

# Human T cell leukemia virus type 1 (HTLV-1) and oncogene or oncomiR addiction?

**Kuan-Teh Jeang**

<sup>1</sup> Kuan-Teh Jeang, Laboratory of Molecular Microbiology, National Institute of Allergy and Infectious Diseases, the National Institutes of Health, Bethesda, Maryland, USA 20892

**Correspondence to:** Kuan-Teh Jeang, e-mail: kjeang@nih.gov

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## ABSTRACT:

**The mechanism of HTLV-1 transformation of cells to Adult T cell leukemia (ATL) remains not fully understood. Currently, the viral Tax oncoprotein is known to be required to initiate transformation. Emerging evidence suggests that Tax is not needed to maintain the transformed ATL phenotype. Recent studies have shown that HTLV-1 transformed cells show deregulated expression of cellular microRNAs (miRNAs). Here we discuss the possibility that early ATL cells are Tax-oncogene-addicted while late ATL cells are oncogenic microRNA (oncomiR) – addicted. The potential utility of interrupting oncomiR addiction as a cancer treatment is broached.**

HTLV-1 was the first human retrovirus to be isolated. It was identified in 1980 by Robert Gallo and co-workers [1]; that initial finding was followed closely by important contributions from Japanese virologists [2]. HTLV-1 is causative of Adult T cell leukemia [3,4], a treatment refractory T cell cancer found endemically in Japan [5] and elsewhere [6]. Studies on this virus over the past three decades have provided insight into oncogene- and oncogenic microRNA- (oncomiR) addiction in leukemic transformation.

HTLV-1 encodes a viral Tax oncoprotein [7-9] whose expression confers prosurvival and proliferative properties to infected cells. Extant findings have shown that Tax is sufficient to transform human T cells [10,11]. Hence, the expression of Tax-alone in transgenic mice was found to be fully proficient for *in vivo* tumorigenesis [12-14]. Indeed, current data are consistent with the notion that Tax expression in infected humans greatly accelerates the *in vivo* cycling of T cells [15]. Intriguingly, when ATL patients are followed over time, a puzzling finding reveals that Tax expression *in vivo* is absent from approximately 60% of late leukemias [16]. Thus, unlike other virus-induced human malignancies such as the cervical cancers caused by human papilloma virus (HPV), in which the expression of the viral E6 and E7 oncoproteins are required for tumor maintenance [17], late ATL cells are

apparently not addicted to the Tax oncoprotein. Why might ATL cells extinguish Tax expression? A possible reason is because this viral protein represents the major target for cytotoxic T-lymphocytes (CTL) in infected patients [18,19]. Accordingly, the loss of Tax expression *in vivo* would facilitate the escape of virus-infected cells from CTL surveillance; and this seemingly would benefit disease progression.

A currently accepted model for ATL genesis by HTLV-1 is that the viral Tax oncogene is used for the initiation, but not the maintenance, of leukemogenesis (Figure 1). In this regard, the HTLV-1–ATL transformation mechanism appears not to subscribe to the oncogene addiction model of carcinogenesis [20]. What might then be some of the factor(s) needed for ATL cells to maintain their leukemic phenotype in the absence of Tax? One possible explanation rests with the observation that all ATL cells exhibit virus-mediated attenuation of the cell's spindle assembly checkpoint [21] and are thus highly aneuploid [9]. Potentially, this selected presentation of aneuploid chromosomes could be sufficient *per se* for maintaining the transformed ATL phenotype [22]. A second possibility is that transformed ATL cells have acquired altered expression of cellular microRNAs that are capable, in a Tax-independent fashion, of maintaining oncogenesis (e.g. oncomiRs [23] [24]).

Altered miRNA expression has indeed been linked to carcinogenesis. Early on, it was found that the loss of

miR-15a and miR-16-1 correlated with B-cell chronic lymphocytic leukemia [25]. Later, miRNA signatures for various cancers were described and linked to oncogenic transformation and found to be diagnostic of tumor types [23,26]. The deregulated expression of miRNAs in HTLV-1 transformed cells has also been reported in three independent publications [27-29]. In parsing the specific miRNA changes published in the three HTLV-1 studies, there appears to be very little overlap amongst most of the miRNA moieties [30]. Nonetheless, there was an intriguing consensus amongst the three findings. For example, in the study by Yeung *et al.*, the authors reported that the tumor suppressor protein TP53INP1 in HTLV-1-infected/transformed cells was targeted for repression by the upregulated expression of miR-93 and miR-130b [27]. By comparison, in the subsequent study by Pichler *et al.*, TP53INP1 was also reported to be targeted in HTLV-1 infected/transformed cells, but by the upregulated expression of miR-21, -24, -146a, and -155 [28]. Remarkably, separate from the *in vitro* HTLV-1 infected/transformed cells, Bellon *et al.* and Yeung *et al.* further investigated *in vivo* ATL leukemic cells from patients; and both noted upregulated miR-155 expression [27,29] which would be consistent with a silencing of TP53INP1 by miR-155 [31]. Thus, collectively, the three studies agree and converge on TP53INP1 as one of the important miRNA-regulated targets in ATL transformation by HTLV-1.

Based on the above data, one biological scenario is that late ATL cells may indeed be oncomiR-addicted while early ATL cells are Tax-oncogene-addicted (Figure 1). Recently, Watashi *et al.* have provided additional evidence that NIH 3T3 mouse cells can be transformed by singular over expression of either miR-93 or miR-130b [32]. They discovered two small molecule compounds

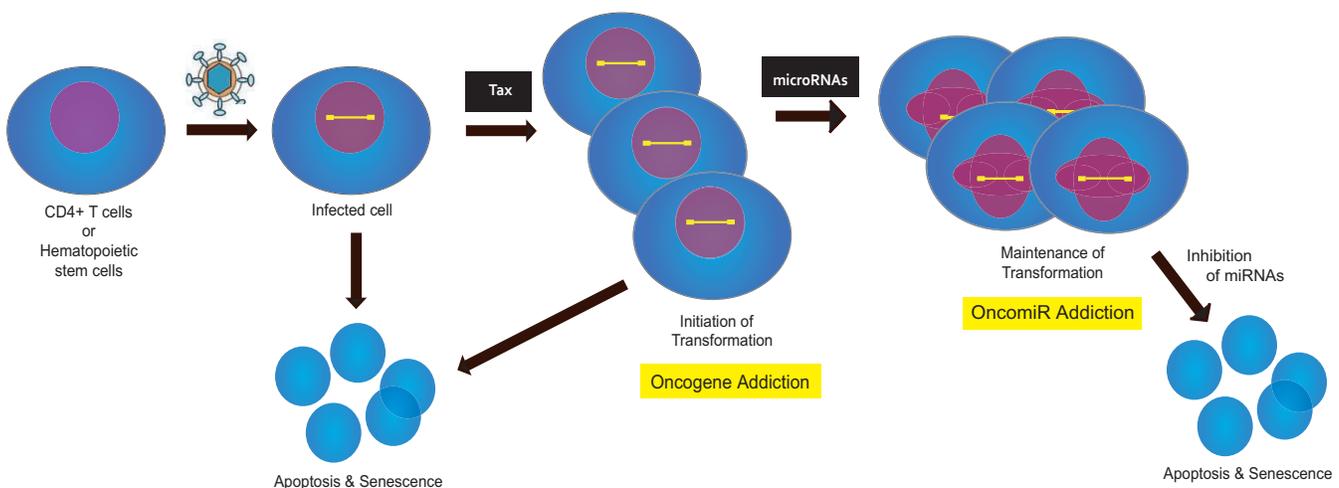
that can be used to reduce the over expression of miR-93 or miR-130b, and they showed that the treatment of miR-93- or miR-130b transformed NIH 3T3 cells using such compounds reversed tumorigenesis [32]. These results support the interpretation that in certain settings oncomiR-addicted tumors exist, and that this addiction could represent a potential treatment target for such cancers.

One might reason that a logical extension is to treat cancers by reducing oncomiR expression as well as targeting oncogene expression. Reality may be more complicated than this simple logic. Some studies have shown that a generalized down regulation of miRNAs is frequently seen in human cancers [26,33]. While it is not fully understood how general miRNA down regulations could propitiate carcinogenesis, such observations do raise caution that small molecule inhibitors of oncomiR activity needs to be utilized judiciously and monitored carefully to ensure that they ameliorate rather than exacerbate cancers. Further investigations are needed to conclusively verify oncomiR inhibition as an important treatment option in cancers.

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## REFERENCE LIST



**Figure 1. Potential stages of oncogene-addiction and oncomiR-addiction in HTLV-1 transformation of ATL leukemic T cells.** Virus-infected cells either initiate transformation after Tax expression or enter apoptosis/senescence. At this stage the cells could be regarded as Tax-oncogene-addicted. Subsequently, the expression of Tax in ATL cells is extinguished, and maintenance of the transformed phenotype in the cells is postulated to emerge from altered miRNA expression (oncomiR-addiction). Inhibition of the activity of oncomiRs can send such cells in tissue culture into apoptosis/senescence. (The figure is modified from Jeang, KT, *JFMA*, 2010, in press).

1. Poiesz BJ, Ruscetti FW, Gazdar AF, Bunn PA, Minna JD, Gallo RC: Detection and isolation of type C retrovirus particles from fresh and cultured lymphocytes of a patient with cutaneous T-cell lymphoma. *Proc Natl Acad Sci U S A* 1980, 77: 7415-7419.
2. Yoshida M, Miyoshi I, Hinuma Y: Isolation and characterization of retrovirus from cell lines of human adult T-cell leukemia and its implication in the disease. *Proc Natl Acad Sci U S A* 1982, 79: 2031-2035.
3. Gallo RC: The discovery of the first human retrovirus: HTLV-1 and HTLV-2. *Retrovirology* 2005, 2: 17.
4. Yoshida M: Discovery of HTLV-1, the first human retrovirus, its unique regulatory mechanisms, and insights into pathogenesis. *Oncogene* 2005, 24: 5931-5937.
5. Takatsuki K: Discovery of adult T-cell leukemia. *Retrovirology* 2005, 2: 16.
6. Proietti FA, Carneiro-Proietti AB, Catalan-Soares BC, Murphy EL: Global epidemiology of HTLV-I infection and associated diseases. *Oncogene* 2005, 24: 6058-6068.
7. Grassmann R, Aboud M, Jeang KT: Molecular mechanisms of cellular transformation by HTLV-1 Tax. *Oncogene* 2005, 24: 5976-5985.
8. Higuchi M, Fujii M: Distinct functions of HTLV-1 Tax1 from HTLV-2 Tax2 contribute key roles to viral pathogenesis. *Retrovirology* 2009, 6: 117.
9. Matsuoka M, Jeang KT: Human T-cell leukaemia virus type 1 (HTLV-1) infectivity and cellular transformation. *Nat Rev Cancer* 2007, 7: 270-280.
10. Rosin O, Koch C, Schmitt I, Semmes OJ, Jeang KT, Grassmann R: A human T-cell leukemia virus Tax variant incapable of activating NF-kappaB retains its immortalizing potential for primary T-lymphocytes. *J Biol Chem* 1998, 273: 6698-6703.
11. Robek MD, Ratner L: Immortalization of CD4(+) and CD8(+) T lymphocytes by human T-cell leukemia virus type 1 Tax mutants expressed in a functional molecular clone. *J Virol* 1999, 73: 4856-4865.
12. Grossman WJ, Kimata JT, Wong FH, Zutter M, Ley TJ, Ratner L: Development of leukemia in mice transgenic for the tax gene of human T-cell leukemia virus type I. *Proc Natl Acad Sci U S A* 1995, 92: 1057-1061.
13. Hasegawa H, Sawa H, Lewis MJ, Orba Y, Sheehy N, Yamamoto Y et al.: Thymus-derived leukemia-lymphoma in mice transgenic for the Tax gene of human T-lymphotropic virus type I. *Nat Med* 2006, 12: 466-472.
14. Ohsugi T, Kumasaka T, Okada S, Urano T: The Tax protein of HTLV-1 promotes oncogenesis in not only immature T cells but also mature T cells. *Nat Med* 2007, 13: 527-528.
15. Zane L, Sibon D, Jeannin L, Zandecki M, Fau-Larue MH, Gessain A et al.: Tax gene expression and cell cycling but not cell death are selected during HTLV-1 infection in vivo. *Retrovirology* 2010, 7: 17.
16. Takeda S, Maeda M, Morikawa S, Taniguchi Y, Yasunaga J, Nosaka K et al.: Genetic and epigenetic inactivation of tax gene in adult T-cell leukemia cells. *Int J Cancer* 2004, 109: 559-567.
17. Laughlin-Drubin ME, Munger K: Oncogenic activities of human papillomaviruses. *Virus Res* 2009, 143: 195-208.
18. Kannagi M, Harada S, Maruyama I, Inoko H, Igarashi H, Kuwashima G et al.: Predominant recognition of human T cell leukemia virus type I (HTLV-I) pX gene products by human CD8+ cytotoxic T cells directed against HTLV-I-infected cells. *Int Immunol* 1991, 3: 761-767.
19. Jacobson S, Shida H, McFarlin DE, Fauci AS, Koenig S: Circulating CD8+ cytotoxic T lymphocytes specific for HTLV-I pX in patients with HTLV-I associated neurological disease. *Nature* 1990, 348: 245-248.
20. Weinstein IB, Joe AK: Mechanisms of disease: Oncogene addiction--a rationale for molecular targeting in cancer therapy. *Nat Clin Pract Oncol* 2006, 3: 448-457.
21. Jin DY, Spencer F, Jeang KT: Human T cell leukemia virus type 1 oncoprotein Tax targets the human mitotic checkpoint protein MAD1. *Cell* 1998, 93: 81-91.
22. Chi YH, Jeang KT: Aneuploidy and cancer. *J Cell Biochem* 2007, 102: 531-538.
23. Croce CM: Causes and consequences of microRNA dysregulation in cancer. *Nat Rev Genet* 2009, 10: 704-714.
24. Scaria V, Jadhav V: microRNAs in viral oncogenesis. *Retrovirology* 2007, 4: 82.
25. Calin GA, Dumitru CD, Shimizu M, Bichi R, Zupo S, Noch E et al.: Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A* 2002, 99: 15524-15529.
26. Lu J, Getz G, Miska EA, Verez-Saavedra E, Lamb J, Peck D et al.: MicroRNA expression profiles classify human cancers. *Nature* 2005, 435: 834-838.
27. Yeung ML, Yasunaga J, Bennasser Y, Dusetti N, Harris D, Ahmad N et al.: Roles for microRNAs, miR-93 and miR-130b, and tumor protein 53-induced nuclear protein 1 tumor suppressor in cell growth dysregulation by human T-cell lymphotropic virus 1. *Cancer Res* 2008, 68: 8976-8985.
28. Pichler K, Schneider G, Grassmann R: MicroRNA miR-146a and further oncogenesis-related cellular microRNAs are dysregulated in HTLV-1-transformed T lymphocytes. *Retrovirology* 2008, 5: 100.
29. Bellon M, Lepelletier Y, Hermine O, Nicot C: Deregulation of microRNA involved in hematopoiesis and the immune response in HTLV-I adult T-cell leukemia. *Blood* 2009, 113: 4914-4917.
30. Ruggero K, Corradin A, Zanollo P, Amadori A, Bronte V, Ciminale V et al.: Role of microRNAs in HTLV-1 infection and transformation. *Mol Aspects Med* 2010.
31. Gironella M, Seux M, Xie MJ, Cano C, Tomasini R, Gommeaux J et al.: Tumor protein 53-induced nuclear protein 1 expression is repressed by miR-155, and its restoration inhibits pancreatic tumor development. *Proc*

Natl Acad Sci U S A 2007, 104: 16170-16175.

32. Watashi K, Yeung ML, Starost MF, Hosmane RS, Jeang KT: Identification of Small Molecules That Suppress MicroRNA Function and Reverse Tumorigenesis. *J Biol Chem* 2010, 285: 24707-24716.
33. Kumar MS, Pester RE, Chen CY, Lane K, Chin C, Lu J et al.: Dicer1 functions as a haploinsufficient tumor suppressor. *Genes Dev* 2009, 23: 2700-2704.