Research Paper

Dapper homolog 1 alpha suppresses metastasis ability of gastric cancer through inhibiting planar cell polarity pathway

Yuegeng Liu¹, Jingwan Zhang¹, Weifang Yu², Xiaoming Zhang¹, Guiqi Wang¹, Zengren Zhao¹

¹Departments of Surgery, First Affiliated Hospital, Hebei Medical University, Shijiazhuang, China

²Endoscopy Center, First Affiliated Hospital, Hebei Medical University, Shijiazhuang, China

Correspondence to: Zengren Zhao, email: zzr-doctor@163.com

Keywords: DACT1a, gastric cancer, metastasis, tumor suppressor

Received: September 20, 2016 Accepted: October 17, 2016

Published: November 09, 2016

ABSTRACT

Dapper homolog 1 alpha (DACT1 α) is a member of DACT family and an important regulator in the planar cell polarity pathway. We aim to clarify its functional role in metastasis ability of gastric cancer. DACT1a was silenced in all gastric cancer cell lines (8/8), but expressed in normal gastric tissue. Ectopic expression of DACT1a in silenced gastric cancer cell lines (AGS, BGC823 and MGC803) by stable transfection significantly suppressed cancer cell spreading (P < 0.05), migration (P < 0.01) and invasion (P < 0.01). These effects were associated with downregulation of planar cell polarity pathway related genes involved in cell proliferation (PDGFB, VEGFA), adhesion (ITGA1, ITGA2, ITGA3, ITGB3) and migration/invasion (PLAU, MMP9, MCAM, DvI-2 and JNK). DACT1a promoter methylation was detected in 205 gastric cancers and 20 normal controls by direct bisulfite genomic sequencing. DACT1a methylation was detected in 29.3% (60/205) of gastric cancer patients, but not in normal tissues. DACT1g methylation was associated with poor survival of gastric cancer patients. In conclusion, DACT1g plays a pivotal role as a potential tumor suppressor in migration and invasion of gastric cancer. DACT10 methylation may serve as a biomarker for the prognosis of gastric cancer.

INTRODUCTION

Gastric cancer is the fourth most common cancer worldwide with 930,000 newly diagnosed cases in 2002. Gastric cancer is the third most common cause of cancer-related death in the world [1]. Since the lack of early symptoms often delays the diagnosis, 80% to 90% of patients with gastric cancer present with locally advanced or metastatic tumors that have poor survival rates [1]. Similar to other types of cancer, gastric cancer develops via a multistage process, including DNA methylation of CpG islands, post-translational modification of histones, microRNAs and nucleosome positioning [2–4]. Among them, promoter methylation is considered as one of the major mechanisms to inactivate tumor suppressor genes, thereby causing gastric carcinogenesis [5]. Promoter methylation is considered as an important hallmark of the progression and metastasis of gastric cancer. So far, many methylated genes have been identified play important roles in cell proliferation, cell cycle, apoptosis, angiogenesis, metastasis, thus contributing to the development of gastric cancer [6-10].

Dapper homolog 1 alpha (DACT1α), a member of DACT family, is an important regulator in the planar cell polarity (PCP) pathway which is an important pathway in cancer metastasis [11, 12]. It has been reported that DACT1a played an important role in regulating PCP pathway by post-translationally regulating central PCP component Dvl2 and vang-like 2 (Vangl2) and modulating PCP downstream Rac1/JNK cascade [11, 12]. Subsequent investigations have revealed that DACT1 α is involved in the pathogenesis of multiple forms of malignant cancers in humans including gastric cancer [13, 14]. Hypermethylation of the DACT1 DNA promoter was frequently identified to play a regulator of tumorigenesis and prognosis in gastric cancer [13, 14]. However, the functional significance of DACT1a in regulation cell metastasis ability is still largely unclear. In this study, we mainly aimed to study the metastasis ability of DACT1 α in gastric cancer.

RESULTS

Silence or down-regulation of DACT1α in gastric cancer cell lines

DACT1 α was expressed in normal human gastric epithelial cell GES1 and in normal stomach tissue, but down-regulated or silenced in gastric cancer cell lines both at mRNA and protein level (Figure 1A).

DACT1a inhibits of gastric cancer cell spreading

The frequent silence of DACT1 α in gastric cancer cell lines but not in normal gastric mucosa suggests that DACT1 α may act as a tumor suppressor. In this connection, we examined the effect of ectopic expression of DACT1 α in gastric cancer cell lines. Three gastric cancer cell lines (AGS, BGC823 and MGC803) with different differentiation and characteristics were selected for functional evaluation of DACT1 *in vitro*. AGS is a well-differentiated gastric epithelial *cell line; BGC823* is an undifferentiated gastric epithelial cell line and MGC803 is a poorly differentiated mucoid stomach adenocarcinoma cell line. Re-expression of DACD1 in the stably transfected BGC823, MGC803 and AGS cells was confirmed by western blot (Figure 1B1). Ectopic expressing DACT1 α in silenced gastric cancer cell lines BGC823 and MGC803 cell lines significantly inhibited in cell spreading in comparison with control cells by stably transfectants with DACT1 α in BGC823 and MGC803 cell lines (BGC823, P = 0.003; MGC803, P = 0.001) (Figure 1C). DACT1 α also down-regulated actin microfilament (stress fiber) formation as determined by staining with rhodamine-labeled phalloidin (Figure 1D).

DACT1α suppresses gastric cancer cell migration

In vitro wound healing assay showed that ectopic expression of DACT1 α markedly slowed cell migration at the edges of scratch wound of AGS and MGC803 (Figure 2A1), while an inverse effect was observed in GES1 with knockdown of DACT1 α (Figure 2A2). Quantitative analyses at 24 h confirmed a significant reduction in wound closure in DACT1 α -transfected AGS cells (P < 0.01) and MGC803 (P < 0.05) compared with empty vector-transfected control cells (Figure 2A1). On the other hand knockdown DACT1 α in normal gastric



Figure 1: DACT1α suppressed cell spreading and F-actin formation. (A) DACT1α expression was silenced in all gastric cancer cell lines, but expressed in normal human epithelial cell line GES1 and in normal gastric tissues. (B1) Ectopic expression of DACT1α in BGC803, MGC823 and AGS cell lines was confirmed by Western blot (cropped gel). (B2) Knockdown effect of DACT1α by shDACT1α in normal gastric epithelial cell line GES1 was confirmed by Western blot (cropped gel). (C) pcDNA3.1-DACT1α vector stably transfected BGC823 and MGC803 cells were significantly slowed the spreading rate on cover slips for 6 hours compared to control vector pcDNA3.1 transfected and BGC823 and MGC803 cells. (D) BGC823 and MGC803 cells stained with rhodamine-phalloidin for detecting F-actin formation.

epithelial cell line GES1 was confirmed by Western blot (Figure 1B2). Knockdown DACT1 α in GES1 cells significantly promoted wound closure rate compared with the control cells (P < 0.01) (Figure 2A2).

DACT1a suppresses gastric cancer cell invasion

We further evaluated the effect of DACT1 α in cell invasion. We found that ectopic expression of DACT1 α significantly dampened cell invasion ability as measured by transwell assays in AGS ((P < 0.01) and MGC803 (P < 0.01) cells stably transfected pcDNA3.1-DACT1 α compared to stably transfected control vector, respectively (Figure 2B1). Inversely, knockdown DACT1 α induced GES1 cell invasive ability (Figure 2B2).

DACT1α regulates planar cell polarity (PCP) pathway

To examine whether DACT1 α regulated PCP pathway in gastric cancer cells [11, 12, 15], expression of central PCP component Dvl-2 and activation of PCP

downstream JNK pathway were determined by western blot in pcDNA3.1 and pcDNA3.1-DACT1 α stably transfected BGC823, MGC803 and AGS cells. As shown in Figure 3A, DACT1 α regulated PCP pathway by promoting Dvl-2 degradation and suppressing the active form of JNK in GC cells.

Downstream target cancer-related genes regulated by DACT1α

To gain insight into the molecular mechanisms underlying the tumor suppression of DACT1 α , gene expression profile in DACT1 α stably transfected BGC823 were analyzed by Cancer Pathway cDNA microarray and further validated by real-time PCR (Figure 3B). Two representative genes PDGFB and MMP9 which included in proliferation and migration pathways were further verified by western blot (Figure 3C). Compared to control vector transfected cells, the anti-tumorigenesis effect by DACT1 α was mediated by regulating important genes in cell proliferation, angiogenesis, adhesion, migration and invasion (Table 1). DACT1 α led to down-regulation of





angiogenic platelet-derived growth factor beta polypeptide (PDGFB), vascular endothelial growth factor A (VEGFA) as well as multiple cell migration and invasion molecules integrin, alpha 1 (ITGA1), integrin, alpha 2 (ITGA2), integrin, alpha 3 (ITGA3), integrin, beta 3 (ITGB3), melanoma cell adhesion molecule (MCAM), matrix metallopeptidase 9 (MMP9) and plasminogen activator, urokinase (PLAU) (Figure 4).

Association of DACT1α methylation with the survival of gastric cancer patients

DACT1 α protein expression level was reduced in gastric cancer tissues compared with their matched adjacent normal tissues by Western blot (Figure 5A). DACT1 α methylation was evaluated in 20 normal gastric biopsies and 205 primary gastric cancer tissues by direct



Figure 3: (A) DACT1 α regulated PCP pathway by promoting Dvl-2 degradation and suppressing the activation of JNK pathway in AGS cells (cropped gel). (B) Real-time PCR validation for gene expression profile in DACT1 α stably transfected BGC823 cells screened by Cancer Pathway cDNA microarray. (C) Two represented genes PDGFB and MMP9 expression decreased in DACT1 α over-expressed GC cell lines (cropped gel).

Genebank Accession	Gene Symbol	Gene Name	Gene Function	Fold Change (DACT1/control)
NM_002608	PDGFB	Platelet-derived growth factor beta polypeptide	Proliferation &Angiogenesis	-1.5
NM_003376	VEGFA	Vascular endothelial growth factor A	Proliferation &Angiogenesis	-1.5
NM_181501	ITGA1	Integrin, alpha 1	Adhesion	-1.5
NM_002203	ITGA2	Integrin, alpha 2	Adhesion	-1.5
NM_002204	ITGA3	Integrin, alpha 3	Adhesion	-1.5
NM_000212	ITGB3	Integrin, beta 3	Adhesion	-1.6
NM_004994	MMP9	Matrix metallopeptidase 9	Invasion & Metastasis	-1.5
NM_006500	MCAM	Melanoma cell adhesion molecule	Invasion & Metastasis	-1.5
NM_002658	PLAU	Plasminogen activator, urokinase	Invasion & Metastasis	-1.8

Table 1: The effect of DACT1α on gene expression profiles of cancer pathways

bisulfite genomic sequencing. None was detected to have DACT1 α methylation in 20 healthy gastric tissue samples. DACT1 methylation was associated with advanced tumor size grade (P < 0.05), lymph node metastasis (P < 0.05) and distant metastasis (P = 0.05) respectively, but was not associated with the clinicopathological characteristics of patients with gastric cancer including age, gender, *H. pylori* infection, Lauren classification and tumor differentiation. Among 205 gastric cancer cases, we found that promoter methylation of DACT1 α was detected in 29.3% (60/205) of gastric cancers, but not in normal

controls. Overall survival of the gastric cancer patients was analyzed for dependence on DACT1 α promoter methylation using Kaplan-Meier survival curves. The overall survival of patients with DACT1 α methylation was significantly shorter than that of other gastric cancer patients (P = 0.007, Figure 5B). Using univariate Cox regression analysis, DACT1 α methylation was associated with a significantly increased risk of cancer-related death (P = 0.007, Table 2). In the multivariate Cox regression analysis, as expected, TNM stages were significantly associated with the survival of gastric cancer patients.



Figure 4: The molecular mechanisms underlying the tumor suppression of DACT1a in gastric cancer. Schematic diagram for the molecular basis of DACT1a as a tumor suppressor gene in gastric cancer. Ectopic expression of DACT1a suppressed gastric cancer cell growth was associated with several biological effects: 1) Suppression of PDGFB and VEGFA contributed to dampened cell proliferation and angiogenesis. 3) DACT1a mediated cell spreading adhesion, migration and invasion inhibition were associated with the suppression of ITGA1, ITGA2, ITGA3, ITGB3, MCAM, MMP9 and PLAU.

DACT1 α methylation was associated with the poor outcome in gastric cancer patients (Table 3).

DISCUSSION

In this study, we found that the mRNA expression of DACT1 α was silenced or reduced in 100% (8 of 8) gastric cancer cell lines, but expressed in normal gastric cell line and tissue. These results suggested that DACT1 α may play a tumor suppressive role in gastric cancer.

As the functional role of DACT1 α in cell migration and invasive abilities has not been well evaluated, we examine the effect of DACT1 α in mediating the cell migration and invasive abilities in gastric cancer cell lines. For metastasis functional study, we found that DACT1 α led to a significant decrease of cell spreading ability in BGC823 and MGC803 cells. At the beginning of cell spreading, most of DACT1a-re-expressed cells maintained a spherical shape whereas a large population of control cells had flatten out and spread onto the matrix. Cell spreading is the first step for cancer cell migrating and invading neighboring tissue. In order for cells to move and invade, they must adhere, flatten out and spread onto the extracellular matrix with extended protrusions. In keeping with reduced cell spreading ability, DACT1a significantly dampened cell migration and invasion as demonstrated by wound healing assay and trans-well invasion assay.

DACT1 α was an important regulator in PCP pathway [11, 12, 16]. Increasing evidences have demonstrated that PCP pathway played important roles in cancer development especially in tumor metastasis and angiogenesis [17]. Dishevelled (Dvl) 2, a core PCP component was over-expressed in non-small cell lung cancer and colorectal cancer and correlated to poor tumor differentiation and advanced TNM stages [18, 19]. Genetic deletion of Dvl2 reduced the intestinal tumor numbers in a dose-dependent way in the Apc(Min) model for colorectal cancer [19]. Therefore DACT1 α could inhibit tumor metastasis partially depending on PCP pathway.

In order to elucidate the other downstream targets modulated by DACT1 α in tumor inhibition, cancer pathway cDNA microarray was performed in BGC823-DACT1 α cell line. 9 markers covering cell proliferation, adhesion, migration, invasion and angiogenesis were involved in the anti-tumorigenesis effect of DACT1 α in gastric cancer. PDGFB and VEGFA were angiogenic factors and mediated tumor growth, angiogenesis and metastasis [20–23]. Therefore, down-regulation of PDGFB and VEGFA may explain the anti-proliferative effect exerted by DACT1 α in gastric cancer cells.

The reduced cell motility and spreading effect caused by DACT1 α in gastric cancer cells was revealed to be at least associated with altered PCP signaling pathway and JNK activity. DACT1 α



Figure 5: Kaplan-Meier survival curves show that gastric cancer patients with DACT1 α methylation had poorer survival than those without DACT1 α methylation. (A) Expression of DACT1 α protein was significantly down-regulated in GC tumor tissues compared to adjacent non-tumor tissues (n = 14) (normalized to GAPDH) (cropped gel). (B) Patients with DACT1 α hypermethylation had poor survival rate than those without methylation (U) based on the log-rank test (P = 0.007).

Variable	HR (95% CI)	P value	
Age	1.01 (0.99–1.03)	0.50	
Gender			
Male	1.07 (0.70–1.64)	0.77	
Female	1.00		
H. pylori infection			
Negative	1.46 (0.80–2.68)	0.22	
Positive	1.00		
Lauren			
Intestinal	1.25 (0.66–2.52)	0.53	
Diffuse	1.83 (0.72–4.66)	0.20	
Mixed	1.00		
TNM stage			
I	0.08 (0.02-0.24)	< 0.0005	
II	0.14 (0.06–0.32)	< 0.0005	
III	0.27 (0.16–0.44)	< 0.0005	
IV	1.00		
DACT1 methylation			
Yes	1.86 (1.08–2.82)	0.007	
No	1.00		

 Table 2: Univariate Cox regression analysis of potential prognostic factors for gastric cancer patients

 Table 3: Multivariate Cox regression analysis of potential prognostic factors for gastric cancer patients

Variable	HR (95% CI)	P value	
Age	1.01 (0.99–1.03)	0.33	
Gender			
Male	1.39 (0.87–2.24)	0.073	
Female	1.00		
TNM stage			
Ι	0.07 (0.02–0.21)	< 0.0005	
II	0.12 (0.05–0.29)	< 0.0005	
III	0.24 (0.14–0.40)	< 0.0005	
IV	1.00		
DACT1 methylation			
Yes	1.712 (1.034–2.87)	0.047	
No	1.00		

promoted central PCP component Dvl-2 degradation and suppressed PCP downstream pathway JNK activity in AGS cells. Consistently, DACT1 α knockout mice showed disregulated PCP signaling pathways by posttranslationally antaganizing Dvl2 and Vangl2 [11, 12, 15]. JNK pathway and RhoA pathway are two main PCP pathways affected by DACT1 α , both of which controlled cytoskeleton reorganization in cell movements and polarity [11, 12]. Less active RhoA but more active JNK was found in mouse embryonic fibroblast derived from DACT1 α knockout mice [11, 12], Notably, it has been well accepted that RhoA normally allowed the cell to maintain a spherical shape in the process of spreading whereas RhoA-depleted cells showed enhanced elongated and extended protrusions, permitting cells to flatten out, spread and invade much faster [24–26]. Therefore, DACT1 α over-expression modulated PCP pathways, contributing to decreased cell spreading activity, F-actin assembly, cell migration and invasion abilities.

Cancer Pathway cDNA microarray analysis revealed that DACT1 α down-regulated cell adhesion receptors from the integrin family including ITGA1, ITGA2,

ITGA3 and ITGB3. Integrins spanned the cell membrane and controlled adhesion to the extracellular matrix. Each integrin consisted of two subunits: α and β . One of the most well-studied integrins in cancer was oncogenic integrin $\alpha_{1}\beta_{2}$, of which ITGB3 (also known as β 3 integrin) was a key component [27]. Integrin $\alpha_{\mu}\beta_{3}$ was usually expressed at low or undetectable levels in most adult epithelia, but can be highly upregulated on the most aggressive tumor cells in a variety of cancer [27]. The expression of ITGB3 was restricted exclusively to cells within vertical growth phase and metastatic melanomas and was correlated with shorter survival [28]. Several integrin $\alpha_1\beta_2$ antagonists have been proved to be efficient in reducing tumor growth and metastasis in many mouse models and clinical studies [27]. In a mouse model of spontaneous tumorigenesis caused by WNT1 overexpression, the luminal epithelial progenitor marker ITGB3 identified a highly tumorigenic cancer stem cell population [29]. By recruiting c-Src to ITGB3 cytoplasmic tail, integrin $\alpha_{\alpha}\beta_{\alpha}$ activated c-Src, substantially increased anchorage-independent tumor cell survival in vitro and metastasis in vivo [27]. In addition, DACT1 α decreased the mRNA expression of ITGA1 and ITGA2, which were components of collagen receptor integrin $\alpha_1\beta_1$ and $\alpha_2\beta_1$. Integrin $\alpha_1\beta_1$ and $\alpha_2\beta_1$ were found to be important regulators in cell invasion in human cancers [30, 31]. Taken together, those down-regulated integrin molecules by DACT1a may at least offer partial explanation for impaired cell adhesion spreading ability, migration and invasion.

Other cell migration and invasion molecules regulated by DACT1a included PLAU, MCAM and MMP9. PLAU played important functions in cell migration, invasion and survival [32]. Elevated PLAU expression was found in many human cancers and correlated with the clinical features of biologically aggressive cancer and poor outcome of cancer patients [33, 34]. By binding urokinase-type plasminogen activator receptor (uPAR), PLAU promoted extracellular matrix (ECM) proteolysis and generated the protease plasmin which cleaved a range of ECM components and activated growth factors and matrix metalloproteases such as MMP9 [35]. Moreover, activated uPAR signals interacted with ITBG3 and triggered intracellular Rac signaling pathway for cell migration, leading to increased F-actin assembly and membrane protrusion [35]. Besides, MMP9 and MCAM were well-established metastasis mediators which actively involved in pathological processes, such as metastasis [36, 37].

To assess the clinical application of DACT1 α in gastric cancer, we examined the promoter methylation of DACT1 α by direct bisulfite sequencing in 205 primary gastric cancer patients and 20 normal controls. We found that promoter methylation of DACT1 α was detected in 29.3% (60/205) of gastric cancers, but not in normal controls. This result indicated that methylation mediated silencing of DACT1 α is a frequent event in gastric cancer. Moreover, in keeping with the dampened migration and invasive ability of gastric cancer cells by DACT1 α *in vitro*, DACT1 α methylation was significantly correlated with patient survival by Kaplan–Meier curves. Moreover, multivariate Cox regression analysis revealed that after adjustment for TNM stage, age and gender, gastric cancer patients with DACT1 α methylation showed poorer survival in gastric cancer patients (P < 0.05), suggesting that DACT1 α methylation may serve as an independent prognostic factor of gastric cancer patient.

In conclusion, we have identified a functional tumor suppressor gene DACT1 α inhibiting cell migration and invasion in gastric cancer. The impaired cell adhesion spreading ability, migration and invasion by DACT1 α are mediated through down-regulated integrin molecules. DACT1 α methylation may serve as a potential epigenetic biomarker to predict outcome of gastric cancer patients.

MATERIALS AND METHODS

Cell lines

Gastric cancer cell lines (AGS, Kato III, MKN28, MKN45, NCI87, SNU1, BGC823 and MGC803) and normal gastric epithelial GES1 cell line were obtained from American Type Culture Collection (Manassas, VA) and Beijing Oncology Hospital (China). Cell lines were maintained in RPMI-1640 or Dulbecco's modified Eagle's medium (DMEM) medium (Gibco BRL, Rockville, MD) with 10% fetal bovine serum. Human normal adult tissue RNA samples were purchased commercially (Stratagene, La Jolla, CA or Millipore Chemicon, Billerica, MA).

Human tissue samples

Gastric tissues samples were obtained from 205 Chinese gastric cancer patients and 20 normal controls. All patients diagnosed to have gastric cancer received surgical resection of the tumor. Clinical data of patients were collected from medical record and structured interview of patients. Clinical information included age, gender, survival time, TNM stage (I-IV), tumor size (I-IV), lymph node status (postive or negative), distant metastasis (postive or negative), differentiation (poor, moderately or well differentiated), H. pylori infection, tumor location, Lauren classification and chemotherapy. The informed consent was obtained from the patients and the study protocol was approved by the Clinical Research Ethics Committee of the First Hospital, Hebei Medical University and the methods were carried out in "accordance" with the approved guidelines.

Bisulfite modification of DNA and bisulfite genomic sequencing

Genomic DNA was extracted from fresh gastric tissues of healthy subjects or paraffin-embedded tissues of gastric cancer patients using DNA mini kit (Qiagen, Hilden, Germany). Extracted DNA was treated with sodium bisulfite using an EZ DNA Methylation-Gold[™] Kit (Zymo Research, Orange, CA). The bisulfitemodified DNA was amplified by using primer pairs that specifically amplify either methylated or unmethylated sequences of the DACT1a gene. The PCR product was separated by electrophoresis on 1% agarose with ethidium bromide staining. The amplicon were purified by using the ExoSAP-IT PCR Clean-up Kit for the removal of unwanted primers and dNTPs (GE Healthcare, Pittsburgh, PA). PCR product was mixed with ExoSAP-IT at the ratio of 5:2 and incubated for 15 min at 37°C followed by enzyme inactivation at 80°C for a further 15 min. The purified DNA was examined by direct sequencing PCR with the Big Dye Terminator Cycle Sequencing kit (Applied Biosystems). Sequencing PCR product was detected by ABI PRISM 7000 Sequence Detection System (Applied Biosystems).

RNA extraction, semi-quantitative RT-PCR and real-time PCR analyses

Total RNA was extracted by TRIzol Reagent (Molecular Research Center Inc, Cincinnati, OH) from cell pellets or tissues. cDNA was synthesised by Transcriptor Reverse Transcriptase (Roche, Indianapolis, IN). Semiquantitative RT-PCR was performed with AmpliTaq Gold DNA polymerase (Applied Biosystems, Foster City, CA). β -actin was employed as internal control. Real-time PCR was performed with SYBR Green master mixture (Roche, Indianapolis, IN) on LightCycler[®] 480 Instrument. 10ng cDNA template and 3pmol of primer pairs were applied in each 15µl PCR reaction. Each sample was tested in triplicate.

Construction of DACT1a expression plasmid

The full-length open reading frame sequence of DACT1 α was obtained by RT-PCR amplification of normal human gastric cDNA. DACT1 was cloned in pcDNA3.1+ and pBABE using primers as below:

c-FLAG- DACT1a-F2	GGactagtATGgactataag gacgatgatgacaagAAGCC GAGTCCGGCC
c-DACT1alpha-R2	ccgctcgagTCAAACCGT CGTCATCAGTTTC
pBABE-DACT1-F	AAAGGCAtacgtaATGA AGCCGAGTCCGGC
pBABE-DACT1-R	ACGCgtcgacTCAAACCG TCGTCATCAGTTTC

The PCR aliquots were subcloned into the mammalian expression vector pcDNA3.1 and then verified by DNA sequencing.

Establishment of stable DACT1α-expressing cells

BGC823, MGC803 or AGS cells were transfected with pcDNA3.1 or pcDNA3.1-DACT1a plasmid using lipofectamine 2000 (Life Technologies). Stable transfections were selected for 2 weeks with G418 antibiotics. For retroviruses production, 293FT cells (Invitrogen) were co-transfected with pBABE-puro-DACT1 or pBABE-puro empty vector, two packaging plasmids pUMVC (Addgene) and pCMV-VSV-G (Addgene) at the ratio of 1: 0.9: 0.1. At 24 hours posttransfection, cells were feeded with fresh medium. After 48 hour post-transfection, the supernatant containing retrovirus pBABE-puro-DACT1a or pBABE-puro control was harvested and stored at -80°C. To generate stable DACT1 α expressing or control cell line, retrovirus pBABE-puro-DACT1a or pBABE-puro controlcontaining supernatant was added to cells. After 48 hour transduction, the media containing retrovirus was removed and replaced with fresh medium containing 0.5 ug/ml antibiotic puromycin (Invitrogen) for selection.

RNA interference and transfection

Knock-down DACT1 α expression in GES1 cell lines was performed by a shRNA targeting DACT1 α . Both GES1-shRNA control and GES1-DACT1 α shRNA cells were selected for 2 weeks with G418 after transfection 48 h. The cells were ready for further experiments.

Cell spreading assay

BGC823 and MGC803 cells stably transfected with pcDNA3.1-DACT1 α or pcDNA3.1 were collected by trypsinization, washed twice with DMEM medium and were resuspended on cover slips and culture plate. Cells were allowed to spread for 6 hours in DMEM containing 10% FBS at 37°C and then photographed. The cell spreading rate was calculated as: number of spreading cells / number of total cells (%). The cover slips with spreading cells were fixed with 3% paraformaldehyde for F-actin staining. The experiments were performed in triplicate at three independent experiments. Every experiment was performed by using the different batches of transfection.

F-actin staining

Cell spreading assay was performed using BGC823 and MGC803 cells stably transfected with pcDNA3.1-DACT1 α or pcDNA3.1. The cover slips with spreading cells were fixed with 3% paraformaldehyde (in PBS) at room temperature for 15 min. Then the cells were washed three times with 1 × TBS and permeabilized with 0.1% Triton X-100 at room temperature for 5 min. Then cells were washed three times with $1 \times TBS$ and blocked with 1% BSA at room temperature for 30 min. For F-actin staining, cells were stained with Rhodamine phalloidin (1:40 dilution in 0.2% BSA) (Invitrogen) at 37°C for 20 min in dark. Finally, were washed and mounted with Mounting Medium containing DAPI (Vector Laboratories). The experiments were performed in triplicate at three independent experiments.

Wound-healing assay

To carry out the wound healing assay, AGS and MGC803 stably transfected with pcDNA3.1-DACT1 α or pcDNA3.1 control vector were seeded into 6-well culture plates. After 24 h, the monolayer AGS and MGC803 cells were then scratched manually with a plastic pipette tip, and after being washed with PBS, the wounded monolayers of the cells were allowed to heal for 24 hours in reduced serum RPMI 1640 and DMEM medium. Photographs were taken at 0 and 24 h respectively. The experiments were performed in triplicate at three independent experiments.

Invasion assay

To measure the cell invasion activity, Transwell assays were done using a BD BioCoat[™] Growth Factor Reduced MATRIGEL[™] Invasion Chamber (Cat. No. 354483, BD Biosciences). AGS and MGC803 cells were seeded into six-well dishes. After 24 h, the cells were transfected with pcDNA3.1-DACT1a or the empty control vector. After 48 h transfection, AGS and MGC803 cells $(2.5 \times 10^4 \text{ in } 500 \text{ uL})$ suspended in serum-free RPMI 1640/DMEM containing 0.1% bovine serum albumin were applied to the upper chamber. RPMI 1640/DMEM containing 20% fetal bovine serum was added to the lower chamber. After the cells were incubated at 37°C for 24 hours, the number of cells that migrated to the lower side of the upper chamber was counted. Three independent experiments were performed and results were shown as the means \pm s.d. The experiments were performed in triplicate at three independent experiments.

Cancer pathway finder PCR array and real-time PCR validation

Gene expression profiles of BGC823 cells stably transfected with pcDNA3.1- DACT1 α or pcDNA3.1 empty vector were analyzed by a commercial gene expression array system named the Human Cancer PathwayFinderTM RT[°] ProfilerTM PCR Array (96-well) (SABiosciences, Frederick, MD). This array contains 84 functionally well characterized genes representative of the six biological pathways involved in human tumorigenesis (Supplementary Table S1). For real-time PCR validation analysis, Ct was measured during the exponential amplification phase, and the amplification plots were analyzed using SDS 1.9.1 software (Applied Biosystems). The relative expression level (defined as fold change) of target gene is given by $2^{-\Delta\Delta Ct}$ ($\Delta Ct = \Delta Ct^{target} - \Delta Ct^{\beta actin}$; $\Delta\Delta Ct = \Delta Ct^{DACT_{1\alpha}-expressing} - \Delta Ct^{control}$) and normalized to the fold change detected in the corresponding control cells, which was defined as 1.0. All reactions were performed in duplicate. Primer sequences of target genes are listed in Supplementary Table S2.

Statistical analysis

The results were expressed as mean \pm standard deviation (SD). The difference in *in vitro* functional studies between the two groups was determined by the Student *t* test. Kaplan-Meier survival curves for trend were used to evaluate the relationship between DACT1 α methylation and the prognosis from the date of primary diagnosis to the end of follow-up. The analysis was performed using SPSS statistical package for Windows (version 16; SPSS). A *P* value of less than 0.05 was taken as statistical significance.

CONFLICTS OF INTEREST

There are no conflicts interest.

Authors' contributions

Y.L., J.Z., W.Y., X.Z. and G.W. performed experiments; Y.L. and J.Z. analyzed data and wrote the manuscript; Z.Z. supervised the study and revised the manuscript.

REFERENCES

- Bryan J. Dicken, David L. Bigam, Carol Cass, John R. Mackey, Anil A. Joy, Stewart M. Hamilton. Gastric Adenocarcinoma: Review and Considerations for Future Directions. Ann Surg. 2005; 241:27–39.
- Banyard J, Chung I, Migliozzi M, Phan DT, Wilson AM, Zetter BR, Bielenberg DR. Identification of genes regulating migration and invasion using a new model of metastatic prostate cancer. BMC Cancer. 2014; 14:387.
- Baud V, Karin M. Is NF-kappaB a good target for cancer therapy? Hopes and pitfalls. Nat Rev Drug Discov. 2009; 8:33–40.
- 4. Cheng X, Lu SH, Cui Y. ECRG2 regulates ECM degradation and uPAR/FPRL1 pathway contributing cell invasion/migration. Cancer Lett. 2010; 290:87–95.
- Cheyette BN, Waxman JS, Miller JR, Takemaru K, Sheldahl LC, Khlebtsova N, Fox EP, Earnest T, Moon RT. Dapper, a Dishevelled-associated antagonist of beta-catenin and JNK signaling, is required for notochord formation. Dev Cell. 2002; 2:449–61.

- Das R, Mahabeleshwar GH, Kundu GC. Osteopontin stimulates cell motility and nuclear factor kappaB-mediated secretion of urokinase type plasminogen activator through phosphatidylinositol 3-kinase/Akt signaling pathways in breast cancer cells. J Biol Chem. 2003; 278:28593–606.
- Deng J, Liang H, Zhang R, Ying G, Xie X, Yu J, Fan D, Hao X. Methylated CpG site count of dapper homolog 1 (DACT1) promoter prediction the poor survival of gastric cancer. Am J Cancer Res. 2014; 4:518–27.
- Desgrosellier JS, Cheresh DA. Integrins in cancer: biological implications and therapeutic opportunities. Nat Rev Cancer. 2010; 10:9–22.
- Gong H, Shen B, Flevaris P, Chow C, Lam SC, Voyno-Yasenetskaya TA, Kozasa T, Du X. G protein subunit Galpha13 binds to integrin alphaIIbbeta3 and mediates integrin "outside-in" signaling. Science. 2010; 327:340–3.
- Guo Y, Yin J, Wang Z, Zha L.Overexpression of plateletderived growth factor-B increases the growth, invasion, and angiogenesis of gastric carcinoma cells through protein kinase B. Neoplasma. 2013; 60:605–12.
- Hieken TJ, Ronan SG, Farolan M, Shilkaitis AL, Das Gupta TK. Molecular prognostic markers in intermediatethickness cutaneous malignant melanoma. Cancer. 1999; 85:375–82.
- Jia Y, Cao B, Yang Y, Linghu E, Zhan Q, Lu Y, Yu Y, Herman JG, Guo M. Silencing NKD2 by promoter region hypermethylation promotes gastric cancer invasion and metastasis by up-regulating SOX18 in human gastric cancer. Oncotarget. 2015; 6:33470–85. doi: 10.18632/ oncotarget.5272.
- Kodama M, Kitadai Y, Sumida T, Ohnishi M, Ohara E, Tanaka M, Shinagawa K, Tanaka S, Yasui W, Chayama K. Expression of platelet-derived growth factor (PDGF)-B and PDGF-receptor β is associated with lymphatic metastasis in human gastric carcinoma. Cancer Sci. 2010; 101:1984–9.
- Li J, Mo ML, Chen Z, Yang J, Li QS, Wang DJ, Zhang H, Ye YJ, Li HL, Zhang F, Zhou HM. HSulf-1 inhibits cell proliferation and invasion in human gastric cancer. Cancer Sci. 2011; 102:1815–21.
- 15. Li X, Cheung KF, Ma X, Tian L, Zhao J, Go MY, Shen B, Cheng AS, Ying J, Tao Q, Sung JJ, Kung HF, Yu J. Epigenetic inactivation of paired box gene 5, a novel tumor suppressor gene, through direct upregulation of p53 is associated with prognosis in gastric cancer patients. Oncogene. 2012; 31:3419–30.
- Mao D, Zhang Y, Lu H, Zhang H. Molecular basis underlying inhibition of metastasis of gastric cancer by anti-VEGFa treatment. Tumour Biol. 2014; 35:8217–23.
- Metcalfe C, Ibrahim AE, Graeb M, de la Roche M, Schwarz-Romond T, Fiedler M, Winton DJ, Corfield A, Bienz M. Dvl2 promotes intestinal length and neoplasia in the ApcMin mouse model for colorectal cancer. Cancer Res. 2010; 70:6629–38.
- Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. CA Cancer J Clin. 2005; 55:74–108.

- 19. Qu Y, Dang S, Hou P. Gene methylation in gastric cancer. Clin Chim Acta. 2013; 424:53–65.
- Rhee JW, Lee KW, Sohn WJ, Lee Y, Jeon OH, Kwon HJ, Kim DS. Regulation of matrix metalloproteinase-9 gene expression and cell migration by NF-kappa B in response to CpG-oligodeoxynucleotides in RAW 264.7 cells. Mol Immunol. 2007; 44:1393–400.
- Staniszewska I, Walsh EM, Rothman VL, Gaathon A, Tuszynski GP, Calvete JJ, Lazarovici P, Marcinkiewicz C. Effect of VP12 and viperistatin on inhibition of collagenreceptor-dependent melanoma metastasis. Cancer Biol Ther. 2009; 8:1507–16.
- Suriben R, Kivimäe S, Fisher DA, Moon RT, Cheyette BN. Posterior malformations in Dact1 mutant mice arise through misregulated Vangl2 at the primitive streak. Nat Genet. 2009; 41:977–85.
- 23. Suspitsin EN, Kashyap A, Shelekhova KV, Sokolenko AP, Kuligina ESh, Iyevleva AG, Kornilov AV, Ehemann V, Yanus GA, Aleksakhina SN, Preobrazhenskaya EV, Zaitseva OA, Yatsuk OS, et al. Evidence for angiogenesisindependent contribution of VEGFR1 (FLT1) in gastric cancer recurrence. Med Oncol. 2013; 30:644.
- 24. Tsai KW1, Wu CW, Hu LY, Li SC, Liao YL, Lai CH, Kao HW, Fang WL, Huang KH, Chan WC, Lin WC. Epigenetic regulation of miR-34b and miR-129 expression in gastric cancer. Int J Cancer. 2011; 129:2600–10.
- 25. Ushijima T, Asada K. Aberrant DNA methylation in contrast with mutations. Cancer Sci. 2010; 101:300–5.
- Vaillant F, Asselin-Labat ML, Shackleton M, Forrest NC, Lindeman GJ, Visvader JE. The mammary progenitor marker CD61/beta3 integrin identifies cancer stem cells in mouse models of mammary tumorigenesis. Cancer Res. 2008; 68:7711–7.
- 27. Van den Eynden GG, Van Laere SJ, Van der Auwera I, Trinh XB, Van Marck EA, Dirix LY, Vermeulen PB. Two distinct expression patterns of urokinase, urokinase receptor and plasminogen activator inhibitor-1 in colon cancer liver metastases. Int J Cancer. 2009; 125:1494–6.
- 28. Vega FM, Fruhwirth G, Ng T, Ridley AJ. RhoA and RhoC have distinct roles in migration and invasion by acting through different targets. J Cell Biol. 2011; 193:655–65.
- Wang S, Kang W, Go MY, Tong JH, Li L, Zhang N, Tao Q, Li X, To KF, Sung JJ, Yu J.Dapper homolog 1 is a novel tumor suppressor in gastric cancer through inhibiting the nuclear factor-kB signaling pathway. Mol Med. 2012; 18:1402–11.
- 30. Wang Y. Wnt/Planar cell polarity signaling: a new paradigm for cancer therapy. Mol Cancer Ther. 2009; 8:2103–9.
- Wei Q, Zhao Y, Yang ZQ, Dong QZ, Dong XJ, Han Y, Zhao C, Wang EH. Dishevelled family proteins are expressed in non-small cell lung cancer and function differentially on tumor progression. Lung Cancer. 2008; 62:181–92.

- 32. Wen J, Chiang YJ, Gao C, Xue H, Xu J, Ning Y, Hodes RJ, Gao X, Chen YG. Loss of Dact1 disrupts planar cell polarity signaling by altering dishevelled activity and leads to posterior malformation in mice. J Biol Chem. 2010; 285:11023–30.
- 33. Xu L, Li X, Chu ES, Zhao G, Go MY, Tao Q, Jin H, Zeng Z, Sung JJ, Yu J. Epigenetic inactivation of BCL6B, a novel functional tumour suppressor for gastric cancer, is associated with poor survival. Gut. 2012; 61:977–85.
- 34. Xu X, Chang X, Li Z, Wang J, Deng P, Zhu X, Liu J, Zhang C, Chen S, Dai D. Aberrant SOX11 promoter

methylation is associated with poor prognosis in gastric cancer. Cell Oncol. 2015; 38:183-94.

- 35. Xu ZY, Chen JS, Shu YQ. Gene expression profile towards the prediction of patient survival of gastric cancer. Biomed Pharmacother. 2010; 64:133–9.
- Singh D, Srivastava SK, Chaudhuri TK, Upadhyay G. Multifaceted role of matrix metalloproteinases (MMPs). Front Mol Biosci. 2015; 2:19.
- 37. Wang Z, Yan X. CD146, a multi-functional molecule beyond adhesion. Cancer Lett. 2013; 330:150–62.