cm in width and 30 cm in length) was used at 18-19 moths. The radial maze was placed into the same water tank as used for the Morris water maze, filled with opaque water (25 ± 2 °C). The height of the walls of the maze was 8 cm above the water level. Distal visual cues were placed on the walls of the testing room. A clear submerged platform (square, 8 cm × 8 cm for the 8-arm maze; circular, 10 cm in diameter

for the 6-arm maze) was placed at the end of one of the arms. In the working memory task, the platform would remain in the same arm for all trials on a given day but its location was changed pseudo-randomly from day to day. On each trial (maximum time 60 s), the mouse was released from one of the non-goal arms and allowed to swim freely to locate the platform. The release arm was pseudo-randomly changed from trial to trial. A mouse was charged with one error each time it entered an arm other than the goal arm or did not enter any arm for 15 s. The trial continued for 60 s or until the mouse ascended the platform. If a mouse did not locate the platform within 60 s, it was guided to the platform. The mouse was removed after 15 seconds on the platform. After three acquisition trials (inter-trial interval 6-8 min), the mouse was placed in a single holding cage for 30 min, after which it was given a fourth retention trial. The error scores for each mouse was averaged over the last three days of testing and used for statistical analysis. Mice were tested until the mean number of errors over three days for WT mice reached performance criteria (2.5 on Trial 3).

Y-maze

Mice were tested in the Y-maze for spatial recognition memory and spatial working memory at 18-19 months. The Y-maze consisted of three arms of equal length (35 cm) and width (5 cm), which were interconnected at 120° and enclosed by walls (10 cm high). The inside of the arms were identical, providing no intra-maze cues. The maze was placed under a bright fluorescent light and was surrounded by distal visual cues.

Two-trial test

The two-trial test test was conducted to assess short-term spatial recognition memory at 18-19 months. During the first trial (training trial), one of the arms of the maze was blocked, and mice were placed into one of the remaining arms of the maze (start arm) and allowed to explore the unblocked two arms for 10 min. After a 1-hr inter-trial interval, the blocked arm was opened (novel arm), and mice were placed in the start arm and allowed to explore freely all three arms of the maze for 5 min (test trial). The number of entries into and the amount of time spent in each arm were registered by the ANY-maze video tracking system. The relative position of the novel vs. known arms (i.e., left or right) was counterbalanced within each genotype to reduce place preference effects. This test takes advantage of the innate tendency of mice to explore novel unexplored areas (e.g., the previously blocked arm). Mice with intact recognition memory will prefer to explore a novel arm over the familiar arms, whereas mice with impaired spatial memory will enter all arms randomly.

Spontaneous alternation test

Ten to thirteen days after the two-trial test, the spontaneous alternation Y-mazetest was conducted to assess spatial working memory. Mice were released to the center of the Y maze with all three arms open and allowed to explore freely for 5 min. The number and the sequence of arm entries were recorded by a video tracking system (ANY-maze). The dependent variables were activity, defined as the number of arms entered, and percent alternation, which was calculated as the number of alternations (entries into three different arms consecutively) divided by the total possible alternations (i.e., the number of arms entered minus 2) and multiplied by 100. For efficient alternation, mice need to use working memory to maintain an ongoing record of most recently visited arms, and a mouse with impaired working memory cannot remember which arm it has just visited and shows decreased spontaneous alternation accordingly.

Fear conditioning

Mice were tested for contextual and cued fear memory at 18-19 months. Fear conditioning was conducted in a mouse conditioning chamber (18 cm \times 20 $cm \times 28 cm$) with a metal grid floor, lit with a single house light and enclosed within a sound-attenuating cubicle (Coulbourn Instruments, Whitehall, PA). The floor grid was connected to a shocker (Coulbourn Instruments) for the delivery of an electric foot shock, which was to be used as an unconditioned stimulus (US). The chamber was also equipped with a speaker connected to an amplifier for the delivery of a pure tone (2.8 kHz, 85 dB), which served as a conditioned stimulus (CS) for cued fear conditioning. The same conditioning chamber was used for testing for contextual fear memory, while, in testing for cued fear memory, the chamber was altered with a rectangular partition placed at a diagonal, wall and floor covers with novel texture, and a novel (vanilla) scent. On the first day, mice were individually placed in the conditioning chamber, and the house light was immediately turned on. One hundred and twenty seconds later, animals were presented with a continuous tone for 30 s, at the end of which an electric shock (0.6 mA) was delivered through the floor grid for 2 s and co-terminated with the tone. Mice remained in the chamber for another 30s before being removed to the home cage. Approximately 24 hr after the training session, animals were tested for contextual fear memory. The mouse was placed in the same conditioning chamber as had been used for training and observed for freezing behavior in the absence of any shock or tone for 5 min. The last 3-min period constituted testing for contextual fear memory. Approximately 24 hr after the test for contextual fear memory, mice were tested for cued fear memory. The animal was placed in the modified chamber (novel environment) and observed for freezing behavior for 2 min. After 2 min, the animal was presented with the tone CS continuously for 3 min, during which time it was again observed for freezing behavior. The last 3-min period with the tone presentation constituted testing for cued fear memory. In both tests, each animal's movements were recorded and the percentage of freezing was calculated by FreezeFrame software (Coulbourn Instruments).

At the completion of the test, mice were assessed for possible genotype effects on shock sensitivity. A sequence of single foot shocks was delivered to animals placed in the same chamber used for fear conditioning at four intensity levels (0.1, 0.2, 0.4, and 0.6 mV) in the ascending order. There were two presentations at each shock intensity level, with a 20-s inter-stimulus interval. At each intensity level, the animal's behavior was evaluated using the following scale to determine the threshold to each of these behavioral responses (0=no response, 1=ambulation, 2=flinch, 3=hop, 4=run, and 5=jump).

Statistical analysis

Statistical analysis of most data was performed by analysis of variance (ANOVA), with one between-subjects factor (genotype) and, when appropriate, a within-subjects factor (e.g., day). When significant effects were found, the data were further analyzed by *post hoc* comparison tests (Tukey's, Sidak's, Dunnett's, or Fisher's LSD). The level of significance was set at p < 0.05. Statistical analyses were carried out using the Prism software (GraphPad, La Jolla, CA).

ACKNOWLEDGMENTS

We thank Bart De Strooper and Lutgarde Serneels for providing the *Aph1B/C*^{-/-} mice.

CONFLICTS OF INTEREST

The AECOM has a patent on the commercial use of FDD_{KI} mice. Luciano D'Adamio is a co-inventor on this patent. AECOM has licensed the patent to Remegenix, a company of which Luciano D'Adamio is a co-founder and a Board member. As a co-founder Luciano D'Adamio owns 35% of Remegenix. The patent and the licensing only covers commercial use of the mice and does not pose any obstacle to distribution of the mice to academic laboratories. There are no further patents, products in development or marketed products to declare. This does not alter the authors' adherence to all the Oncotarget's policies on sharing data and materials, as detailed online in the guide for authors.

GRANT SUPPORT

The work was funded by the following grants: R01AG052286, D'ADAMIO, LUCIANO (PD/PI), R01AG041531, D'ADAMIO, LUCIANO (PD/PI), R01AG033007, D'ADAMIO, LUCIANO (PD/PI).

Author contributions

F.B. planned, performed and analyzed all the experiments; K.I. planned and conducted some of the experiments with F.B. and analyzed the data; D.D.P. helped generating the mice; L.D. conceived the experiments and produced the mouse strains, K.I., F.B. and L.D. prepared the figures and wrote the paper. All authors reviewed the manuscript.

REFERENCES

- Matsuda S, Giliberto L, Matsuda Y, Davies P, McGowan E, Pickford F, Ghiso J, Frangione B, D'Adamio L. The familial dementia BRI2 gene binds the Alzheimer gene amyloid-beta precursor protein and inhibits amyloid-beta production. J Biol Chem. 2005; 280:28912-28916.
- Matsuda S, Giliberto L, Matsuda Y, McGowan EM, D'Adamio L. BRI2 inhibits amyloid beta-peptide precursor protein processing by interfering with the docking of secretases to the substrate. J Neurosci. 2008; 28:8668-8676.
- Matsuda S, Matsuda Y, D'Adamio L. BRI3 inhibits amyloid precursor protein processing in a mechanistically distinct manner from its homologue dementia gene BRI2. J Biol Chem. 2009; 284:15815-15825.
- 4. Tanzi RE. The genetics of Alzheimer disease. Cold Spring Harb Perspect Med. 2012,2.
- Vidal R, Frangione B, Rostagno A, Mead S, Revesz T, Plant G, Ghiso J. A stop-codon mutation in the BRI gene associated with familial British dementia. Nature. 1999; 399:776-781.
- Garringer HJ, Murrell J, D'Adamio L, Ghetti B, Vidal R. Modeling familial British and Danish dementia. Brain Struct Funct. 2010; 214:235-244.
- Vidal R, Revesz T, Rostagno A, Kim E, Holton JL, Bek T, Bojsen-Moller M, Braendgaard H, Plant G, Ghiso J, Frangione B. A decamer duplication in the 3' region of the BRI gene originates an amyloid peptide that is associated with dementia in a Danish kindred. Proc Natl Acad Sci U S A. 2000; 97:4920-4925.
- De Strooper B. Loss-of-function presenilin mutations in Alzheimer disease. Talking Point on the role of presenilin mutations in Alzheimer disease. EMBO Rep. 2007; 8:141-146.
- 9. De Strooper B, Vassar R, Golde T. The secretases: enzymes with therapeutic potential in Alzheimer disease. Nat Rev Neurol. 2010; 6:99-107.

- 10. De Strooper B, Voet T. Alzheimer's disease: A protective mutation. Nature. 2012; 488:38-39.
- Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, Stefansson H, Sulem P, Gudbjartsson D, Maloney J, Hoyte K, Gustafson A, Liu Y, Lu Y, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature. 2012; 488:96-99.
- Matsuda S, Matsuda Y, Snapp EL, D'Adamio L. Maturation of BRI2 generates a specific inhibitor that reduces APP processing at the plasma membrane and in endocytic vesicles. Neurobiol Aging. 2011; 32:1400-1408.
- Choi SI, Vidal R, Frangione B, Levy E. Axonal transport of British and Danish amyloid peptides via secretory vesicles. FASEB J. 2004; 18:373-375.
- Kim SH, Creemers JW, Chu S, Thinakaran G, Sisodia SS. Proteolytic processing of familial British dementiaassociated BRI variants: evidence for enhanced intracellular accumulation of amyloidogenic peptides. J Biol Chem. 2002; 277:1872-1877.
- 15. Holton JL, Lashley T, Ghiso J, Braendgaard H, Vidal R, Guerin CJ, Gibb G, Hanger DP, Rostagno A, Anderton BH, Strand C, Ayling H, Plant G, et al. Familial Danish dementia: a novel form of cerebral amyloidosis associated with deposition of both amyloid-Dan and amyloid-beta. J Neuropathol Exp Neurol. 2002; 61:254-267.
- Askanas V, Engel WK, Alvarez RB, Frangione B, Ghiso J, Vidal R. Inclusion body myositis, muscle blood vessel and cardiac amyloidosis, and transthyretin Val122Ile allele. Ann Neurol. 2000; 47:544-549.
- 17. Holton JL, Ghiso J, Lashley T, Rostagno A, Guerin CJ, Gibb G, Houlden H, Ayling H, Martinian L, Anderton BH, Wood NW, Vidal R, Plant G, et al. Regional distribution of amyloid-Bri deposition and its association with neurofibrillary degeneration in familial British dementia. Am J Pathol. 2001; 158:515-526.
- Yan R, Vassar R. Targeting the beta secretase BACE1 for Alzheimer's disease therapy. Lancet Neurol. 2014; 13:319-329.
- Giliberto L, Matsuda S, Vidal R, D'Adamio L. Generation and initial characterization of FDD knock in mice. PLoS One. 2009; 4:e7900.
- Tamayev R, Matsuda S, Fa M, Arancio O, D'Adamio L. Danish dementia mice suggest that loss of function and not the amyloid cascade causes synaptic plasticity and memory deficits. Proc Natl Acad Sci U S A. 2010; 107:20822-20827.
- Tamayev R, Giliberto L, Li W, d'Abramo C, Arancio O, Vidal R, D'Adamio L. Memory deficits due to familial British dementia BRI2 mutation are caused by loss of BRI2 function rather than amyloidosis. J Neurosci. 2010; 30:14915-14924.
- Tamayev R, Matsuda S, Giliberto L, Arancio O, D'Adamio L. APP heterozygosity averts memory deficit in knockin mice expressing the Danish dementia BRI2 mutant. EMBO J. 2011; 30:2501-2509.

- 23. Tamayev R, D'Adamio L. Memory deficits of British dementia knock-in mice are prevented by Abeta-precursor protein haploinsufficiency. J Neurosci. 2012; 32:5481-5485.
- Tamayev R, Akpan N, Arancio O, Troy CM, D'Adamio L. Caspase-9 mediates synaptic plasticity and memory deficits of Danish dementia knock-in mice: caspase-9 inhibition provides therapeutic protection. Mol Neurodegener. 2012; 7:60.
- 25. Tamayev R, Matsuda S, Arancio O, D'Adamio L. beta- but not gamma-secretase proteolysis of APP causes synaptic and memory deficits in a mouse model of dementia. EMBO Mol Med. 2012; 4:171-179.
- Tamayev R, Matsuda S, D'Adamio L. beta but not gammasecretase proteolysis of APP causes synaptic and memory deficits in a mouse model of dementia. Mol Neurodegener. 2012; 7 Suppl 1:L9.
- 27. Matsuda S, Tamayev R, D'Adamio L. Increased AbetaPP processing in familial Danish dementia patients. J Alzheimers Dis. 2011; 27:385-391.
- Xia D, Watanabe H, Wu B, Lee SH, Li Y, Tsvetkov E, Bolshakov VY, Shen J, Kelleher RJ, 3rd. Presenilin-1 Knockin Mice Reveal Loss-of-Function Mechanism for Familial Alzheimer's Disease. Neuron. 2015; 85:967-981.
- 29. Shen J. Function and dysfunction of presenilin. Neurodegener Dis. 2014; 13:61-63.
- Wu B, Yamaguchi H, Lai FA, Shen J. Presenilins regulate calcium homeostasis and presynaptic function via ryanodine receptors in hippocampal neurons. Proc Natl Acad Sci U S A. 2013; 11015091-6.
- Heilig EA, Gutti U, Tai T, Shen J, Kelleher RJ, 3rd. Trans-Dominant Negative Effects of Pathogenic PSEN1 Mutations on gamma-Secretase Activity and Abeta Production. J Neurosci. 2013; 33:11606-11617.
- 32. Heilig EA, Xia W, Shen J, Kelleher RJ, 3rd. A presenilin-1 mutation identified in familial Alzheimer disease with cotton wool plaques causes a nearly complete loss of gamma-secretase activity. J Biol Chem. 2010; 285:22350-22359.
- Tabuchi K, Chen G, Sudhof TC, Shen J. Conditional forebrain inactivation of nicastrin causes progressive memory impairment and age-related neurodegeneration. J Neurosci. 2009; 29:7290-7301.
- Lee SH, Sharma M, Sudhof TC, Shen J. Synaptic function of nicastrin in hippocampal neurons. Proc Natl Acad Sci U S A. 2014; 111:8973-8978.
- Zhang C, Wu B, Beglopoulos V, Wines-Samuelson M, Zhang D, Dragatsis I, Sudhof TC, Shen J. Presenilins are essential for regulating neurotransmitter release. Nature. 2009; 460:632-636.
- 36. Saura CA, Choi SY, Beglopoulos V, Malkani S, Zhang D, Shankaranarayana Rao BS, Chattarji S, Kelleher RJ, 3rd, Kandel ER, Duff K, Kirkwood A, Shen J. Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration.

Neuron. 2004; 42:23-36.

- 37. Mitani Y, Yarimizu J, Saita K, Uchino H, Akashiba H, Shitaka Y, Ni K, Matsuoka N. Differential Effects between gamma-Secretase Inhibitors and Modulators on Cognitive Function in Amyloid Precursor Protein-Transgenic and Nontransgenic Mice. J Neurosci. 2012; 32:2037-2050.
- 38. Doody RS, Raman R, Farlow M, Iwatsubo T, Vellas B, Joffe S, Kieburtz K, He F, Sun X, Thomas RG, Aisen PS; Alzheimer's Disease Cooperative Study Steering Committee, Siemers E, Sethuraman G, Mohs R; Semagacestat Study Group. A phase 3 trial of semagacestat for treatment of Alzheimer's disease. N Engl J Med. 2013; 369:341-350.
- Bao J, Wolpowitz D, Role LW, Talmage DA. Back signaling by the Nrg-1 intracellular domain. J Cell Biol. 2003; 161:1133-1141.
- 40. De Strooper B, Annaert W, Cupers P, Saftig P, Craessaerts K, Mumm JS, Schroeter EH, Schrijvers V, Wolfe MS, Ray WJ, Goate A, Kopan R. A presenilin-1-dependent gamma-secretase-like protease mediates release of Notch intracellular domain. Nature. 1999; 398:518-522.
- 41. Kopan R, Ilagan MX. Gamma-secretase: proteasome of the membrane? Nat Rev Mol Cell Biol. 2004; 5:499-504.
- Nicolas M, Wolfer A, Raj K, Kummer JA, Mill P, van Noort M, Hui CC, Clevers H, Dotto GP, Radtke F. Notch1 functions as a tumor suppressor in mouse skin. Nat Genet. 2003; 33:416-421.
- Scheinfeld MH, Matsuda S, D'Adamio L. JNK-interacting protein-1 promotes transcription of A beta protein precursor but not A beta precursor-like proteins, mechanistically different than Fe65. Proc Natl Acad Sci U S A. 2003; 100:1729-1734.
- Scheinfeld MH, Ghersi E, Laky K, Fowlkes BJ, D'Adamio L. Processing of beta-amyloid precursor-like protein-1 and -2 by gamma-secretase regulates transcription. J Biol Chem. 2002; 277:44195-44201.
- 45. De Strooper B. Aph-1, Pen-2, and Nicastrin with Presenilin generate an active gamma-Secretase complex. Neuron. 2003; 38:9-12.
- Takasugi N, Tomita T, Hayashi I, Tsuruoka M, Niimura M, Takahashi Y, Thinakaran G, Iwatsubo T. The role of presenilin cofactors in the gamma-secretase complex. Nature. 2003; 422:438-441.
- 47. Edbauer D, Winkler E, Regula JT, Pesold B, Steiner H, Haass C. Reconstitution of gamma-secretase activity. Nat Cell Biol. 2003; 5:486-488.
- 48. Hebert SS, Serneels L, Dejaegere T, Horre K, Dabrowski M, Baert V, Annaert W, Hartmann D, De Strooper B. Coordinated and widespread expression of gamma-secretase in vivo: evidence for size and molecular heterogeneity. Neurobiol Dis. 2004; 17:260-272.
- Shirotani K, Edbauer D, Prokop S, Haass C, Steiner H. Identification of distinct gamma-secretase complexes with different APH-1 variants. J Biol Chem. 2004; 279:41340-

41345.

- Ma G, Li T, Price DL, Wong PC. APH-1a is the principal mammalian APH-1 isoform present in gamma-secretase complexes during embryonic development. J Neurosci. 2005; 25:192-198.
- 51. Serneels L, Dejaegere T, Craessaerts K, Horre K, Jorissen E, Tousseyn T, Hebert SS, Coolen M, Martens G, Zwijsen A, Annaert W, Hartmann D, De Strooper B. Differential contribution of the three Aph1 genes to gamma-secretase activity in vivo. Proc Natl Acad Sci U S A. 2005; 102:1719-1724.
- 52. Serneels L, Van Biervliet J, Craessaerts K, Dejaegere T, Horre K, Van Houtvin T, Esselmann H, Paul S, Schafer MK, Berezovska O, Hyman BT, Sprangers B, Sciot R, et al. gamma-Secretase heterogeneity in the Aph1 subunit: relevance for Alzheimer's disease. Science. 2009; 324:639-642.
- 53. Dejaegere T, Serneels L, Schafer MK, Van Biervliet J, Horre K, Depboylu C, Alvarez-Fischer D, Herreman A, Willem M, Haass C, Hoglinger GU, D'Hooge R, De Strooper B. Deficiency of Aph1B/C-gamma-secretase disturbs Nrg1 cleavage and sensorimotor gating that can be reversed with antipsychotic treatment. Proc Natl Acad Sci U S A. 2008; 105:9775-9780.
- 54. Maei HR, Zaslavsky K, Teixeira CM, Frankland PW. What is the Most Sensitive Measure of Water Maze Probe Test Performance? Front Integr Neurosci. 2009; 3:4.
- 55. Garthe A, Kempermann G. An old test for new neurons: refining the Morris water maze to study the functional relevance of adult hippocampal neurogenesis. Front Neurosci. 2013; 7:63.
- 56. Kim JJ, Fanselow MS. Modality-specific retrograde amnesia of fear. Science. 1992; 256:675-677.
- Fanselow M. Factors governing one-trial contextual conditioning. Animal Learning & Behavior. 1990; 18:264-270.
- Phillips RG, LeDoux JE. Differential contribution of amygdala and hippocampus to cued and contextual fear conditioning. Behav Neurosci. 1992; 106:274-285.
- 59. Anagnostaras SG, Gale GD, Fanselow MS. Hippocampus and contextual fear conditioning: recent controversies and advances. Hippocampus. 2001; 11:8-17.