Editorial Material

IκBα beyond the NF-kB dogma

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The NF- κ B pathway participates in a myriad of processes including cell differentiation, proliferation, survival, and regulation of the immune response. In mammals there are five NF- κ B proteins: p65/RelA, p50, RelB, cRel and p52. All these proteins can form dimers to generate more than a dozen of possible combinations. Different NF- κ B dimers bind the DNA at specific κ B sites, acting as activators or repressors of gene transcription. However, this is only an example of how complex the NF- κ B pathway is, since it also involves several post-translational modifications of upstream and downstream elements and multiple negative and positive regulatory loops.

I κ B proteins, including I κ B α , I κ B β and I κ B ϵ constitute the canonical inhibitors of the NF- κ B pathway. However p105 and p100 proteins, the precursors of p50 and p52, contain a C-terminal end that is functionally comparable to I κ B (this is the reason they are also called I κ B γ and I κ B δ , respectively).

In unstimulated cells, NF- κ B dimers reside in the cytoplasm bound to I κ B proteins. Upon stimulation, I κ Bs become phosphorylated, subsequently polyubiquitinated and degraded by the proteasome. Degradation of I κ B leads to the release of the NF- κ B factors that then translocate to the nucleus to activate transcription of specific genes including I κ B α itself. Termination of the transcriptional response primarily depends on I κ B α resynthesis that is required not only for retaining NF- κ B in the cytoplasm but also for removing active NF- κ B dimers from the DNA [1]. Because this mechanism implies that I κ B α continuously shuttles from the cytoplasm to the nucleus to maintain the cells in a silent NF- κ B state [2], it is not so surprising that I κ B α has been found associated with other proteins that are primarily nuclear such as histone

deacetylases (HDACs) or nuclear corepressors [3, 4]. Now, we have demonstrated that $I\kappa B\alpha$ is predominantly nuclear in primary keratinocytes where it binds the chromatin at specific genes promoters associated with their transcriptional repression [5]. Mechanistically, we found that $I\kappa B\alpha$ facilitates the association of the Polycomb Repressive Complex 2 (PRC2) to specific promoters and supply PRC2 with the capacity to respond to TNFa stimulation, thus establishing an unexpected link between inflammatory signals and skin homeostasis. This result is particularly relevant since to date it is still not clear how PRC2 is recruited to the chromatin in mammals, unlike in flies where specific PRC2 recognition DNA sequences have been identified. The physiologic relevance of IkB and Polycomb (Pc) association has been validated in Drosophila using compose mutants of Cactus, the homolog of IkBa in flies, and Pc, which in fact indicates the elevated conservation of this function during evolution.

The identification of $I\kappa B\alpha$ as a chromatin interacting protein apparently contradicts the notion that $I\kappa B\alpha$ cannot bind the DNA. However, it is not just $I\kappa B\alpha$ but a SUMOylated and phosphorylated form of the protein that binds the chromatin, suggestive of structural changes that affect the biochemistry of the protein including its capacity to bind NF- κ B or the chromatin in a mutually exclusive manner. Another member of the family, $I\kappa B\beta$, also binds the chromatin but only when it is hypophosphorylated [6], suggesting that post-translational modifications of the NF- κ B elements link this pathway with specific cellular functions. This is also the case of IKK α and NEMO that exert additional NF- κ B-related but independent functions such as cell cycle control and DNA-damage repair, respectively [7, 8]. However, which are the modifications



🜔 Cytoplasmic ΙκΒα

Nuclear ΙκΒα

- Cytoplasmic ΙκΒα

Figure 1: Cytoplasmic localization of I κ B α in the skin is associated with tumorigenesis. I κ B α is localized in the cytoplasm of most cell types where it plays a major role as a negative regulator of the NF- κ B signaling pathway. However, I κ B α is detected in the nucleus of basal layer keratinocytes, whereas loss of nuclear I κ B α and its cytoplasmic accumulation is a hallmark of human Squamous Cell Carcinoma.

that regulate alternative IKK α and NEMO functions is not completely understood.

Detection of cytoplasmic IkBa has never been considered as pathologic in any tissue, however this seems to be the case in the skin (Fig. 1). In Mulero et al we found that nuclear exclusion and cytoplasmic accumulation of IkBa strongly associated with progression of Squamous Cell Carcinoma (SCC) in patient samples. Although this is not addressed in the study, it is plausible that this is not cytoplasmic accumulation of conventional IkBa but of SUMOylated IkBa what exerts pro-tumorigenic functions, likely by relieving the nucleus from transcriptional regulators. Further studies should be performed to validate this idea. Why this is observed in the skin but not in other tissues can be basically a matter of levels of either phosphatases and/or desumoylases, which will be deciphered in the near future. However, this does not exclude that low amounts of SUMOylated IkBa exert comparable functions in cells other than the keratinocytes, which will be also investigated.

In conclusion, the identification of SUMOylated and phosphorylated $I\kappa B\alpha$ as a spinoff of $I\kappa B\alpha$ that plays unexpected but important physiologic function opens a whole field of research and the possibility to re-interpret some old unexpected results with a novel and renewed perspective.

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REFERENCES

- Hoffmann A et al. Immunological Reviews. 2006; 210: 171-186.
- 2. Arenzana-seisdedos F et al. J Cell Sci. 1997; 110: 369-78.
- 3. Viatour P et al. J Biol Chem. 2003; 278:46541-46548.
- Aguilera C et al. Proceedings of the National Acad. Sci. USA. 2004; 101(47): 16537-42.
- 5. Mulero MC et al. Cancer Cell. 2013; 24(2): 151-166.
- 6. Rao P et al. Nature. 2010; 466(7310): 1115-9.
- 7. Zhu F et al. Mol Cell 2007; 27: 214-227.
- 8. Huang TT et al. Cell. 2003 26;115(5):565-76.