

## 6-Shogaol attenuates LPS-induced inflammation in BV2 microglia cells by activating PPAR- $\gamma$

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### ABSTRACT

**6-Shogaol, a pungent agent isolated from *Zingiber officinale Roscoe*, has been known to have anti-tumor and anti-inflammatory effects. However, the anti-inflammatory effects and biological mechanism of 6-Shogaol in LPS-activated BV2 microglia remains largely unknown. In this study, we evaluated the anti-inflammatory effects of 6-Shogaol in LPS-activated BV2 microglia. 6-Shogaol was administered 1 h before LPS treatment. The production of inflammatory mediators were detected by ELISA. The expression of NF- $\kappa$ B and PPAR- $\gamma$  were detected by western blot analysis. Our results revealed that 6-Shogaol inhibited LPS-induced TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and PGE2 production in a concentration dependent manner. Furthermore, 6-Shogaol inhibited LPS-induced NF- $\kappa$ B activation by inhibiting phosphorylation and nuclear translocation of NF- $\kappa$ B p65. In addition, 6-Shogaol could increase the expression of PPAR- $\gamma$ . Moreover, inhibition of PPAR- $\gamma$  by GW9662 could prevent the inhibition of 6-Shogaol on LPS-induced inflammatory mediator production. In conclusion, 6-Shogaol inhibits LPS-induced inflammation by activating PPAR- $\gamma$ .**

### INTRODUCTION

The incidence of neurodegenerative disease, particularly Parkinson disease (PD) and Alzheimer's disease, increased markedly in the last decades [1, 2]. Microglia, the major immune cells in the brain, plays a key role in host defence response to injury or infectious agents [3]. Microglia is exquisitely sensitive to brain injury and disease [4]. Overactivation of microglia leads to the production of inflammatory mediators which plays a critical role in the development of neuroinflammation [5, 6]. Neuroinflammation has recently been implicated as an important mechanism responsible for the pathological processes of neurodegenerative diseases [7, 8]. Therefore, the identification of agents to inhibit neuroinflammation might be an effective approach for the treatment of neurodegenerative diseases.

6-Shogaol, a pungent agent from *Zingiber officinale Roscoe*, has been reported to have anti-tumor and anti-inflammatory effects. 6-Shogaol has been reported to

protect against LPS-induced acute lung injury in mice. Also, 6-Shogaol was found to attenuate neuroinflammation and cognitive deficits in animal models of dementia [9]. Furthermore, 6-Shogaol has been reported to inhibit LPS-induced iNOS and COX-2 expression in macrophages [10]. In addition, studies showed that 6-Shogaol could protect against LPS-induced toxicity in murine astrocytes [11]. However, whether 6-Shogaol could inhibit LPS-induced anti-inflammatory response in activated microglial cells remains unclear. In the present study, we evaluated the anti-inflammatory effects of 6-Shogaol in LPS-stimulated BV2 microglia.

### RESULTS

#### Effects of 6-Shogaol on cell viability

To test whether 6-Shogaol has cytotoxicity on BV2 microglia, MTT assay were used in this study. The results showed that 6-Shogaol had no cytotoxicity on BV2

microglia at the concentration of 0 to 20  $\mu\text{g/mL}$  (Figure 1). Therefore, 6-Shogaol (5, 10, 20  $\mu\text{g/mL}$ ) were used in the following experiments.

### 6-Shogaol inhibited LPS-induced TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and PGE<sub>2</sub> production

To investigate the anti-inflammatory effects of 6-Shogaol, the expression of inflammatory mediators were detected in this study by ELISA. As shown in Figure 2, LPS dramatically increased the production of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and PGE<sub>2</sub>. However, 6-Shogaol concentration dependently down-regulated the production of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and PGE<sub>2</sub> induced by LPS.

### 6-Shogaol inhibited LPS-induced NF- $\kappa\text{B}$ activation

NF- $\kappa\text{B}$  has been known to be involved in the regulation of inflammatory mediators. To investigate the anti-inflammatory mechanism of 6-Shogaol, LPS-induced NF- $\kappa\text{B}$  activation were detected in the present study. The results showed that LPS significantly up-regulated the phosphorylation levels of NF- $\kappa\text{B}$  p65 and I $\kappa\text{B}\alpha$ . Pretreatment of 6-Shogaol concentration dependently inhibited LPS-induced NF- $\kappa\text{B}$  p65 phosphorylation and I $\kappa\text{B}\alpha$  phosphorylation and degradation (Figure 3).

### Effects of 6-Shogaol on PPAR- $\gamma$ expression

Previous studies showed that activation PPAR- $\gamma$  could inhibit LPS-induced NF- $\kappa\text{B}$  activation. Thus, we detected whether 6-Shogaol could up-regulated the expression of PPAR- $\gamma$ . As shown in Figure 4, 6-Shogaol

increased the expression of PPAR- $\gamma$  in a concentration dependent manner (Figure 4).

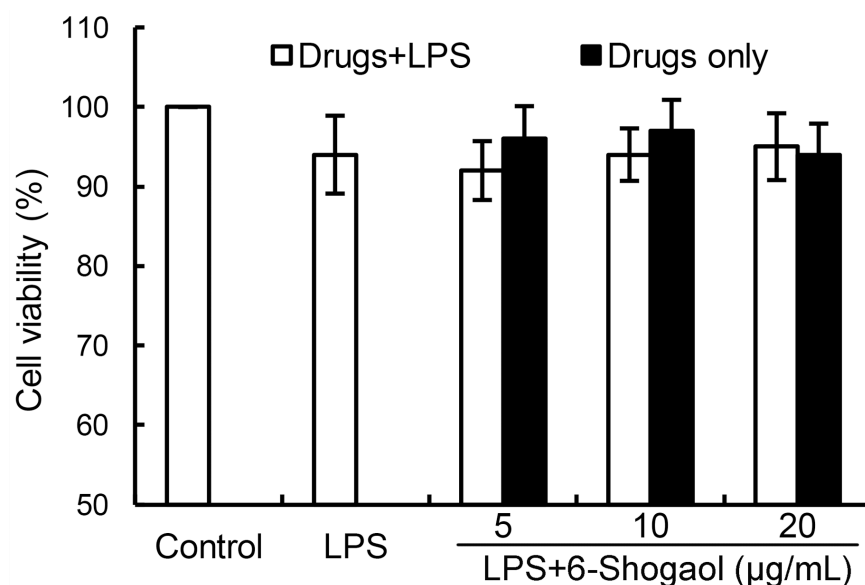
### GW9662 prevented the anti-inflammatory effects of 6-Shogaol

To further evaluate the anti-inflammatory mechanism of 6-Shogaol, PPAR- $\gamma$  was blocked by its inhibitor GW9662. As shown in Figure 5, our results indicated that the inhibition of 6-Shogaol on TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and PGE<sub>2</sub> production were prevented by GW9662. These results suggested that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia by activating PPAR- $\gamma$ .

## DISCUSSION

Microglia has been known to play an important role in neurodegenerative diseases [12]. Increasing evidences suggested that controlling the activation of microglia may have protective effects against neurodegenerative diseases [13]. In this study, the results showed that 6-Shogaol inhibited LPS-induced microglia activation by activating PPAR- $\gamma$ .

Microglia, the prime effector cells in the brain, plays a critical role in immune defense and inflammatory responses [14]. However, overactivation of microglia could lead to the pathological process of neurodegenerative diseases [15]. LPS has the ability to induce microglia activation, which lead to the release of inflammatory mediators [12, 16]. These inflammatory mediators, such as TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and PGE<sub>2</sub>, play an important role in the pathological process of

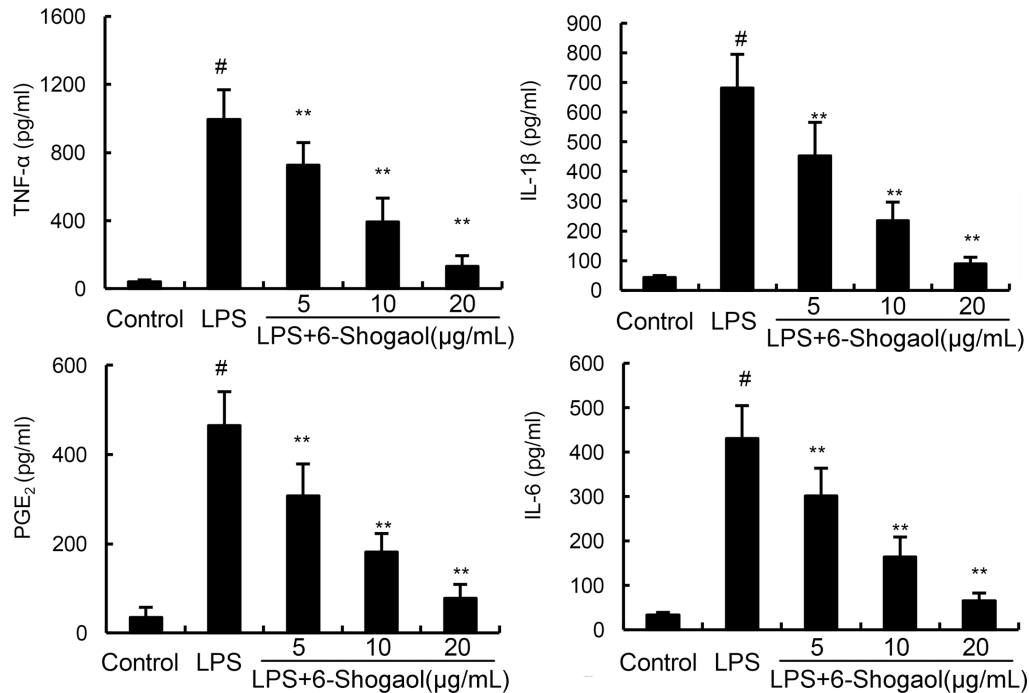


**Figure 1: Effects of 6-Shogaol on the cell viability of BV2 microglial cells.** Cells were cultured with different concentrations of 6-Shogaol (5, 10, 20  $\mu\text{g/mL}$ ) in the absence or presence of 0.5  $\mu\text{g/mL}$  LPS for 24 h. The cell viability was determined by MTT assay. The values presented are the means  $\pm$  SEM of three independent experiments.

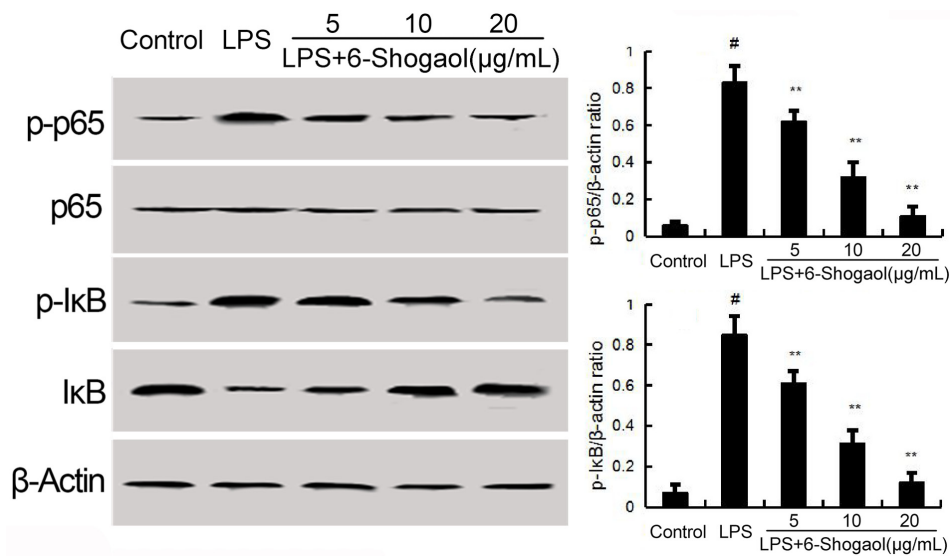
neurodegenerative diseases [17]. In the present study, our results showed that 6-Shogaol significantly inhibited LPS-induced inflammatory mediators production in BV2 microglia. The results indicated that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia.

It has been reported that NF- $\kappa$ B played a critical role in neuroinflammation [18]. LPS could induce NF- $\kappa$ B activation and inflammatory cytokines release [19].

Inhibition of LPS-induced NF- $\kappa$ B activation could attenuate neuroinflammation [20]. To clarify the anti-inflammatory mechanism of 6-Shogaol, NF- $\kappa$ B activation were measured in this study. We demonstrated that 6-Shogaol significantly inhibited LPS-induced NF- $\kappa$ B activation. PPAR- $\gamma$ , belongs to a nuclear receptor superfamily, is a ligand-activated transcription factor [21]. Activation of PPAR- $\gamma$  could regulate metabolism



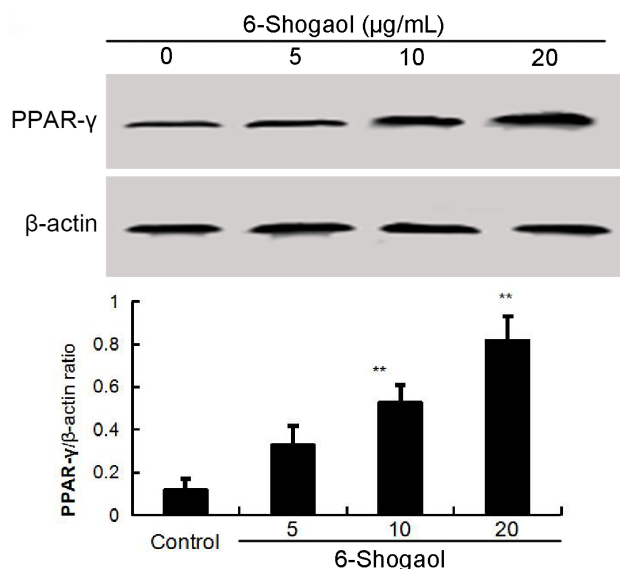
**Figure 2: Effects of 6-Shogaol on LPS-induced TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and PGE<sub>2</sub> production.** The production of TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and PGE<sub>2</sub> were measured by ELISA. The data presented are the means  $\pm$  SEM of three independent experiments. # $p$  < 0.05 vs. control group; \* $p$  < 0.05, \*\* $p$  < 0.01 vs. LPS group.



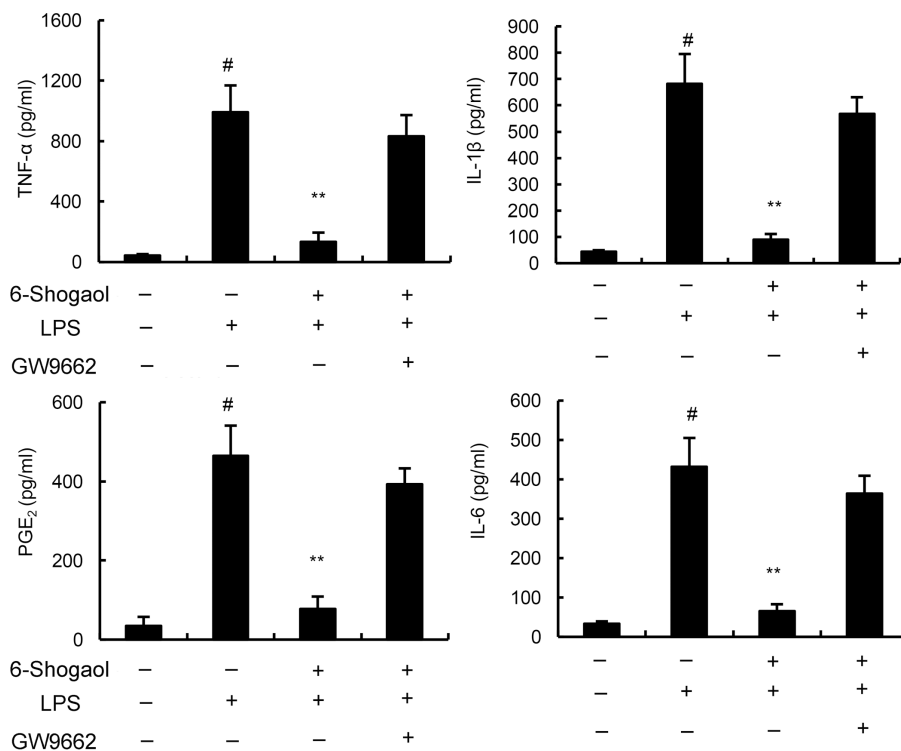
**Figure 3: Effects of 6-Shogaol on NF- $\kappa$ B expression.** The values presented are the means  $\pm$  SEM of three independent experiments. # $p$  < 0.05 vs. control group; \* $p$  < 0.05, \*\* $p$  < 0.01 vs. LPS group.

and inflammation [22]. Previous studies PPAR- $\gamma$  agonists inhibited LPS-induced airway inflammation [23]. Also, PPAR- $\gamma$  agonists could suppress LPS-induced inflammatory response in RAW264.7 cells [24].

Furthermore, PPAR- $\gamma$  agonists have been reported to have therapeutic role in diabetes, inflammation, and cancer [25]. In this study, our results showed that 6-Shogaol increased the expression of PPAR- $\gamma$ . And GW9662, a PPAR- $\gamma$



**Figure 4: Effects of 6-Shogaol on PPAR- $\gamma$  expression.** The values presented are the means  $\pm$  SEM of three independent experiments. # $p$  < 0.05 vs. control group; \* $p$  < 0.05, \*\* $p$  < 0.01 vs. LPS group.



**Figure 5: Effects of PPAR- $\gamma$  inhibitor GW9662 on the anti-inflammatory effects of 6-Shogaol.** Cells were treated with GW9662 for 12 h. Then, the cells were treated with 6-Shogaol and stimulated by LPS. The productions of inflammatory mediator were detected 24 h after LPS treatment. The values presented are the means  $\pm$  SEM of three independent experiments. # $p$  < 0.05 vs. control group; \* $p$  < 0.05, \*\* $p$  < 0.01 vs. LPS group.

inhibitor, could prevent the inhibition of 6-Shogaol on TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and PGE<sub>2</sub> production. These results indicated that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia by activating PPAR- $\gamma$ .

In conclusion, our results demonstrated that 6-Shogaol suppressed LPS-induced inflammatory mediators production by activating PPAR- $\gamma$ , which subsequently inhibited LPS-induced NF- $\kappa$ B activation. 6-Shogaol might be an effective agent for the treatment of neurodegenerative diseases.

## MATERIALS AND METHODS

### Materials

6-Shogaol (purity > 98%) was obtained from the National Institute for the Control of Pharmaceutical and Biological Products (Beijing, China). LPS (*Escherichia coli* O55:B5) and MTT were purchased from Sigma (St. Louis, MO, USA). ELISA kits of PGE<sub>2</sub>, TNF- $\alpha$ , IL-6 and IL-1 $\beta$  were purchased from BioLegend (San Diego, CA). PPAR- $\gamma$  monoclonal antibody was obtained from Santa Cruz Biotechnology (Heidelberg, Germany). NF- $\kappa$ B p65, I $\kappa$ B $\alpha$ , and  $\beta$ -actin monoclonal antibodies were obtained from Cell Signaling Technology Inc (Boston, MA, USA).

### Cell culture

Murine BV2 microglia cells were purchased from China Center for Type Culture Collection (CCTCC, Wuhan, China). The cells were cultured in DMEM with 5% fetal bovine serum, 100 U/ml penicillin, and 100 mg/ml streptomycin. The cells were treated with 6-Shogaol 1 h before LPS treatment.

### Cell viability

For determination of cell viability, MTT assay was applied in this study. BV2 microglia was incubated with 6-Shogaol alone and with LPS for 18 h. Then, the cells were treated with MTT for 4 h and the formazan formed was dissolved with DMSO (150  $\mu$ l/well). The optical density was determined at 570 nm using a Bio-Rad spectrophotometer.

### ELISA assay

24 h after LPS treatment, the levels of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, and PGE<sub>2</sub> in culture media were tested using commercially available ELISA kits (BioLegend, San Diego, CA). The assay was performed following the instructions provided by the manufacturers.

### Western blot analysis

The cells were lysed using RIAP lysis buffer and the concentration was measured by BCA method. Equal amount of protein was resolved using 12% SDS-

polyacrylamide gel. The proteins were transferred onto PVDF membranes. The membranes were blocked with 5% skimmed milk and incubated with primary antibodies and HRP-conjugated goat anti-rabbit IgG. The proteins were tested using the chemiluminescence detection system (Amersham, Berkshire, UK). Finally, the bands were analyzed using ImageJ software.

### Statistical analysis

Data were presented as means  $\pm$  SEM. Statistical comparison of the data were analyzed by one-way ANOVA with post-test Neuman-Keuls. A *p* value < 0.05 was considered as significant.

## CONFLICTS OF INTEREST

All authors declare that they have no conflicts of interest.

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## REFERENCES

1. Tanzi RE, Bertram L. Twenty years of the Alzheimer's disease amyloid hypothesis: a genetic perspective. *Cell*. 2005; 120:545–555.
2. Driver JA, Logroscino G, Gaziano JM, Kurth T. Incidence and remaining lifetime risk of Parkinson disease in advanced age. *Neurology*. 2009; 72:432–438.
3. Liu B, Hong JS. Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther*. 2003; 304:1–7.
4. Rock RB, Gekker G, Hu S, Sheng WS, Cheeran M, Lokensgard JR, Peterson PK. Role of microglia in central nervous system infections. *Clinical microbiology reviews*. 2004; 17:942–964.
5. Hanisch UK. Microglia as a source and target of cytokines. *Glia*. 2002; 40:140–155.
6. Block ML, Hong JS. Microglia and inflammation-mediated neurodegeneration: Multiple triggers with a common mechanism. *Prog Neurobiol*. 2005; 76:77–98.
7. Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH. Mechanisms Underlying Inflammation in Neurodegeneration. *Cell*. 2010; 140:918–934.
8. Zipp F, Aktas O. The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases. *Trends Neurosci*. 2006; 29:518–527.
9. Moon M, Kim HG, Choi JG, Oh H, Lee PK, Ha SK, Kim SY, Park Y, Huh Y, Oh MS. 6-Shogaol, an active constituent of ginger, attenuates neuroinflammation and

cognitive deficits in animal models of dementia. *Biochem Biophys Res Commun.* 2014; 449:8–13.

10. Pan MH, Hsieh MC, Hsu PC, Ho SY, Lai CS, Wu H, Sang SM, Ho CT. 6-Shogaol suppressed lipopolysaccharide-induced up-expression of iNOS and COX-2 in murine macrophages. *Mol Nutr Food Res.* 2008; 52:1467–1477.
11. Shim S, Kim S, Kwon YB, Kwon J. Protection by [6]-shogaol against lipopolysaccharide-induced toxicity in murine astrocytes is related to production of brain-derived neurotrophic factor. *Food Chem Toxicol.* 2012; 50:597–602.
12. Dheen ST, Kaur C, Ling EA. Microglial activation and its implications in the brain diseases. *Curr Med Chem.* 2007; 14:1189–1197.
13. Liu B, Hong JS. Role of microglia in inflammation-mediated neurodegenerative diseases: Mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther.* 2003; 304:1–7.
14. Glezer I, Simard AR, Rivest S. Neuroprotective role of the innate immune system by microglia. *Neuroscience.* 2007; 147:867–883.
15. Block ML, Zecca L, Hong JS. Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nat Rev Neurosci.* 2007; 8:57–69.
16. Wang MJ, Lin WW, Chen HL, Chang YH, Ou HC, Kuo JS, Hong JS, Jeng KC. Silymarin protects dopaminergic neurons against lipopolysaccharide-induced neurotoxicity by inhibiting microglia activation. *Eur J Neurosci.* 2002; 16:2103–2112.
17. Smith JA, Das A, Ray SK, Banik NL. Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases. *Brain Res Bull.* 2012; 87:10–20.
18. Dilshara MG, Jayasooriya RG, Lee S, Choi YH, Kim GY. Morin downregulates nitric oxide and prostaglandin E2 production in LPS-stimulated BV2 microglial cells by suppressing NF-kappaB activity and activating HO-1 induction. *Environmental toxicology and pharmacology.* 2016; 44:62–68.
19. Covert MW, Leung TH, Gaston JE, Baltimore D. Achieving stability of lipopolysaccharide-induced NF-kappaB activation. *Science.* 2005; 309:1854–1857.
20. Wang YP, Wu Y, Li LY, Zheng J, Liu RG, Zhou JP, Yuan SY, Shang Y, Yao SL. Aspirin-triggered lipoxin A4 attenuates LPS-induced pro-inflammatory responses by inhibiting activation of NF-kappaB and MAPKs in BV-2 microglial cells. *Journal of neuroinflammation.* 2011; 8:95.
21. Yessoufou A, Wahli W. Multifaceted roles of peroxisome proliferator-activated receptors (PPARs) at the cellular and whole organism levels. *Swiss Med Wkly.* 2010; 140:w13071.
22. Chawla A, Barak Y, Nagy L, Liao D, Tontonoz P, Evans RM. PPAR-gamma dependent and independent effects on macrophage-gene expression in lipid metabolism and inflammation. *Nat Med.* 2001; 7:48–52.
23. Birrell MA, Patel HJ, McCluskie K, Wong S, Leonard T, Yacoub MH, Belvisi MG. PPAR-gamma agonists as therapy for diseases involving airway neutrophilia. *Eur Respir J.* 2004; 24:18–23.
24. Huang C, Yang Y, Li WX, Wu XQ, Li XF, Ma TT, Zhang L, Meng XM, Li J. Hyperin attenuates inflammation by activating PPAR-gamma in mice with acute liver injury (ALI) and LPS-induced RAW264.7 cells. *Int Immunopharmacol.* 2015; 29:440–447.
25. Murphy GJ, Holder JC. PPAR-gamma agonists: therapeutic role in diabetes, inflammation and cancer. *Trends Pharmacol Sci.* 2000; 21:469–474.