Research Paper

Correlation between serum IL-1β and miR-144-3p as well as their prognostic values in LUAD and LUSC patients

Chen Wu^{1,2,3}, Bin Xu^{3,4}, You Zhou^{3,4}, Mei Ji^{1,3}, Dachuan Zhang⁵, Jingting Jiang^{2,3,4}, Changping Wu^{1,2,3}

¹Department of Oncology, The Third Affiliated Hospital of Soochow University, Changzhou, 213003, P.R.China

²Institute of Cell Therapy, Soochow University, Changzhou, 213003, P.R.China

³Jiangsu Engineering Research Center for Tumor Immunotherapy, Changzhou, 213003, P.R.China

⁴Department of Biological Treatment, The Third Affiliated Hospital of Soochow University, Changzhou, 213003, P.R.China

⁵Department of Pathology, The Third Affiliated Hospital of Soochow University, Changzhou, 213003, P.R.China

Correspondence to: Jingting Jiang, email: Jiangjingting@suda.edu.cn Changping Wu, email: wcpjjt@163.com

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ABSTRACT

Background: IL-1 β is an essential factor of inflammation initiation, and it also promotes malignant transformation, indicating its tumorigenic property. We aimed to investigate the correlation between IL-1 β and miR-144-3p as well as their prognostic values in LUAD and LUSC patients.

Results: The IL-1 β level in both LUAD and LUSC patients was significantly higher than that of healthy donors (P < 0.001). In both populations, patients with low IL-1 β level had better prognosis than high IL-1 β level (P < 0.001 and P = 0.010, respectively). In A549 cells, miR-144 showed the biggest expression change (-4.38 fold) after IL-1 β exposure. In LUAD patients, a negative correlation was detected between IL-1 β and miR-144-3p (r = -0.540, P < 0.001) and the high miR-144-3pgroup had better prognosis (P = 0.003), which was validated by TCGA data. Clinical stage, IL-1 β and miR-144-3p were independent risk factors in LUAD patients. *In vitro*, IL-1 β and miR-144-3p mimics would attenuate the promoting effect of IL-1 β .

Materials and Methods: ELISA and qRT-PCR were applied respectively to detected cytokines and miR-144-3p in 129 LUAD, 54 LUSC and 40 healthy donors. Moreover, miRNA array was carried out for miRNA profiling. TCGA database was employed for validation, and follow up data were collected for prognosis evaluation. MTT assay and western-blot were carried out for proliferation evaluation.

Conclusions: In LUAD patients, the serum IL-1 β level was correlated with miR-144-3p may affect miR-144-3p at transcriptional level. Both of them were independent risk factors for LUAD prognosis. In addition, IL-1 β and miR-144-3p might mediate inflammation-promoted tumorigenesis in LUAD patients.

INTRODUCTION

Lung cancer remains the leading cause of cancerrelated death worldwide. It accounts for approximately 1.58 million deaths in 2015 in USA, and such deaths are projected to continue rising [1]. According to histological characteristics, lung cancer can be categorized into non-small-cell lung cancer (NSCLC) and small-cell lung cancer (SCLC). In NSCLC, lung adenocarcinoma (LUAD) and lung squamous cell carcinoma (LUSC) account for 60% and 25% of this population, respectively [2].

Besides histological differences, many aspects, including genetics, pathogenesis, biological behavior, treatment and prognosis, significantly vary between LUAD and LUSC [3–7]. As a result, it is highly recommended by both scientists and clinical doctors to distinguish them as two individual diseases.

It has been well established that chronic inflammation and infection are pivotal factors of carcinogenesis [8]. Interleukin-1 beta (IL-1 β), which is produced by activated macrophages as a proprotein and processed to its active form by Caspase 1, is a member of the IL-1 cytokine family [9]. IL-1 β is an important mediator of the inflammatory response and involved in a variety of cell behaviors. The induction of cyclooxygenase-2 (PTGS2/ COX2) by IL-1 β in the central nervous system (CNS) is found to contribute to inflammatory pain hypersensitivity [10]. Besides, IL-1 β promotes malignant transformation and tumor aggressiveness in oral cancer [11], indicating its tumorigenic role.

Recently, mounting evidence has shown that microRNAs (miRNAs) are a novel class of inflammationassociated molecules and involved in multi-stages of tumor development, including initiation, tumor progression, invasion and metastasis [12–14]. Tumorrelated miRNAs can be regulated/influenced by various inflammatory stimuli, including cytokines [15–17]. Lin et al. demonstrated that IL-1 β upregulates Lin28B by downregulating miR-101 and promotes proliferation and migration of NSCLC cells, defining an IL-1 β /miR-101/Lin28B pathway in NSCLC [18].

As NSCLC is a tissue specific tumor, its corresponding treatment strategies are dissimilar widely. Oncologists and scientists currently consider that LUAD and LUSC are two individual diseases with completely different genetic background [3]. In this study, we detected cytokines in both LUAD and LUSC patients and found that the IL-1 β levels in both LUAD and LUSC patients were significantly higher than that of healthy donors. Moreover, in both LUAD and LUSC, low IL-1B cases had better prognosis that high IL-1ß ones. In addition, miR-144-3p was a potential downstream miRNA correlated with and influenced by IL-1 β in LUAD. LUAD patients with high miR-144-3p level had better prognosis. Based on TCGA database, the miR-144-3p level in tumor tissue was crucially higher than that in adjacent area in both LUAD and LUSC. However, miR-144-3p was associated with prognosis only in LUAD population, , which was consistent with our results. IL-1ß and miR-144-3p suppression could enhance proliferation of LUAD cells. Therefore, our findings suggested a potential link between IL-1 β and miR-144-3p, which might be associated with LUAD prognosis, providing new insights into inflammation-promoted tumorigenesis.

RESULTS

Comparison of various cytokines in serum from LUAD, LUSC and healthy donors

A total of six cytokines were detected using ELISA method in LUAD, LUSC patients and healthy donors. The results showed that the IL-1 β level in LUAD patients was significantly higher than that of healthy donors

(P < 0.001), whereas levels of IL-4, IL-6, IL-8, IL-10 or TNF- α slightly varied between LUAD patients and healthy donors (Figure 1A). The IL-1 β level in LUSC patients was significantly higher than that of healthy donors, while levels of IL-8, IL-10 and TNF- α were lower in LUSC than healthy donors (P < 0.001). However, IL-4 and IL-6 levels slightly varied between the two groups (Figure 1B).

Association between IL-1β and clinical features in LUAD and LUSC patients

The IL-1 β level was defined as "high" and "low" using median (28.3992 pg/mL and 43.5495 pg/mL in LUAD and LUSC, respectively) as threshold. IL-1 β level of LUAD and LUSC patients was analyzed according to age, gender, smoking status and pathological stage, as well as target therapy status in LUAD patients. The results showed that in LUSC patients, the IL-1 β level was associated with pathological stage (P < 0.05). However, no significant associations were observed between IL-1 β level and other above-mentioned clinical features in either LUAD or LUSC population (Table 1).

Prognosis evaluation between high and low cytokine levels in LUAD and LUSC patients

Levels of cytokines were defined as "high" and "low" using medians as thresholds. In both LUAD (Figure 2A) and LUSC (Figure 2B) populations, patients with low IL-1 β level showed better prognosis than those with high IL-1 β level (P < 0.001 and P = 0.010, respectively). In LUAD population, patients with high TNF- α level exhibited better prognosis than those with low TNF- α level (P = 0.017). No significant differences in prognosis were observed between patients with high and low IL-4, IL-6, IL-8 or IL-10 levels (Figure 2).

The prognostic value of miR-144-3p and its correlation with IL-1 β

Figure 3A reveals that hsa-miR-17, hsa-miR-338-3p, hsa-miR-579, hsa-miR-3156-5p, hsa-miR-144 and hsa-miR-1254 ranked as the top six mostly altered miRNAs after IL-1 β exposure in A549 cells, with an expression change greater than \pm 4 fold-change, among which hsa-miR-144 showed the biggest change (-4.38 fold). Real-time PCR was carried out to validate the array data, and results showed that both pri-miR-144-3p and miR-144-3p were down-regulated in A549-IL-1 β group (P < 0.05), indicating that IL-1 β could regulate the transcription of miR-144-3p.

In LUAD patients, a negative correlation was detected between IL-1 β and miR-144-3p (r = -0.540, P < 0.001, Figure 4A), and the high miR-144-3p group had better prognosis than low miR-144-3p group (P = 0.003, Figure 4C). In contrast, miR-144-3p was neither correlated with IL-1 β (r = -0.159, P = 0.250, Figure 4B) nor

prognosis (P = 0.633, Figure 4D) in LUSC patients. TCGA data displayed that miR-144-3p was significantly higher in both tumor tissues compared with adjacent area in both LUAD (Figure 5A) and LUSC (Figure 5B) populations, but its expression was only associated with prognosis in LUAD patients (Figure 5C and 5D). By χ^2 test, we confirmed that miR-144-3p was correlated with IL-1 β only in LUAD patients (Table 2).

In LUAD patients, univariate analysis showed that age, clinical stage, target therapy, IL-1 β , TNF- α and miR-144-3p were significant prognostic factors, while gender and smoking status showed no statistical differences (Table 2). As an association was determined between IL-1 β and miR-144-3p, two separate multivariate models (IL-1 β and miR-144-3p) were run to avoid problems of multicollinearity. Multivariate analyses demonstrated that



Figure 1: Comparison of various cytokines in serum from a set of 129 LUAD, 54 LUSC and 40 healthy donors. IL-1 β level in LUAD patients was significantly higher than Healthy donors (P < 0.001), whereas IL-4, IL-6, IL-8, IL-10 or TNF- α levels varied slightly between LUAD and healthy donors (**A**); IL-1 β level in LUSC patients were significantly higher than healthy donors, while IL-8, IL-10 and TNF- α were lower in LUSC than healthy donors (P < 0.001), however, IL-4 and IL-6 levels varied slightly between the two groups (**B**).

	LUAD				LUSC			
Characteristics	All patients	IL-1β Low (< median ^a)	IL-1β High (≥ medianª)	р	All patients	IL-1β Low (< median ^b)	IL-1β High (≥ median ^b)	р
n	129	64 (49.60%)	65 (50.40%)		54	27 (50.00%)	27 (50.00%)	
Age (years)				0.842				0.695
Median (range)	65 (48-85)	65 (49–84)	65 (48–85)		64 (50-85)	64 (50-83)	64 (51–85)	
Gender				0.401				0.761
Male	93 (72.09%)	44 (34.11%)	49 (37.98%)		39 (%72.22)	20 (37.05%)	19 (35.17%)	
Female	36 (27.91%)	20 (15.50%)	16 (12.40%)		15 (27.78%)	7 (12.97%)	8 (14.81%)	
Smoke				0.334				0.248
Yes	61 (47.29%)	33 (25.58%)	28 (21.71%)		36 (66.67%)	20 (37.04%)	16 (29.63%)	
No	68 (52.71%)	31 (24.03%)	37 (28.68%)		18 (33.33%)	7 (12.96%)	11 (20.37%)	
Pathological stage ^c				0.330				0.023
IA	17 (13.18%)	11 (8.53%)	6 (4.65%)		11 (20.37%)	8 (14.81%)	3 (5.56%)	
IB	34 (26.36%)	20 (15.50%)	14 (10.85%)		10 (18.52%)	8 (14.81%)	2 (3.71%)	
IIA	23 (17.83%)	12 (9.30%)	11 (8.53%)		11 (20.37%)	5 (9.26%)	6 (11.11%)	
IIB	32 (24.81%)	13 (10.08%)	19 (14.73%)		13 (24.07%)	5 (9.26%)	8 (14.81%)	
IIIA	11 (8.53%)	4 (3.10%)	7 (5.43%)		8 (14.81%)	1 (1.85%)	7 (12.96%)	
IIIB	12 (9.30%)	4 (3.10%)	8 (6.20%)		1 (1.85%)	0 (0.00%)	1 (1.85%)	
Target therapy				0.537				NT^{d}
Yes	65 (50.39%)	34 (26.36%)	31 (24.03%)		NT^{d}	NT^d	NT^d	
No	64 (49.61%)	30 (23.26%)	34 (26.36%)		NT ^d	NT ^d	NT^d	

Table 1: Association between IL-1 β and clinical features in LUAD and LUSC patients

Median^a = 28.3992 pg/ml Median^b = 43.5495 pg/ml NT^d = Not Tested

Pathological stage^C was based on NCCN Guidelines Version 3.2011 NSCLC.

clinical stage, IL-1 β and miR-144-3p were independent risk factors for OS (Table 3). In LUSC patients, no independent risk factor was verified (Table 4).

The regulatory effect of IL-1β and miR-144-3p on LUAD cell proliferation

In A549 cells, IL-1 β exposure and miR-144-3p mimics enhanced and suppressed cell proliferation, respectively (*P* < 0.001, Figure 6A). The combination of IL-1 β exposure and miR-144-3p silencing promoted proliferation more significantly than mono-treatment (*P* < 0.0001, Figure 6B). Western-blot validated MTT result that PCNA protein level was increased after IL-1 β exposure (Figure 6C and 6D), while it was increased and decreased after miR-144-3p mimics and antagomir transfection, respectively.

DISCUSSION

In the present study, we found that the IL-1 β level was significantly higher in LUAD and LUSC patients. In LUAD patients, serum IL-1 β level was correlated with miR-144-3p, both of which were associated with prognosis. Moreover, IL-1 β was involved in regulation of miR-144-3p

at the transcriptional level; IL-1 β and miR-144-3p overexpression both could promote the cell proliferation.

Cytokines play key roles in initiation and development of cancer [19, 20]. IL-1ß has been approved to be an essential factor in the inflammation initiation [9]. Gui et al. demonstrated that IL-1 β can trigger HIF-1 through NF-kB/COX-2 signaling pathway, suggesting that HIF-1 is a bridge linking inflammation and tumor and IL-1 β possesses a cancer promoting property [21]. IL-1β-induced COX-2 would mediate the evolution from inflammation to cancer through HIF-1, and such a process could be inhibited or suppressed by nonsteroidal anti-inflammatory drugs (NSAIDs), such as Aspirin and Celecoxib, leading to apoptosis of cancer cells [22]. Nevertheless, a large portion of NSCLC patients can not benefit from NSAIDs but suffer tumor progression quickly in clinic [23]. A few studies have approved that Celecoxib can promote epithelial-mesenchymal transition (EMT) of cancer cells, leading to increased metastatic risk [24, 25]. The conflict results indicated that COX-2/HIF1 is probably not the only cancer promoting pathway in downstream of IL-1 β . In general, the existing evidence is not convincing enough to elucidate carcinogenetic mechanism of IL-1 β , and new breakthroughs are urgently desired.

Despite under the same catalog of NSCLC, LUAD and LUSC greatly differ in genetics, histology, treatment and prognosis. Recently, Campbell JD et al. identified distinct patterns of somatic genome alterations in LUAD and LUSC. The genetics of the two major types of NSCLC significantly vary, which can be defined as individual diseases in certain level. Although targeted therapies for LUAD and LUSC are largely distinct, immunotherapy may aid in treatment for both types [3]. In the present study, we found that only in LUAD, serum IL-1 β level was correlated with miR-144-3p, and both of them were independent risk factors for prognosis. However, no such



Figure 2: Prognosis evaluation between high and low cytokine levels in LUAD and LUSC patients. In LUAD, patients with low IL-1 β and high TNF- α levels showed better prognosis than those with high IL-1 β (P < 0.001) and low TNF- α (P = 0.017) levels, respectively, whereas no significant differences in prognosis were observed between patients with high and low IL-4 (P = 0.509), IL-6 (P = 0.952), IL-8 (P = 0.665) or IL-10 (P = 0.649) levels (**A**); In LUSC, patients with low IL-1 β level had better prognosis than high IL-1 β (P = 0.010) level. However, no significant prognostic differences were detected between patients with high and low IL-4 (P = 0.802), IL-6 (P = 0.362), IL-8 (P = 0.439), IL-10 (P = 0.288) or TNF- α (P = 0.605) levels (**B**).

significant prognostic value of IL-1 β or miR-144-3p was obtained in LUSC. Our result was validated by TCGA database. Therefore, we suggested to investigate LUAD and LUSC separately to avoid major confounding factors in the future studies.

In this study, IL-8, IL-10 and TNF- α levels in LUSC were lower than those of healthy donors (P < 0.001). However, no prognostic value was observed. Although a number of studies have demonstrated the correlation between cytokine levels and prognosis in NSCLC patients, the population size was limited in the present study. The potential reason and mechanism remain unclear, and further investigation is necessary.

Although great efforts have been made to improve diagnosis and effective treatment, the prognosis of lung cancer remains poor due to lack of fine biomarkers. Accumulating evidence has confirmed that miRNAs are involved and play pivotal role in the occurrence, development and prognosis of tumor [26, 27]. Our results indicated that IL-1 β probably promoted cell proliferation via suppression of miR-144-3p. Tumor miRNA is ubiquitous and stable in peripheral blood, and it is convenient to detect by real-time PCR or miRNA sequencing [28–30], making it a potential marker with synergetic diagnostic value. miRNA participates in every step of "inflammation--premalignancy--malignancy" process, including cytokine release and immune response [14]. So far, it remains unclear how miRNA mediates and regulates "inflammation-cancer transduction" in lung cancer. Therefore, such an investigation is important for the patients. Based on our findings, we hypothesized that IL-1β might regulate miR-144-3p transcription and control various biological behaviors, and its expression was associated with prognosis in LUAD patients. However, it still remains undefined how miR-144-3p acts and what the cross-talk between miR-144-3p and COX-2/ HIF-1 pathway is.

Taken together, we demonstrated that in LUAD patients, the serum IL-1 β level was correlated with

miR-144-3p, and it could affect miR-144-3p at the transcriptional level. Moreover, both of them were associated with prognosis and independent risk factors for LUAD patients. In addition, IL-1 β might promote cell proliferation via suppression of miR-144-3p. As the limit on the sample size, more elaborate studies are necessary for further exploration of the link between IL-1 β and miR-144-3p and their roles in tumorigenesis.

MATERIALS AND METHODS

Patients, samples and cell line

The human NSCLC cell line A549 was obtained from American Type Culture Collection (ATCC, Manassas, USA) and cultured according to the guideline.

The study was performed on a total of 183 randomly selected NSCLC patients (129 LUADs and 54 LUSCs). Specimens were collected during biopsies and surgery from the Third Affiliated Hospital of Soochow University (Changzhou, China), and freshly collected samples were immediately fixed in formaldehyde. Each patient participated in the study after informed consent was provided. All specimens were histologically and blindly classified by two professional pathologists according to the national NCCN guidelines for NSCLC. Serum samples were collected from the same 183 patients and 40 extra healthy donors and stored in liquid nitrogen.

Enzyme linked immunosorbent assay (ELISA)

Serum samples from patients and healthy donors were obtained before biopsy or surgery from the Third Affiliated Hospital of Soochow University (Changzhou, China). The levels of cytokines, including IL-1 β , IL-4, IL-6, IL-8, IL-10 and TNF- α , were determined using ELISA assay kits according to manufacturers instructions (R&D, Shanghai, China).



Figure 3: IL-1 β was involved in regulation of miR-144-3p at the transcriptional level. Via miRNA array, a total of six miRNAs varied greater than 4 folds (including miR-144-3p) in A549 cells after high IL-1 β exposure. (**A**); Real-time PCR validated the result of miRNA array and showed that both pri-miR-144-3p and miR-144-3p were down-regulated in A549-IL-1 β group (P < 0.05, **B**).



Figure 4: Serum miR-144-3p level was correlated with IL-1β level and associated with prognosis in LUAD. Via Pearson correlation analysis, serum level of miR-144-3p was negatively correlated with IL-1β (r = -0.540, P < 0.001) (A) and LUAD patient with high miR-144-3p level showed significantly better prognosis than the ones with low miR-144-3p level (P = 0.003) (C); In the contrary, in LUSC patients, serum miR-144-3p level was neither correlated with IL-1β (r = -0.550) (B) nor associated with prognosis (P = 0.633) (D).



Figure 5: TCGA data validated the correlation of miR-144-3p and prognosis. Based on TCGA database, miR-144-3p level in tumor tissue was crucially higher than that in adjacent area in both LUAD (P < 0.001) (**A**) and LUSC (P < 0.001) (**B**) patients; in LUAD, patients with high miR-144-3p had significantly longer OS than low miR-144-3p group (P < 0.001) (**C**), while no distinct prognostic difference was found in LUSC population (P = 0.073) (**D**).

Characteristics All patients miR-144-3p Low (< median
n 129 (100%) 64 (49.61%) 65 (50.39%) 54 (100%) 27 (50.00%) 27 (50.00%) Age (years) 0.982 0.982 0.982 0.817 Median (range) 65 (48-85) 65 (48-85) 65 (49-84) 64 (50-85) 64 (51-85) 64 (50-83) Gender 0.736 0.362 0.363 0.363 0.363 0.363 Male 93 (72.09%) 47 (36.43%) 46 (35.66%) 39 (72.22%) 21 (38.89%) 18 (33.33%) 0.363 Female 36 (27.91%) 17 (17.18%) 19 (14.73%) 15 (27.78%) 6 (11.11%) 9 (16.67%) 1.000 Smoke 0.425 1.000 36 (27.91%) 32 (24.80%) 36 (66.67%) 18 (33.33%) 18 (33.33%) 18 (33.33%) Yes 68 (52.71%) 36 (27.91%) 32 (24.80%) 36 (66.67%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 18 (33.33%) 10 (35.65%) 10 (35.65%)
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Pathological stage ^c 0.726 0.14
IA 17 (13.18%) 10 (7.75%) 7 (5.43%) 11 (20.37%) 3 (5.56%) 8 (14.81%)
IB 34 (26.35%) 14 (10.85%) 20 (15.50%) 10 (18.52%) 6 (11.11%) 4 (7.41%)
IIA 23 (17.83%) 11 (8.53%) 12 (9.30%) 11 (20.37%) 8 (14.81%) 3 (5.56%)
IIB 32 (24.81%) 17 (13.18%) 15 (11.63%) 13 (24.07%) 7 (12.96%) 6 (11.11%)
IIIA 11 (8.53%) 7 (5.43%) 4 (3.10%) 8 (14.81%) 2 (3.70%) 6 (11.11%)
IIIB 12 (9.30%) 5 (3.88%) 7 (5.43%) 1 (1.85%) 1 (1.85%) 0 (0.00%)
IL-1β Level <0.001 0.05
Low 64 (49.61%) 11 (8.52%) 53 (41.09%) 27 (50.00%) 10 (18.52%) 17 (31.48%)
High 65 (50.39%) 53 (41.09%) 12 (9.30%) 27 (50.00%) 17 (31.48%) 10 (18.52%)
Target therapy0.660NTd
Yes 64 (49.61%) 33 (25.58%) 31 (24.03%) NT ^d NT ^d NT ^d
No 65 (50.39%) 31 (24.03%) 34 (26.36%) NT ^d NT ^d

Fable 2: Correlation between miR-144-	p and clinical features in LUAD and LUSC pa	atients
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Median^a = 8.1731 Median^b = 5.8994 NT^d = Not Tested

Pathological stage^C was based on NCCN Guidelines Version 3.2011 NSCLC.

NCCN, National Comprehensive Cancer Network; NSCLC, non-small-cell lung cancer.

miRNA isolation

Total RNA was isolated from 400 µL serum of each patient using the miRNeasy Mini Kit (Qiagen, Shanghai, China) according to the manufacturer's instructions. Purified total RNA was used for further real-time PCR and miRNA array test.

miRNA array

Serum miRNA profile was generated using RiboArray TM miDETECT TM Human Array 1 × 12K (Ribobio, Guangzhou, China) according to the manufacturer's instructions.

Quantitative reverse transcription PCR (qRT-PCR)

The level of miR-144-3p was quantified by qRT-PCR using Bulge-Loop[™] miRNA qRT-PCR Primer Set (Ribobio, Guangzhou, China), and results were analyzed and expressed by $^{\Delta\Delta}$ CT method.

The cancer genome atlas (TCGA) database

LUAD and LUSC samples with miR-144-3p expression and clinical information were obtained from TCGA database, yielding a total of 452 LUADs and 343 LUSCs patients. The K-means cluster method (K = 2) was carried out to cluster the patients into two groups based on their miR-144-3p expression. The log-rank test and Kaplan-Meier curve were performed to evaluate the statistical significance and prognosis of the two groups.

Cell culture, miR-144-3p mimics and antagomir transfection

LUAD cell line A549 was employed for in vitro experiments. For treatment, cells were treated with 10 ng/mL IL-1β (PrimeGene, Shanghai, China). Mimics

Clinicopathological	Comparison	Univariate analys	is	Multi-variate analysis		
parameters	reference	Hazard ratio (95%CI)	P value	Hazard ratio (95%CI)	P value	
Age	< 65	Ref		Ref		
	≥ 65	1.906 (1.079–3.366)	0.026	1.566 (0.838–2.926) ^a	0.159ª	
Gender	Male	Ref				
	Female	1.176 (0.640-2.160)	0.602			
Smoke	No	Ref				
	Yes	1.577 (0.891-2.790)	0.118			
Clinical Stage	IA	Ref	< 0.001	Ref	$< 0.001^{a}$	
	IB	1.058 (0.177-6.338)	0.951	0.907 (0.150–5.499) ^a	0.916ª	
	IIA	4.515 (0.934–21.817)	0.061	4.513 (0.919–22.164) ^a	0.064ª	
	IIB	6.102 (1.397–26.660)	0.016	4.278 (0.966–18.945) ^a	0.056ª	
	IIIA	19.666 (4.234–91.348)	< 0.001	13.101 (2.796–61.386) ^a	0.001ª	
	IIIB	10.963 (2.415-49.760)	0.002	6.050 (1.307–27.996) ^a	0.021ª	
Target therapy	Yes	Ref		Ref		
	No	2.000 (1.129-3.542)	0.018	1.697 (0.934–3.083) ^a	0.083ª	
IL-1β level	Low	Ref		Ref		
	High	3.818 (1.986-7.339)	< 0.001	3.166 (1.592–6.297) ^a	0.001ª	
TNF-α level	Low	Ref		Ref		
	High	1.954 (1.101-3.466)	0.022	1.332 (0.730–2.431) ^a	0.350ª	
miR-144-3p level	Low	Ref		Ref		
	High	0.427 (0.238-0.765)	0.004	0.365 (0.196–0.681) ^b	0.002 ^b	

Table 3: Univariate and multi-variate analysis of the clinicopathological and molecular features for OS in LUAD

^aMultivatiate model including Age, Clinical Stage, Target therapy, IL-1β & TNF-α.

^bMultivatiate model including Age, Clinical Stage, Target therapy, miR-144-3p & TNF-α.

and antagomir were purchased from Ribobio Co., Ltd. (Guangzhou, China) in order to induce and suppress the miR-144-3p expression, respectively.

MTT assay

Cell proliferation was determined using Vybrant MTT Cell Proliferation Assay Kit (Invitrogen, Shanghai, China). Cells were collected at 0, 24, 48 and 72 h after IL-1 β exposure and miR-144-3p mimics/antagomir transfection. Absorbance at a wavelength of 570 nm was determined on a spectrometer. The experiment was performed in triplicates.

Protein extraction and Western-blot analysis

Total proteins were extracted using CHAPS lysis buffer. Proteins were separated by sodiem dodecyl sulfate polyacrylamide gel electrophoresis and transferred onto polyvinylidene fluoride (PVDF) membrane. Blots were blocked in 4% dry milk at room temperature for 1 h, and immuno-stained with primary antibodies (anti-PCNA, 1:2,000; Cell signaling Technology, Shanghai, China and β -actin 1:10,000; Dako, Glostrup, Denmark) at 4°C overnight. The results were visualized via a chemiluminescent detection system (Pierce ECL Substrate Western-blot detection system; Thermo, Rockford, IL) and exposed in Molecular Imager ChemiDoc XRS System (Bio-Rad, Hercules, CA).

Statistical analysis

Statistical analysis was performed using GraphPad Prism (version 5.01; GraphPad Software, Inc., La Jolla, USA) and SPSS (version 17, SPSS Inc., Chicago, USA) statistical softwares. The Student's *t* test and paired t test were used to analyze significance between independent groups and paired materials, respectively. The correlation test was used to analyze the correlation between IL-1 β and miR-144-3p. The χ^2 test was used to test the significance of observed differences in proportions except when the cell size was less than 5 (Fisher's exact tests). Survival

Clinicopathological	Comparison	Univariate analysi	S	Multi-variate analysis	
parameters	reference	Hazard ratio (95%CI)	P value	Hazard ratio (95%CI)	P value
Age	< 65	Ref			
	≥ 65	0.649 (0.280–1.503)	0.307		
Gender	Male	Ref			
	Female	0.913 (0.375-2.224)	0.842		
Smoke	No	Ref			
	Yes	0.909 (0.384–2.153)	0.828		
Clinical Stage	IA	Ref	0.039	Ref	0.221
	IB	2.830 (0.255-31.346)	0.397	3.088 (0.275-34.686)	0.361
	IIA	5.361 (0.626-45.921)	0.125	4.805 (0.550-41.949)	0.156
	IIB	9.739 (1.186–79.956)	0.034	8.430 (0.995–71.386)	0.050
	IIIA	17.498 (2.100–145.829)	0.008	13.287 (1.442–122.460)	0.022
	IIIB	19.248 (1.157–320.344)	0.039	14.606 (0.812-262.349)	0.069
IL-1β level	Low	Ref		Ref	
	High	3.106 (1.218-7.917)	0.018	1.531 (0.503–4.657)	0.453
TNF-α level	Low	Ref			
	High	0.810 (0.357-1.839)	0.615		
miR-144-3p level	Low	Ref			
	High	1.216 (0.533–2.777)	0.642		

 Table 4: Univariate and multi-variate analysis of the clinicopathological and molecular features for OS in LUSC



Figure 6: L IL-1 β and miR-144-3p were involved in regulation of cell proliferation. In A549, IL-1 β exposure and miR-144-3p mimics enhanced and suppressed cell proliferation, respectively (P < 0.001) (**A**). The combination of IL-1 β exposure and miR-144-3p silencing promoted proliferation more significantly than mono-treatment (P < 0.0001) (**B**). Western-blot validated MTT result that PCNA protein level was increased after IL-1 β exposure (**C** and **D**), while it was increased and decreased after miR-144-3p mimics and antagomir transfection, respectively.

curves were made using the Kaplan-Meier method and compared using the log-rank test. All variables that achieved a significance at $P \le 0.05$ in univariate analyses were enrolled in multivariate Cox's proportional hazard model. $P \le 0.05$ was considered statistically significant unless otherwise specified.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

Authors' contributions

Chen Wu designed experiments, analyzed clinical data and wrote the manuscript; Bin Xu and You Zhou analyzed data; Mei Ji provided patient samples and analyzed clinical data. Dachuan Zhang performed research and analyzed data. Jingting Jiang and Changping Wu conceived the study, supervised experiments and revised the manuscript.

REFERENCES

- 1. Siegel RL, MillerKD, Jemal A. Cancer statistics, 2015. CA: a cancer journal for clinicians. 2015; 65:5–29.
- 2. Herbst RS, HeymachJV, Lippman SM. Lung cancer. The New England journal of medicine. 2008; 359:1367–1380.
- Campbell JD, Alexandrov A, Kim J, Wala J, Berger AH, Pedamallu CS, Shukla SA, Guo G, Brooks AN, Murray BA, Imielinski M, Hu X, Ling S, et al. Distinct patterns of somatic genome alterations in lung adenocarcinomas and squamous cell carcinomas. Nature genetics. 2016; 48:607–616.
- Huang T, Li J, Zhang C, Hong Q, Jiang D, YeM, Duan S. Distinguishing Lung Adenocarcinoma from Lung Squamous Cell Carcinoma by Two Hypomethylated and Three Hypermethylated Genes: A Meta-Analysis. PloS one. 2016; 11:e0149088.
- Zhan C, Yan L, Wang L, Sun Y, Wang X, Lin Z, Zhang Y, Shi Y, Jiang W, Wang Q. Identification of immunohistochemical markers for distinguishing lung adenocarcinoma from squamous cell carcinoma. Journal of thoracic disease. 2015; 7:1398–1405.
- Comprehensive molecular profiling of lung adenocarcinoma. Nature. 2014; 511:543–550.

- Gottschling S, Jauch A, Kuner R, Herpel E, Mueller-Decker K, Schnabel PA, Xu EC, Muley T, Sultmann H, Bender C, Granzow M, Efferth T, Hoffmann H, et al. Establishment and comparative characterization of novel squamous cell non-small cell lung cancer cell lines and their corresponding tumor tissue. Lung Cancer. 2012; 75:45–57.
- CoussensLM, Werb Z. Inflammation and cancer. Nature. 2002; 420:860–867.
- 9. Dinarello CA. A clinical perspective of IL-1beta as the gatekeeper of inflammation. European journal of immunology. 2011; 41:1203–1217.
- Vardeh D, Wang D, Costigan M, Lazarus M, Saper CB, Woolf CJ, FitzgeraldGA, Samad TA. COX2 in CNS neural cells mediates mechanical inflammatory pain hypersensitivity in mice. The Journal of clinical investigation. 2009; 119:287–294.
- Lee CH, Chang JS, Syu SH, Wong TS, Chan JY, Tang YC, Yang ZP, Yang WC, Chen CT, Lu SC, Tang PH, Yang TC, Chu PY, et al. IL-1beta promotes malignant transformation and tumor aggressiveness in oral cancer. Journal of cellular physiology. 2015; 230:875–884.
- 12. Guo J, Feng Z, Huang Z, WangH, Lu W. MicroRNA-217 functions as a tumour suppressor gene and correlates with cell resistance to cisplatin in lung cancer. Molecules and cells. 2014; 37:664–671.
- KasinskiAL, Slack FJ. miRNA-34 prevents cancer initiation and progression in a therapeutically resistant K-ras and p53induced mouse model of lung adenocarcinoma. Cancer research. 2012; 72:5576–5587.
- Bu P, Wang L, Chen KY, Srinivasan T, Murthy PK, Tung KL, Varanko AK, Chen HJ, Ai Y, King S, LipkinSM, Shen X. A miR-34a-Numb Feedforward Loop Triggered by Inflammation Regulates Asymmetric Stem Cell Division in Intestine and Colon Cancer. Cell stem cell. 2016; 18:189–202.
- Peng HY, Jiang SS, Hsiao JR, Hsiao M, Hsu YM, Wu GH, Chang WM, Chang JY, JinSC, Shiah SG. IL-8 induces miR-424–5p expression and modulates SOCS2/STAT5 signaling pathway in oral squamous cell carcinoma. Molecular oncology. 2016.
- Qayum AA, Paranjape A, Abebayehu D, Kolawole EM, Haque TT, McLeod JJ, Spence AJ, Caslin HL, Taruselli MT, Chumanevich AP, Baker B, OskeritzianCA, Ryan JJ. IL-10-Induced miR-155 Targets SOCS1 To Enhance IgE-Mediated Mast Cell Function. J Immunol. 2016; 196:4457–4467.
- McCoy CE, Sheedy FJ, Qualls JE, Doyle SL, Quinn SR, MurrayPJ, O'Neill LA. IL-10 inhibits miR-155 induction by toll-like receptors. The Journal of biological chemistry. 2010; 285:20492–20498.
- Wang L, Zhang LF, Wu J, Xu SJ, Xu YY, Li D, LouJT, Liu MF. IL-1beta-mediated repression of microRNA-101 is crucial for inflammation-promoted lung tumorigenesis. Cancer research. 2014; 74:4720–4730.
- Shi M. 6P Expression of PD-L1 in lung cancer cells regulates Treg cell differentiation. Journal of thoracic oncology. 2016; 11:S59.

- 20. Savas P, Salgado R, Denkert C, Sotiriou C, Darcy PK, SmythMJ, Loi S. Clinical relevance of host immunity in breast cancer: from TILs to the clinic. Nature reviews Clinical oncology. 2016; 13:228–241.
- Jung YJ, Isaacs JS, Lee S, TrepelJ, Neckers L. IL-1betamediated up-regulation of HIF-1alpha via an NFkappaB/ COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. FASEB journal. 2003; 17:2115–2117.
- Hou Z, Falcone DJ, SubbaramaiahK, Dannenberg AJ. Macrophages induce COX-2 expression in breast cancer cells: role of IL-1beta autoamplification. Carcinogenesis. 2011; 32:695–702.
- 23. Groen HJ, Sietsma H, Vincent A, Hochstenbag MM, van Putten JW, van den Berg A, Dalesio O, Biesma B, Smit HJ, Termeer A, Hiltermann TJ, van den BorneBE, Schramel FM. Randomized, placebo-controlled phase III study of docetaxel plus carboplatin with celecoxib and cyclooxygenase-2 expression as a biomarker for patients with advanced non-small-cell lung cancer: the NVALT-4 study. Journal of clinical oncology. 2011; 29:4320–4326.
- Liu R, Zheng J, Li C, Pang Y, Zheng Q, XuX, Liu P. Celecoxib induces epithelial-mesenchymal transition in epithelial ovarian cancer cells via regulating ZEB1 expression. Archives of gynecology and obstetrics. 2015; 291:1361–1369.

- Wang ZL, Fan ZQ, JiangHD, Qu JM. Selective Cox-2 inhibitor celecoxib induces epithelial-mesenchymal transition in human lung cancer cells via activating MEK-ERK signaling. Carcinogenesis. 2013; 34:638–646.
- Garzon R, MarcucciG, Croce CM. Targeting microRNAs in cancer: rationale, strategies and challenges. Nature reviews Drug discovery. 2010; 9:775–789.
- 27. Tili E, MichailleJJ, Croce CM. MicroRNAs play a central role in molecular dysfunctions linking inflammation with cancer. Immunological reviews. 2013; 253:167–184.
- 28. Jin P, Wang E, Ren J, Childs R, Shin JW, Khuu H, MarincolaFM, Stroncek DF. Differentiation of two types of mobilized peripheral blood stem cells by microRNA and cDNA expression analysis. Journal of translational medicine. 2008; 6:39.
- Vaz C, Ahmad HM, Sharma P, Gupta R, Kumar L, KulshreshthaR, Bhattacharya A. Analysis of microRNA transcriptome by deep sequencing of small RNA libraries of peripheral blood. BMC genomics. 2010; 11:288.
- Weber DG, Casjens S, Rozynek P, Lehnert M, Zilch-Schoneweis S, Bryk O, Taeger D, Gomolka M, Kreuzer M, Otten H, Pesch B, JohnenG, Bruning T. Assessment of mRNA and microRNA Stabilization in Peripheral Human Blood for Multicenter Studies and Biobanks. Biomarker insights. 2010; 5:95–102.