

Impact of HLA-B*58:01 allele and allopurinol-induced cutaneous adverse drug reactions: evidence from 21 pharmacogenetic studies

Ran Wu^{1,*}, Yi-ju Cheng^{2,3,*}, Li-li Zhu⁴, Lei Yu⁵, Xue-ke Zhao⁶, Min Jia¹, Chang-hui Wen¹, Xing-zhen Long¹, Ting Tang¹, Ai-juan He¹, Yi-yan Zeng¹, Zun-feng Ma¹, Zhi Zheng³, Mu-zi Ni³, Gong-jing Cai³

¹Department of Dermatology, The First Affiliated Hospital of Guiyang College of Traditional Chinese Medicine, Guiyang 550000, Guizhou, China

²Department of Respiratory Medicine, The Affiliated Hospital of Guizhou Medical University, Guiyang 550004, Guizhou, China

³Department of Respiratory Medicine, The Affiliated Baiyun Hospital of Guizhou Medical University, Guiyang 550014, Guizhou, China

⁴Blood Transfusion Department, The Affiliated Baiyun Hospital of Guizhou Medical University, Guiyang 550014, Guizhou, China

⁵Prenatal Diagnostic Center, The Affiliated Hospital of Guizhou Medical University, Guiyang 550004, Guizhou, China

⁶Department of Infectious Diseases, The Affiliated Hospital of Guizhou Medical University, Guiyang 550004, Guizhou, China

*These authors have contributed equally to this work

Correspondence to: Lei Yu, email: gmcLei_Yu@hotmail.com
Min Jia, email: Jiamin0851@yahoo.com.cn

Keywords: allopurinol, cutaneous adverse drug reactions, HLA-B*58:01, diagnosis, meta-analysis

Received: July 28, 2016

Accepted: October 27, 2016

Published: November 09, 2016

ABSTRACT

Allopurinol is widely used for hyperuricemia and gouty arthritis, but is associated with cutaneous adverse drug reactions (CADRs). Recently, HLA-B*58:01 allele was identified as a strong genetic marker for allopurinol-induced CADRs in Han Chinese. However, the magnitude of association and diagnosis value of HLA-B*58:01 in allopurinol-induced CADRs remain inconclusive. To investigate this inconsistency, we conducted a meta-analysis of 21 pharmacogenetic studies, including 551 patients with allopurinol-induced CADRs, and 2,370 allopurinol-tolerant controls as well as 9,592 healthy volunteers. The summary OR for allopurinol-induced CADRs among HLA-B*58:01 carriers was 82.77 (95% CI: 41.63 – 164.58, $P < 10^{-5}$) and 100.87 (95% CI: 63.91 – 159.21, $P < 10^{-5}$) in matched and population based studies, respectively. Significant results were also observed when stratified by outcomes and ethnicity. Furthermore, the summary estimates for quantitative analysis of HLA-B*58:01 allele carriers in allopurinol-induced CADRs screening were as follows: sensitivity, 0.93 (95% CI: 0.85 – 0.97); specificity, 0.89 (95% CI: 0.87 – 0.91); positive likelihood ratio, 8.24 (95% CI: 6.92 – 9.81); negative likelihood ratio, 0.084 (95% CI: 0.039 – 0.179); and diagnostic odds ratio, 98.59 (95% CI: 43.31 – 224.41). The AUSROC was 0.92 (95% CI: 0.89–0.94), indicating the high diagnostic performance. Our results indicated that allopurinol–SCAR is strongly associated with HLA-B*58:01, and HLA-B*58:01 is a highly specific and effective genetic marker for the detection allopurinol-induced CADRs, especially for Asian descents.

INTRODUCTION

Allopurinol, a structural analog of hypoxanthine, is an effective xanthine oxidase inhibitor that has been widely used as antihyperuricemic agent [1]. In general, allopurinol

is well tolerated with gastrointestinal discomfort being the most frequent complaint. However, allopurinol causes a variety of cutaneous adverse drug reactions (CADRs) ranging from milder form, such as maculopapular eruption (MPE), to severe cutaneous adverse reactions (SCARs)

including drug-induced hypersensitivity syndrome (HSS), Stevens–Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN) [2]. Although SCARs rarely occur, the mortality rate ranges from 5 - 10% in SJS, 10% in HSS, and increases to 30 – 40% in TEN [2–4].

Allopurinol-induced CADR is regarded as a complex process with interaction between environmental and genetic factors related to drug metabolism and immune responses. Environmental factors such as cigarette smoking, alcohol abuse, drug-drug interactions, pre-existing diseases (e.g., diabetes, chronic kidney disease), and viral infections have been already well studied so far [5]. To investigate the relationship between human leucocyte antigen (HLA) genetic markers and CADR induced by allopurinol, recent pharmacogenetic studies have shown HLA-B*58:01 allele as the most strong association signal for allopurinol-induced CADR [6–8]. However, inconsistent findings were subsequently reported [9, 10]. Individual study may have failed to detect difference due to inadequate statistical power, phenotypic heterogeneity, multiple hypothesis testing, and publication bias. Besides, accumulated evidences have been reported in recent years and there is a need to reconcile these data. Furthermore, HLA-B*58:01 genotyping is a cost-prohibitive test for routine clinical practice, which are mainly used in medical research rather than in clinical practice [11]. Moreover, uncertainty still persists about the clinical performance of HLA-B*58:01 genotype for diagnosing of SCARs caused by allopurinol. Here, we conducted a comprehensive meta-analysis from all eligible pharmacogenetic studies to assess the association of HLA-B*58:01 allele in the development of allopurinol-induced CADR and to evaluate the diagnosis value of CADR.

RESULTS

Literature selection and studies characteristics

The flow of our literature search is shown in Supplementary Figure S1. We identified 308 records after searching different databases. After reviewing the title and abstracts, 287 records were excluded. After full-text review, the remaining 21 studies [7–10, 12–28] were included in our study, with 12,513 individuals in total, including 551 patients with allopurinol-induced CADR. The 11,962 individuals without allopurinol-induced CADR were included in these studies as control groups, which comprised 2,370 allopurinol-tolerant controls from 16 matched studies and 9,592 healthy volunteers or general populations from 13 studies. Most studies were conducted among East Asian populations, 2 studies examined individuals of white race [16, 24], and 1 studies evaluated multi-ethnic populations [27]. Ten studies reported the allopurinol dosages data [7, 9, 14, 15, 17, 20–23, 28], while 9 studies [7, 9, 17, 20–23, 25, 28] provided

information on allopurinol exposure duration. Most studies (except for the study by Ye et al [14] and study by Zeng [15]) specified the diagnostic criteria for SJS and TEN cases [29, 30]. The main study characteristics were summarized in Supplementary Table S1. Additionally, only the general population data from the study by Hung et al [7] were used in the overall comparison [7] as for sample overlapping.

Overall association of HLA-B*58:01 with allopurinol-induced CADR risk

Table 1 shows the summary of the meta-analysis for HLA-B*58:01 and allopurinol-induced CADR. Overall, the HLA-B*58:01 allele showed a strong association with the risk of allopurinol-induced CADR in matched studies (OR = 82.77, 95% CI: 41.63 – 164.58, $P < 10^{-5}$; Figure 1) and population-based studies (OR = 100.87, 95% CI: 63.91 – 159.21, $P < 10^{-5}$; Figure 2). When only the severe form of CADR were considered, a significant increased risks of allopurinol-induced SCARs for carrier of the HLA-B*58:01 allele were detected for matched studies and population-based studies with OR of 92.06 (95% CI: 59.54 – 142.32, $P < 10^{-5}$) and 108.39 (95% CI: 73.73 – 159.36, $P < 10^{-5}$), respectively. In addition, significantly increased risk for SJS/TEN was observed among studies using matched control (OR = 79.01, 95% CI: 44.23 – 141.12, $P < 10^{-5}$), and population-control (OR = 106.48, 95% CI: 65.66 – 172.66, $P < 10^{-5}$). When all included studies were stratified based on ethnicity, significantly increased risks of allopurinol-induced CADR among HLA-B*58:01 carrier were found both in Asians and Caucasians (Table 1). For multiple testing, all associations remain significant after Bonferroni correction. Statistical amount of between-study heterogeneity was found (I^2 values $> 50\%$); we therefore conducted a meta-regression analysis which showed that the study size may be the source of heterogeneity ($P < 0.05$). By contrast, source of controls, study quality, age, sex, allopurinol dosage and exposure duration were not correlated with the overall ORs ($P > 0.05$). To further explore sources of heterogeneity between individual studies, Galbraith plot analyses were used and 2 studies were identified (Supplementary Figure S2).

MPE and EEM have been considered to be distinct from SJS/TEN, the overall association between HLA-B*58:01 carriers with MPE and EEM is much weaker than SCARs caused by allopurinol. Overall, the random-effect OR of the HLA-B*58:01 for MPE and EEM was 29.33 (95% CI: 5.89 – 145.98, $P < 10^{-4}$; Supplementary Figure S3) and 12.95 (95% CI: 2.30–72.85, $P = 0.004$; Supplementary Figure S4), respectively.

Under recessive genetic model, we further analysed the gene dosage effect of HLA-B*58:01 on -CADR induced by allopurinol. The distribution of HLA-B*58:01 genotypes among CADR cases and tolerant controls

Table 1: Results of meta-analysis for HLA-B*58:01 with allopurinol-induced CADR

Overall and subgroups analyses	No. of data sets	No. of cases/controls	OR (95% CI)	P(Z)	P(Q)	I ² (%)
Study with tolerant controls						
All types of CADR	16	551/2370	82.77 (41.63-164.58)	<10 ⁻⁵	0.001	60.3
CADR in Asians	15	526/2347	87.66 (42.44-181.10)	<10 ⁻⁵	0.001	63.0
CADR in Caucasians	1	25/23	39.11 (4.49-340.50)	0.001	NA	NA
SCARs	16	466/2370	92.06 (59.54-142.32)	<10 ⁻⁵	0.66	0
SCARs in Asians	15	441/2347	95.45 (61.18-148.91)	<10 ⁻⁵	0.63	0
SCARs in Caucasians	1	25/23	39.11 (4.49-340.50)	0.001	NA	NA
SJS/TEN	14	211/2207	79.01 (44.23-141.12)	<10 ⁻⁵	0.83	0
SJS/TEN in Asians	13	205/2184	81.42 (44.92-147.51)	<10 ⁻⁵	0.79	0
SJS/TEN in Caucasians	1	6/23	44.00 (3.18-608.16)	0.005	NA	NA
MPE	6	99/548	29.33 (5.89-145.98)	<10 ⁻⁴	0.003	72.1
MPE in Asians	5	93/525	40.45 (6.43-254.57)	<10 ⁻⁴	0.002	76.8
MPE in Caucasians	1	6/23	4.40 (0.23-82.98)	0.32	NA	NA
EEM (All Asians)	3	9/112	12.95 (2.30-72.85)	0.004	0.38	0
Study with population controls						
All types of CADR	13	414/9592	100.87 (63.91-159.21)	<10 ⁻⁵	0.19	25.5
CADR in Asians	10	351/4455	122.57 (73.79-203.84)	<10 ⁻⁵	0.42	2.5
CADR in Caucasians	3	63/5137	64.59 (25.42-164.11)	<10 ⁻⁵	0.09	58.3
SCARs	13	381/9592	108.39 (73.73-159.36)	<10 ⁻⁵	0.56	0
SCARs in Asians	10	318/3360	147.88 (86.69-252.25)	<10 ⁻⁵	0.98	0
SCARs in Caucasians	3	63/5137	64.59 (25.42-164.11)	<10 ⁻⁵	0.09	58.3
SJS/TEN	12	190/9312	106.48 (65.66-172.66)	<10 ⁻⁵	0.73	0
SJS/TEN in Asians	9	146/4175	156.32 (78.22-312.41)	<10 ⁻⁵	0.99	0
SJS/TEN in Caucasians	3	44/5137	58.35 (16.90-201.54)	<10 ⁻⁵	0.09	58.7

MPE: Maculopapular eruption; SJS: Stevens–Johnson syndrome; TEN: Toxic epidermal necrolysis.

NA: not available; EEM: erythema exudativum multiforme.

P(Z): Z test used to determine the significance of the overall OR.

P(Q): Cochran’s Chi-square Q statistic used to assess the heterogeneity in subgroups.

were extracted from 4 studies. Patients with homozygous HLA-B*58:01 had 16.78-fold (95% CI: 5.90 – 47.73, $P < 10^{-5}$; Supplementary Figure S5) risk of CADR compared with individuals with one or no copy of the risk allele.

Sensitivity analyses confirmed the significant association of HLA-B*58:01 with allopurinol-induced CADR (Supplementary Figure S6). No small study effects were observed according to funnel plot inspection (Egger’s test $P > 0.05$; Supplementary Figure S7).

Diagnosis value of HLA-B*58:01 on allopurinol-induced CADR

For diagnosis allopurinol-induced CADR, the summary specificity was 0.89 (95% CI: 0.87 – 0.91), and the sensitivity was 0.93 (95% CI: 0.85 – 0.97; Figure 3). The pooled PLR was 8.24 (95% CI: 6.92 – 9.81); whereas, the NLR was 0.084 (95% CI: 0.039 – 0.179). The pooled DOR of HLA-B*58:01 was 98.59 (95% CI: 43.31 –

224.41), with significant heterogeneity ($I^2=98.6$, $Q=21.7$, $P < 10^{-5}$). To assess covariates, univariate meta-regression find that ethnic population ($P = 0.01$) and study size ($P = 0.02$) may affect the ability of HLA-B*58:01 to diagnosis CADR.

As shown in Figure 4, the HSROC curve indicated a high level of overall accuracy as measured by AUC (0.92, 95% CI: 0.89 – 0.94). The Deek’s regression test was performed to detect potential small study effects and no significant selection or publication bias was detected ($P = 0.35$, Supplementary Figure S8).

DISCUSSION

Because of the rareness of serious adverse drug reactions, pharmacogenetic studies were underpowered to detect modest differences on small sample sizes. Obtaining adequate numbers of cases with adverse drug reactions for

specific drug-gene interaction, and thus improves safety before drug administration is a real challenge. Quantitative synthesis of data from single study, meta-analysis with sufficient power is helpful to address this issue [31]. The present meta-analysis of 21 pharmacogenetic studies, involving 12,513 individuals provided evidence regarding the casual relationship between HLA-B*58:01 carriers and the development of allopurinol-related CADR; while the association of HLA-B*58:01 allele with with allopurinol-induced MPE or EEM was much weaker.

We observed significant heterogeneity between studies and conducted subgroup analyses and meta-regression to investigate the potential sources. In fact, ethnicity was identified as a potential source of heterogeneity and the effect estimates in Asians (OR range from 73 to 152) was much stronger than that in white populations (OR range from 39 to 64). Thus, differences in genetic background could influence the response to allopurinol. Indeed, the HLA-B*58:01 allelic frequency

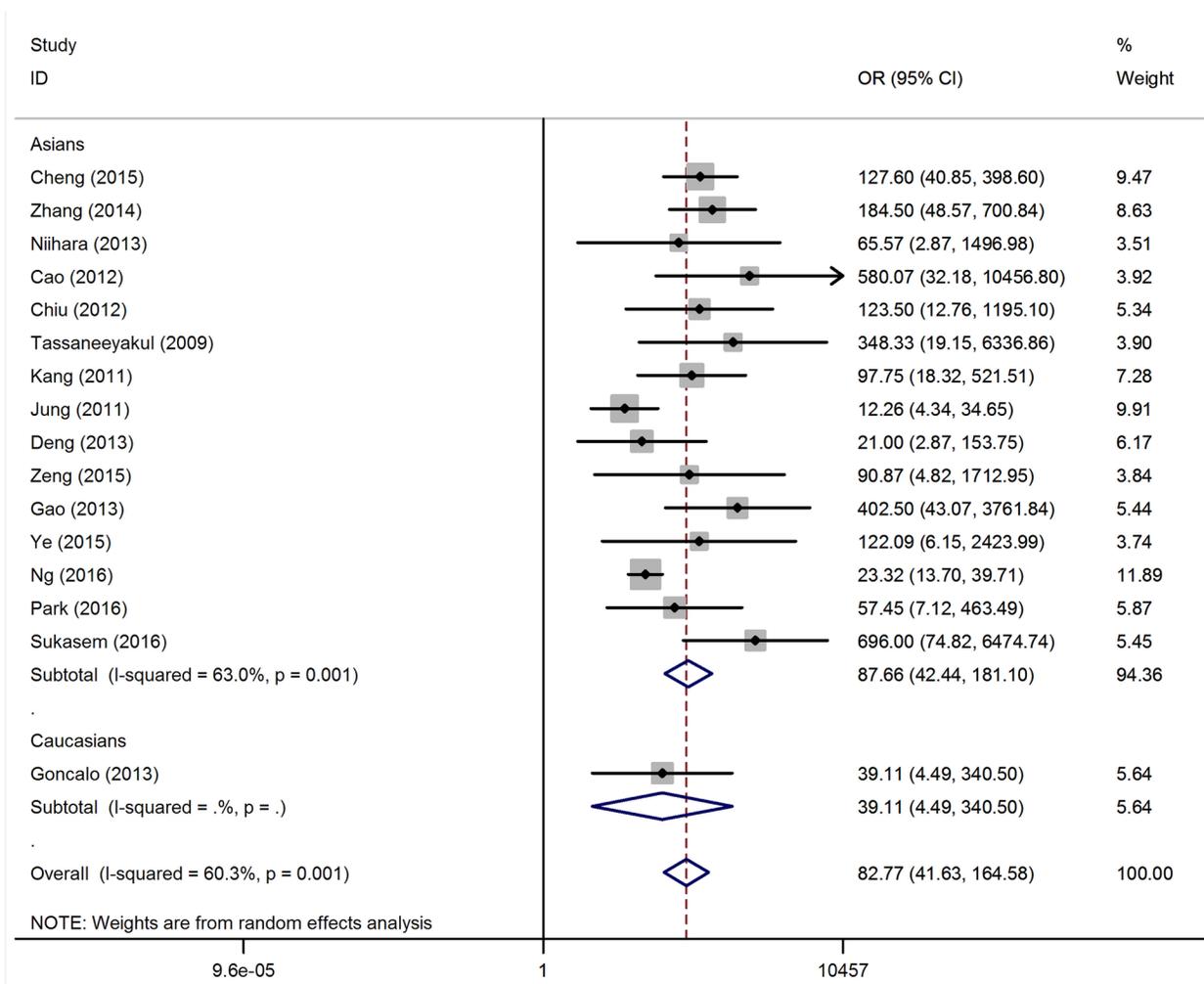


Figure 1: Forest plot for the meta-analysis of the association between HLA-B*58:01 allele carriers and risk of allopurinol-induced CADR stratified by ethnicity in matched study.

is much higher in Asians (10 – 15%) than Europeans (1 – 3%) [32]. Due to limited allopurinol-induced CADR patients investigated in Caucasians, it's possible that different effect estimates among ethnic groups might arise simply by chance as for insufficient statistical power. Therefore, more studies with large sample size are warranted to validate the effect of HLA-B*58:01 on allopurinol-induced CADR among difference ethnic populations. To check the influence of individual study, sensitivity analysis was performed and showed robust associations even when the largest study was removed.

The area under ROC serves as a global measure of diagnostic performance. According to the suggested guideline for interpretation of area under ROC [33], HLA-B*58:01 had high diagnostic accuracy ($0.9 < AUC < 1$) for detection of allopurinol-induced CADR. The DOR combines sensitivity and specificity as one indicator for diagnostic accuracy [34]. In overall analysis, HLA-B*58:01 also showed a high diagnostic performance with a DOR of 90.12. The likelihood ratios are more

clinically meaningful indicators [35]. The summary PLR was 8.24 suggesting about 8.3 times higher chance of a allopurinol-induced CADR case to be identified from a positive result for HLA-B*58:01 testing, which was not high enough to be used as a robust diagnostic indicator of allopurinol-induced CADR. While a negative result for HLA-B*58:01 screening means allopurinol-tolerant control only have 8.6% of probability to develop CADR. These values indicated that a negative result of HLA-B*58:01 allele could be used as a justification to deny allopurinol-induced CADR.

As lacks of sufficient evidence about the cost-effectiveness of HLA-B*58:01 typing, and the conflicting results reported, screening HLA-B*58:01 before allopurinol administration is still a remaining issue. Recently, Jung et al. [6] reported that genetic screening test could help to identify HLA-B*58:01-positive patients and thus improve safety of allopurinol treatment. More recently, a large cohort study further demonstrated the usefulness of prospective HLA-B*58:01 genotyping

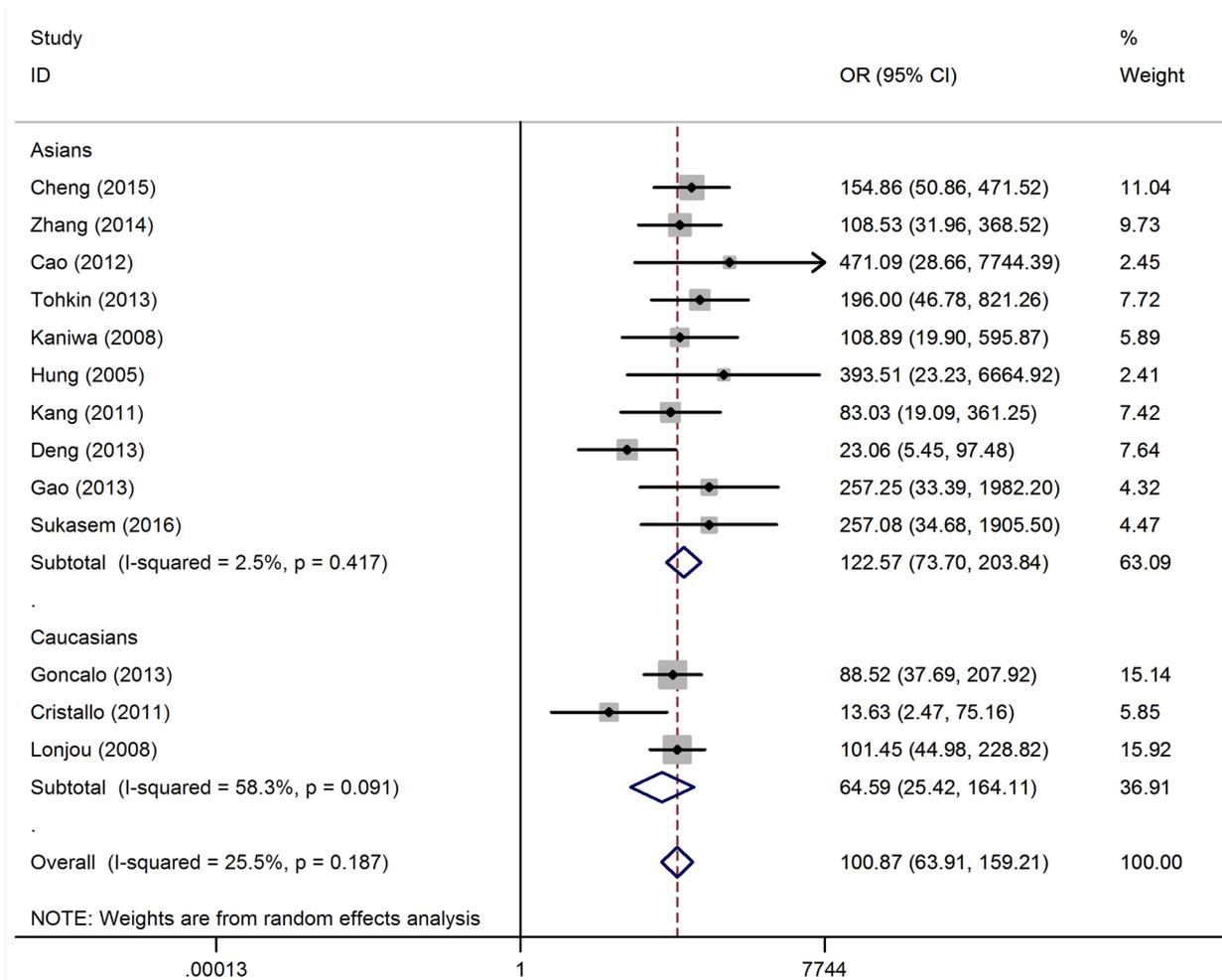


Figure 2: Forest plot for the meta-analysis of the association between HLA-B*58:01 allele carriers and risk of allopurinol-induced CADR stratified by ethnicity in population based study.

for prevention of SCARs induced by allopurinol [36]. Furthermore, two cost-effectiveness studies conducted among Koreans and Thais indicated that HLA-B*58:01 screening before allopurinol treatment is a more cost effective intervention than benzbromarone or febuxostat as an alternative medication [37, 38]. However, Dong et al. reported that allopurinol treatment for chronic gout without HLA-B*58:01 genetic tests remain the optimal strategy from a cost-effectiveness perspective in Singapore [39]. With the development of new technologies and decreasing

cost of genetic testing, HLA screening could implement widely and cost effectively in near future.

The detailed pathogenesis mechanisms of CADR caused by allopurinol remain unknown. Accumulated evidence suggested that T-cell-mediated immunologic response play a central role in CADR. Through interactions with class I HLA-restricted antigen-presenting cells (APC) and T-cell receptor, CD8-positive cytotoxic T cells believed to trigger an immunologic reaction of SCARs [40]. *In vitro* study by Lin et al. demonstrates

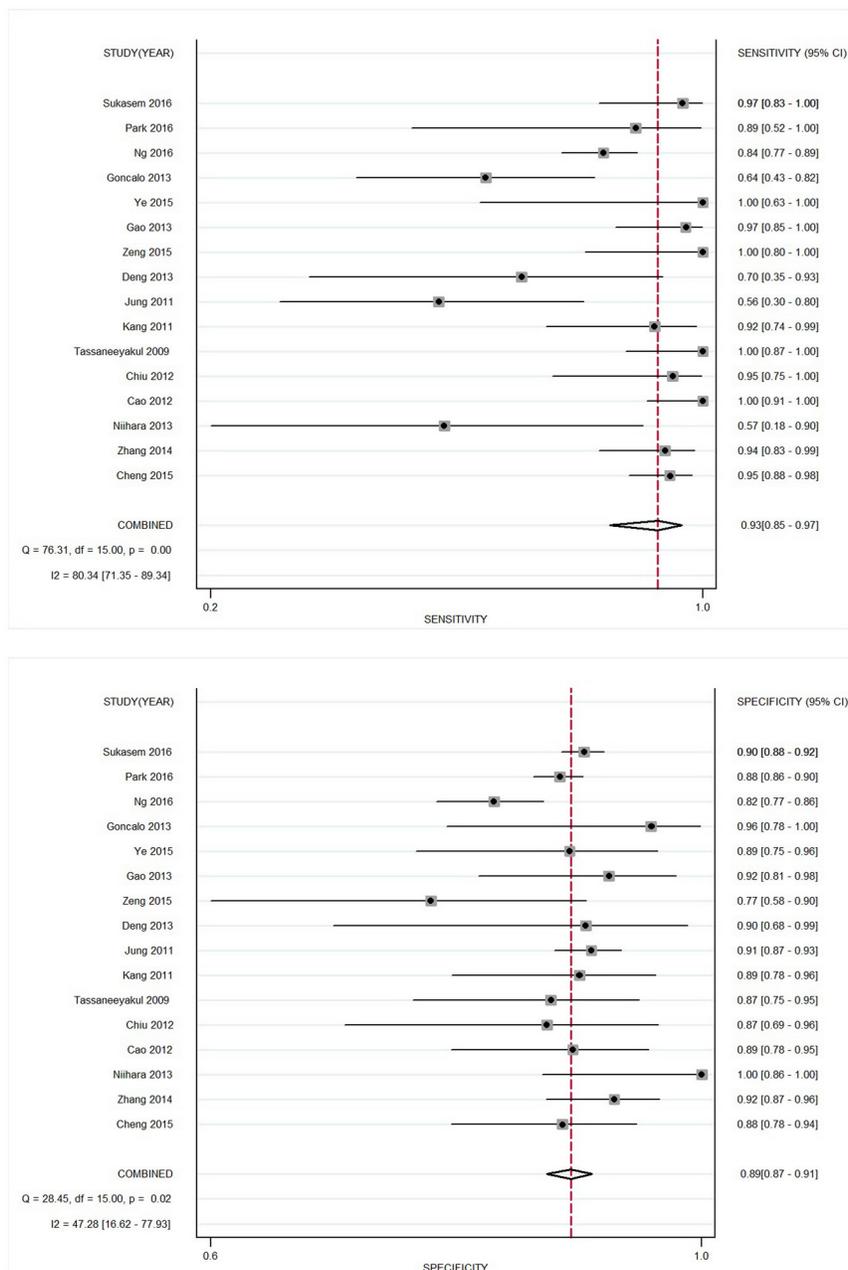


Figure 3: Forest plot of estimates of sensitivity and specificity for HLA-B*58:01 status in the diagnosis of allopurinol-induced CADR.

that only APC with HLA-B*58:01 allele present strong cytotoxicity against CD8 + T lymphocytes collected from patients with allopurinol–caused CADR [41].

Some limitations of the study must be addressed to prevent misinterpretation of our findings. First, substantial heterogeneity was detected and study-level data did not allow us to further explore potential sources of heterogeneity. Second, most cases of allopurinol-induced CADR were of Asian origin, and results from Caucasian populations may be biased. Third, only single-factor estimates were available and we failed to provide results with further adjustment of potential confounders. For stratified analyses investigating HLA-B*58:01 with MPE and EEM, very few patients were available. Thus, selection bias is inevitable and the results may be easily over inflated. Large studies are needed to address these issues.

Taken together, our meta-analysis from 21 pharmacogenetic studies summarizes the strong correlation of allopurinol-related CADR with HLA-B*58:01 allele, especially among Asians. Our findings suggested that screening for HLA-B*58:01 may be helpful in allopurinol-induced CADR detection because of its high level of diagnostic accuracy.

MATERIALS AND METHODS

Data sources and search strategy

Pharmacogenetic association studies published before July 2016, on HLA-B*58:01 and CADR in patients treated with allopurinol were sought by computer-based searches, scanning of the reference lists of all relevant studies and review articles, hand searching of relevant journals. Systematic search of the literature in EMBASE, PubMed, clinicaltrials.gov, The Cochrane Library, Web of Knowledge, MEDLINE, IPA (International Pharmaceutical Abstracts), CINAHL (Cumulative Index to Nursing and Allied Health Literature), and HuGENet (Human Genome Epidemiology Network) used keywords relating to the HLA-B (e.g., “Human leukocyte antigen”) and allopurinol in combination with CADR (e.g., “drug adverse reaction”, “maculopapular exanthema”, “hypersensitivity syndrome”, “Stevens Johnson syndrome”, “toxic epidermal necrolysis”, and “erythema exudativum multiforme”). No restriction was imposed on the language and the year of publication. Furthermore, citations in the retrieved articles were hand-searched to identify additional relevant reports. The titles and abstracts

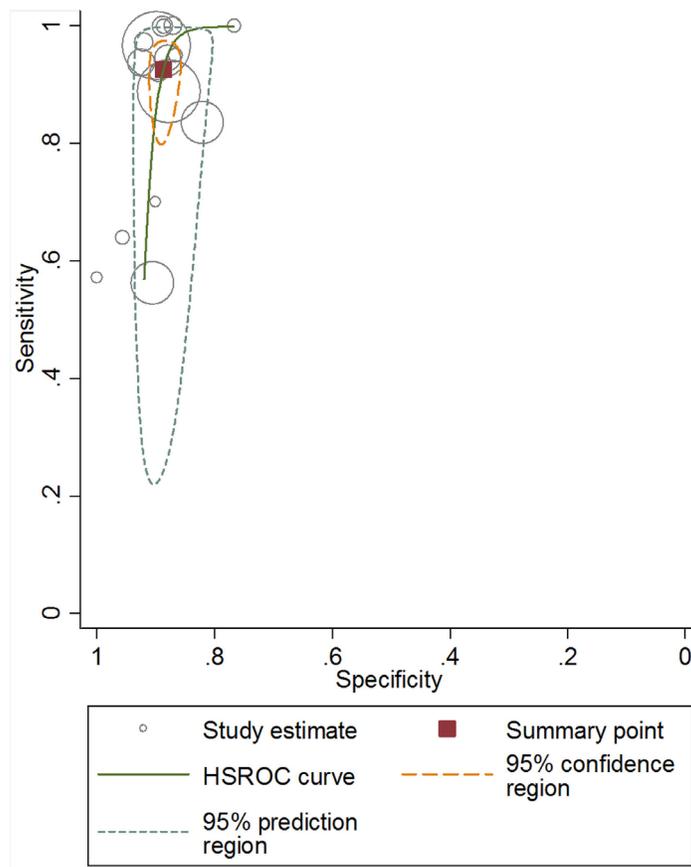


Figure 4: Hierarchical summarized receiver operating characteristic (HSROC) curves of HLA-B*58:01 status for allopurinol-induced CADR diagnosis.

were read to determine their relevance, and potentially relevant studies were retained for further evaluation. For retrieved articles, the full texts of the articles were read to determine whether they contained information on the topic of interest.

Selection criteria and quality assessment

Two investigators (R.W and Y.J.C) independently assessed abstracts and titles retrieved from the comprehensive searches for eligible study. Studies included in the current meta-analysis had to meet all the following criteria: (a) original papers containing independent data, (b) investigated the relationship between HLA-B*58:01 and allopurinol-induced CADR, (c) reported sufficient data for estimating an odds ratio (OR) with 95% confidence interval (95% CI), sensitivity and specificity. Exclusion criteria were as follows: (a) case-only studies, (b) duplicated studies using the same case series, and (c) reviews, editorials, comments, reports from scientific sessions or discussions. A procedure known as ‘Newcastle–Ottawa Scale (NOS)’ has been used to assess the quality of included observational studies. Details are published elsewhere [42].

Data extraction

Information was carefully extracted from all eligible publications independently by the two reviewers according to a fixed protocol. First author, study design, year of publication, ethnicity, eligibility criteria, diagnosis and phenotypic definition for CADR patient demographics, CADR type, dosage of allopurinol and duration of use, genotyping method, HLA-B*58:01 status among cases and controls, results of Hardy-Weinberg equilibrium (HWE) in the control group, and sensitivity and specificity data were collected from eligible studies. Review reports from the two were then compared to identify any inconsistency, and differences were resolved by further discussion among all authors.

Statistical analysis

The data from each study was divided into two groups according to study design: allopurinol-induced CADR vs. allopurinol-tolerant patients; allopurinol-induced CADR vs. healthy controls without allopurinol exposure or subjects obtained from the population database. The strength of the association between the presence of HLA-B*58:01 in at least 1 allele [43] and allopurinol-induced CADR was estimated using crude odds ratios (ORs), with the corresponding 95% confidence intervals (CIs). The heterogeneity across individual studies was measured by the Cochran’s Q test and the inconsistency index (I^2) [44]. A random-effects model, which is usually more conservative, was used to calculate the pooled ORs [45]. Quantitative assessment of sources

of heterogeneity was undertaken by meta-regression analysis using ethnicity, sample size, source of controls, age, sex, allopurinol dosage and exposure duration as covariates [46]. Subgroup analyses by ethnicity (Asians, and Caucasians) and clinical outcomes (SCARs, SJS/TEN, MPE, and EEM) were also performed to seek for potential sources of between-study heterogeneity. Furthermore, the Galbraith plot was used to identify the outliers contributing toward heterogeneity.

Sensitivity, specificity, diagnostic odds ratio (DOR), positive likelihood ratio (PLR), and negative likelihood ratio (NLR) with corresponding 95% confidence intervals (CI) were calculated for each matched study. Meta-analysis of diagnostic test evaluations was performed using standard methods under random-effects model [47]. Hierarchical summary receiver operating characteristic (HSROC) curves were also plotted to graphically present the results [48]. The area under the curve (AUC) results are considered excellent for AUC values of 0.9 – 1.0, good for values of 0.8 – 0.9, fair for values of 0.7 – 0.8, and poor for values of 0.6 – 0.7.

To assess the stability of results, one-way sensitivity analyses were performed by removing each individual study in turn from the total and reanalysing the remainder. Small study effects, such as publication bias, were assessed by inspecting the funnel plots for asymmetry and Egger’s linear regression test, as well as Deeks’ test [49–51]. Since the P-values of less than 0.05 were considered significant, alpha was firstly set at 0.05, and the Holm-Bonferroni method was used to control the type I error in multiple comparisons with an alpha of 0.0056 (0.05/9). Statistical analyses were carried out using the STATA software version 11.0 (Stata Corporation, College Station, TX, USA).

CONFLICTS OF INTEREST

The authors have no conflicts of interest to declare.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

REFERENCES

1. Terkeltaub RA. Clinical practice. Gout. *N Engl J Med*. 2003; 349: 1647–1655.
2. Roujeau JC. Clinical heterogeneity of drug hypersensitivity. *Toxicology*. 2005; 209:123–129.
3. Finkelstein Y, Macdonald EM, Li P, Hutson JR, Juurlink DN. Recurrence and mortality following severe cutaneous adverse reactions. *JAMA*. 2014; 311: 2231–2232.
4. Roujeau JC, Stern RS. Severe adverse cutaneous reactions to drugs. *N Engl J Med*. 1994; 331: 1272–1285.

5. Yu Q, Huang JF. The DEER database: a bridge connecting drugs, environmental effects, and regulations. *Gene*. 2013;520:98–105.
6. Jung JW, Kim DK, Park HW, Oh KH, Joo KW, Kim YS, Ahn C, Lee KW, Cho SH, Min KU, Kang HR. An effective strategy to prevent allopurinol-induced hypersensitivity by HLA typing. *Genet Med*. 2015; 17:807–814.
7. Hung SI, Chung WH, Liou LB, Chu CC, Lin M, Huang HP, Lin YL, Lan JL, Yang LC, Hong HS, Chen MJ, Lai PC, Wu MS, Chu CY, Wang KH, Chen CH, Fann CS, Wu JY, Chen YT. HLA-B*5801 allele as a genetic marker for severe cutaneous adverse reactions caused by allopurinol. *Proc Natl Acad Sci U S A*. 2005;102:4134–4139.
8. Tohkin M, Kaniwa N, Saito Y, Sugiyama E, Kurose K, Nishikawa J, Hasegawa R, Aihara M, Matsunaga K, Abe M, Furuya H, Takahashi Y, Ikeda H, et al. A whole-genome association study of major determinants for allopurinol-related Stevens-Johnson syndrome and toxic epidermal necrolysis in Japanese patients. *Pharmacogenomics J*. 2013;13:60–69.
9. Cheng L, Xiong Y, Qin CZ, Zhang W, Chen XP, Li J, Zhou HH. HLA-B*58:01 is strongly associated with allopurinol-induced severe cutaneous adverse reactions in Han Chinese patients: a multicentre retrospective case-control clinical study. *Br J Dermatol*. 2015; 173:555–558.
10. Zhang X, Ma H, Hu C, Yu B, Ma W, Wu Z, Luo X, Zou H, Guan M. Detection of HLA-B*58:01 with TaqMan assay and its association with allopurinol-induced sCADR. *Clin Chem Lab Med*. 2015;53:383–390.
11. Zineh I, Mummaneni P, Lyndly J, Amur S, La Grenade LA, Chang SH, Rogers H, Pacanowski MA. Allopurinol pharmacogenetics: assessment of potential clinical usefulness. *Pharmacogenomics*. 2011;12:1741–1749.
12. Ng CY, Yeh YT, Wang CW, Hung SI, Yang CH, Chang YC, Chang WC, Lin YJ, Chang CJ, Su SC, Fan WL, Chen DY, Wu YJ, et al. Impact of the HLA-B(*)58:01 Allele and Renal Impairment on Allopurinol-Induced Cutaneous Adverse Reactions. *J Invest Dermatol*. 2016;136:1373–1381.
13. Park HJ, Kim YJ, Kim DH, Kim J, Park KH, Park JW, Lee JH. HLA Allele Frequencies in 5802 Koreans: Varied Allele Types Associated with SJS/TEN According to Culprit Drugs. *Yonsei Med J*. 2016;57:118–126.
14. Ye XL, Chen CS, Xu RA, Zhang CH, Pan XF, Zhang XH. HLA-B*5801 carrying station and the correlation of allergic reactions caused by allopurinol. *J Wenzhou Medical University*. 2015;45:143–45.
15. Zeng DY, Wang CL, Huang PF, Liu YW, Chen DD. Research on detection methods for HLA-B*5801: a biomarker for allopurinol induced serious cutaneous adverse drug reactions. *Chin J Mod Appl Pharm*. 2015;32:700–704.
16. Gonçalo M, Coutinho I, Teixeira V, Gameiro AR, Brites MM, Nunes R, Martinho A. HLA-B*58:01 is a risk factor for allopurinol-induced DRESS and Stevens-Johnson syndrome/toxic epidermal necrolysis in a Portuguese population. *Br J Dermatol*. 2013;169:660–665.
17. Gao J, Zhang JJ, Wang J, Zhang WW, Yu D, Qian YL, Zheng XX, Ding XL, Miao LY. Correlation between HLA-B*5801 allele and allopurinol-induced severe cutaneous adverse reaction in Han ethnic group patients in Jiangsu province. *Adverse Drug Reactions J*. 2013;15:258–262.
18. Deng ZG, Yang J, Yang WL. Detection of the HLA-B*5801 allele in Han Chinese with allopurinol-induced severe drug eruption. *J Diagn Ther Dermatol-Venereol*. 2013; 20:379–382.
19. Niihara H, Kaneko S, Ito T, Sugamori T, Takahashi N, Kohno K, Morita E. HLA-B*58:01 strongly associates with allopurinol-induced adverse drug reactions in a Japanese sample population. *J Dermatol Sci*. 2013;71:150–152.
20. Chiu ML, Hu M, Ng MH, Yeung CK, Chan JC, Chang MM, Cheng SH, Li L, Tomlinson B. Association between HLA-B*58:01 allele and severe cutaneous adverse reactions with allopurinol in Han Chinese in Hong Kong. *Br J Dermatol*. 2012;167:44–49.
21. Cao ZH, Wei ZY, Zhu QY, Zhang JY, Yang L, Qin SY, Shao LY, Zhang YT, Xuan JK, Li QL, Xu JH, Xu F, Ma L, et al. HLA-B*58:01 allele is associated with augmented risk for both mild and severe cutaneous adverse reactions induced by allopurinol in Han Chinese. *Pharmacogenomics*. 2012;13:1193–1201.
22. Kang HR, Jee YK, Kim YS, Lee CH, Jung JW, Kim SH, Park HW, Chang YS, Jang IJ, Cho SH, Min KU, Kim SH, Lee KW; Adverse Drug Reaction Research Group in Korea. Positive and negative associations of HLA class I alleles with allopurinol-induced SCARs in Koreans. *Pharmacogenet Genomics*. 2011;21:303–307.
23. Jung JW, Song WJ, Kim YS, Joo KW, Lee KW, Kim SH, Park HW, Chang YS, Cho SH, Min KU, Kang HR. HLA-B58 can help the clinical decision on starting allopurinol in patients with chronic renal insufficiency. *Nephrol Dial Transplant*. 2011;26:3567–3572.
24. Cristallo AF, Schroeder J, Citterio A, Santori G, Ferrioli GM, Rossi U, Bertani G, Cassano S, Gottardi P, Ceschini N, Barocci F, Ribizzi G, Cutrupi V, et al. A study of HLA class I and class II 4-digit allele level in Stevens-Johnson syndrome and toxic epidermal necrolysis. *Int J Immunogenet*. 2011;38:303–309.
25. Tassaneeyakul W, Jantararoungtong T, Chen P, Lin PY, Tiamkao S, Khunarkornsiri U, Chucherd P, Konyoung P, Vannaprasaht S, Choonhakarn C, Pisuttimarn P, Sangviroon A, Tassaneeyakul W. Strong association between HLA-B*5801 and allopurinol-induced Stevens-Johnson syndrome and toxic epidermal necrolysis in a Thai population. *Pharmacogenet Genomics*. 2009;19:704–709.
26. Kaniwa N, Saito Y, Aihara M, Matsunaga K, Tohkin M, Kurose K, Sawada J, Furuya H, Takahashi Y, Muramatsu M, Kinoshita S, Abe M, Ikeda H, et al. HLA-B locus in Japanese patients with anti-epileptics and allopurinol-related

- Stevens-Johnson syndrome and toxic epidermal necrolysis. *Pharmacogenomics*. 2008;9:1617–1622.
27. Lonjou C, Borot N, Sekula P, Ledger N, Thomas L, Halevy S, Naldi L, Bouwes-Bavinck JN, Sidoroff A, de Toma C, Schumacher M, Roujeau JC, Hovnanian A, et al. A European study of HLA-B in Stevens-Johnson syndrome and toxic epidermal necrolysis related to five high-risk drugs. *Pharmacogenet Genomics*. 2008;18:99–107.
 28. Sukasem C, Jantararoungtong T, Kuntawong P, Puangpetch A, Koomdee N, Satapornpong P, Supapsophon P, Klaewsongkram J, Rerkpattanapipat T. HLA-B (*) 58:01 for Allopurinol-Induced Cutaneous Adverse Drug Reactions: Implication for Clinical Interpretation in Thailand. *Front Pharmacol*. 2016;7:186.
 29. Roujeau JC, Kelly JP, Naldi L, Rzany B, Stern RS, Anderson T, Auquier A, Bastuji-Garin S, Correia O, Locati F, Mockenhaupt M, Paoletti C, Shapiro S, et al. Medication use and the risk of Stevens–Johnson syndrome or toxic epidermal necrolysis. *N Engl J Med*. 1995; 333:1600–1607.
 30. Kardaun SH, Sidoroff A, Valeyrie-Allanore L, Halevy S, Davidovici BB, Mockenhaupt M, Roujeau JC. Variability in the clinical pattern of cutaneous side-effects of drugs with systemic symptoms: does a DRESS syndrome really exist? *Br J Dermatol*. 2007;156:609–611.
 31. Munafò MR, Flint J. Meta-analysis of genetic association studies. *Trends Genet*. 2004; 20: 439–444.
 32. Hershfield MS, Callaghan JT, Tassaneeyakul W, Mushiroda T, Thorn CF, Klein TE, Lee MT. Clinical pharmacogenetics implementation consortium guidelines for human leukocyte antigen-B genotype and allopurinol dosing. *Clin Pharmacol Ther*. 2013; 93:153–158.
 33. Swets JA. Measuring the accuracy of diagnostic systems. *Science*. 1988;240:1285–1293.
 34. Glas AS, Lijmer JG, Prins MH, Bossel GJ, Bossuyt PM. The diagnostic odds ratio: a single indicator of test performance. *J Clin Epidemiol*. 2003;56:1129–1135.
 35. Gallagher EJ. Clinical utility of likelihood ratios. *Ann Emergency Med*. 1998;31:391–397.
 36. Ko TM, Tsai CY, Chen SY, Chen KS, Yu KH, Chu CS, Huang CM, Wang CR, Weng CT, Yu CL, Hsieh SC, Tsai JC, Lai WT, et al. Use of HLA-B*58:01 genotyping to prevent allopurinol induced severe cutaneous adverse reactions in Taiwan: national prospective cohort study. *BMJ*. 2015;351:h4848.
 37. Saokaew S, Tassaneeyakul W, Maenthaisong R, Chaiyakunapruk N. Cost-effectiveness analysis of HLA-B*5801 testing in preventing allopurinol-induced SJS/TEN in Thai population. *PLoS One*. 2014;9:e94294.
 38. Park DJ, Kang JH, Lee JW, Lee KE, Wen L, Kim TJ, Park YW, Park SH, Lee SS. Cost-effectiveness analysis of HLA-B5801 genotyping in the treatment of gout patients with chronic renal insufficiency in Korea. *Arthritis Care Res (Hoboken)*. 2015;67:280–287.
 39. Dong D, Tan-Koi WC, Teng GG, Finkelstein E, Sung C. Cost-effectiveness analysis of genotyping for HLA-B*5801 and an enhanced safety program in gout patients starting allopurinol in Singapore. *Pharmacogenomics*. 2015;16:1781–1793.
 40. Nassif A, Bensussan A, Boumsell L, Deniaud A, Moslehi H, Wolkenstein P, Bagot M, Roujeau JC. Toxic epidermal necrolysis: effector cells are drug-specific cytotoxic T cells. *J Allergy Clin Immunol*. 2004;114:1209–1215.
 41. Lin C, Chen Y, Hung S. The pathogenic role of HLA-B*5801 in allopurinol-induced severe cutaneous adverse reactions. Presented at: Drug Hypersensitivity Meeting – DHM5, Munich, 2012, Programme and Abstract Book, 2012; 125.
 42. Stang A. Critical evaluation of the Newcastle–Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. *Eur J Epidemiol*. 2010; 25:603–605.
 43. Tanaka H, Akaza T, Juji T. Report of the Japanese Central Bone Marrow Data Center. *Clin Transpl*. 1996; 139–144.
 44. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med*. 2002;21:1539–1558.
 45. DerSimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials*. 1986; 7:177–188.
 46. Thompson SG, Higgins JP. How should meta-regression analyses be undertaken and interpreted? *Stat Med*. 2002; 21:1559–1573.
 47. Deville WL, Buntinx F, Bouter LM, Montori VM, de Vet HC, van der Windt DA, Bezemer PD. Conducting systematic reviews of diagnostic studies: didactic guidelines. *BMC Med Res Methodol*. 2002;2:9.
 48. Rutter CM, Gatsonis CA. A hierarchical regression approach to meta-analysis of diagnostic test accuracy evaluations. *Stat Med*. 2001;20:2865–2884.
 49. Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics*. 1994;50:1088–1101.
 50. Egger M, Davey Smith G, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315:629–634.
 51. Deeks JJ, Macaskill P, Irwig L. The performance of tests of publication bias and other sample size effects in systematic reviews of diagnostic test accuracy was assessed. *J Clin Epidemiol*. 2005;58:882–893.