Epithelial-to-mesenchymal transition, circulating tumor cells and cancer metastasis: Mechanisms and clinical applications

Xiao-Xiang Jie^{1,2,3}, Xiao-Yan Zhang^{1,2,3} and Cong-Jian Xu^{1,2,3,4}

¹Obstetrics and Gynecology Hospital, Fudan University, Shanghai 200011, People's Republic of China

²Department of Obstetrics and Gynecology of Shanghai Medical School, Fudan University, Shanghai 200032, People's Republic of China

³Shanghai Key Laboratory of Female Reproductive Endocrine Related Diseases, Shanghai 200011, People's Republic of China

⁴Institute of Biomedical Sciences, Fudan University, Shanghai 200032, People's Republic of China

Correspondence to: Xiao-Yan Zhang, email: xyzhang80@gmail.com Cong-Jian Xu, email: xucongjian@gmail.com

Keywords: epithelial-to-mesenchymal transition, circulating tumor cells, metastasis, EMT markers

Received: March 29, 2017 **Accepted:** May 10, 2017 **Published:** May 26, 2017

Copyright: Jie et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0),

which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Epithelial-to-mesenchymal transition (EMT) endows epithelial cells with enhanced motility and invasiveness, allowing them to participate in many physiological and pathological processes. Epithelial-to-mesenchymal transition contributes to the generation of circulating tumor cells (CTCs) in epithelial cancers because it increases tumor cell invasiveness, promotes tumor cell intravasation and ensures tumor cell survival in the peripheral system. Although the contribution of epithelialto-mesenchymal transition to tumor cell invasiveness has been confirmed, the role epithelial-to-mesenchymal transition plays in metastasis remains debated. As a favorable material for a "liquid biopsy", circulating tumor cells have been shown to have promising values in the clinical management of tumors. Furthermore, an increasing number of studies have begun to explore the value of CTC-related biomarkers, and some studies have found that the expression of EMT and stemness markers in circulating tumor cells, in addition to CTC detection, can provide more information on tumor diagnosis, treatment, prognosis and research.

INTRODUCTION

Epithelial cancer metastasis is a multi-step process that includes a loss of intercellular connections, the invasion of basal membrane and surrounding tissues, intravasation into venous or lymphatic vessels that generates circulating tumor cells (CTCs), survival in the peripheral system, extravasation and proliferation at secondary sites. In this metastatic cascade, epithelialto-mesenchymal transition (EMT) is believed to play an important role [1–3]. EMT is characterized by decreased epithelial properties and increased mesenchymal attributes and has been implicated in a number of physiological processes, such as embryonic development and pathological conditions, including organ fibrosis and cancer progression [2–4].

One important aspect of EMT's role in cancer is that EMT contributes to the generation of CTCs. CTCs are

tumor cells released into blood and/or lymphatic vessels that can circulate in the human body, which are predestined sources of metastasis as the "seeds" in Paget's "seed and soil" hypothesis [5]. Moreover, EMT has been shown to contribute to tumor resistance [6–8] and may also be related to tumor cell stemness [9, 10]. However, because of technical restrictions and a lack of convincing *in vivo* evidence, the role of EMT in cancer metastasis remains debated. Some believe that EMT is of great importance in the formation of metastases, whereas others hold that the effect of EMT might have been overestimated, given the lack of convincing evidence. In fact, some have suggested that EMT is not required for metastasis [6, 8, 11–13].

Because of their crucial role in metastasis, CTCs have had attracted much attention since their discovery. Furthermore, the development of CTC detection technology has facilitated the real-time dynamic monitoring of cancer by using CTCs as a material for "liquid biopsies", which are superior to traditional biopsy [14, 15]. In the last decade, the detection and enumeration of CTCs have been shown to provide information on prognosis, metastasis, therapeutic efficacy and chemoresistance in several cancer types [16–20]. Recent studies of CTCs are not limited to the detection and enumeration of these cells, and increasing efforts have been made to elucidate the molecular features of CTCs and the potential values of these biomarkers, among which EMT markers are of great interest.

In this review, we summarize the mechanisms underlying EMT and their role in CTC generation, shed light on the controversial role of EMT in metastasis, and review recent advances that have been made in the clinical application of EMT markers in CTC detection.

THE MECHANISMS UNDERLYING EMT AND THEIR ROLE IN METASTASIS

The mechanisms underlying EMT

Epithelial-to-mesenchymal transition (EMT) is a multi-step process involving many molecular and cellular changes, including the down-regulation of epithelial proteins such as E-cadherin, claudins and cytokeratins and the up-regulation of mesenchymal proteins, such as N-cadherin, fibronectin and vimentin, which endow the cell with increased motility and invasiveness [1–3, 21].

These molecular changes during EMT are regulated by transcription factors called EMT-inducing transcription factors (EMT-TFs), including Snail 1, Snail 2 (Slug), ZEB1, Twist, TCF4, and FOXC2 [22, 23]. In addition to EMT-TFs, some extracellular molecules in the tumor microenvironment (TGF- β , FGF, EGF, HGF, Wnt, Notch, Hedgehog, etc.) and related pathways (MAPK, PI3K, NF- κ B, Wnt/ β -catenin, Notch, etc.) are thought to induce EMT [23–27]. Moreover, hypoxia results in the accumulation of hypoxia-inducible factor (HIF), and HIF-1 α activates EMT-TFs, such as Twist and Snail, to induce EMT [28, 29] (Figure 1).

EMT plays a role in a number of physiological processes and pathological conditions. EMT has been categorized into three types based on function [2]: type I is involved in embryonic development, type II participates in wound healing and fibrosis, and type III is associated with cancer progression, which is the focus of this review.

EMT promotes CTC generation

The generation of CTCs includes several steps: the detachment from the tumor mass, invasion of the basal membrane and surrounding tissues, entry of vessels and survival in the peripheral system. Specifically, EMT and related regulatory networks primarily promote CTC generation in three aspects: i) increase tumor cell

invasiveness, ii) promote tumor cell intravasation and iii) facilitate tumor cell survival in the peripheral system. (Figure 1)

As shown in Figure 1, the EMT regulatory networks involves not only molecular changes directly regulated by EMT-TFs but also other related factors and pathways, such as HIF, TGF- β and VEGF-A, which together promote the ultimate generation of CTCs.

Epithelial cells are immobile due to the precise regulation of strong cell-cell and cell-extracellular matrix adhesions, which consist of adherent junctions, tight junctions and desmosomes, and the well-constructed cytoskeleton. During EMT, the key components of intercellular junctions, such as E-cadherin, claudins, occludins and desmosomes, are directly down-regulated by EMT-TFs, such as Snail, Slug and SIP1 [22, 30-32]. Some cell and animal assays also demonstrated that the reorganization of adhesive molecules is associated with increased invasiveness [33-35]. The phenotype switch from epithelial to mesenchymal during EMT also involves the reconstruction of the cytoskeleton, which changes their morphology to a spindle-like shape that is appropriate for migration [36]. In addition to adhesive molecules, EMT-TFs can also induce the expression of matrix metalloproteinases (MMPs), which facilitate the degradation of the basal membrane and surrounding tissues [37–39]. Remarkably, some important extracellular factors, such as TGF- β , FGF and Wnt, participate in the regulatory network by inducing EMT and/or the expression of MMPs [23–27, 29, 40, 41]. The mechanisms of hypoxia in this regulatory network are similar to those of extracellular factors, as shown in Figure 1 [28, 29, 42, 43].

The EMT regulatory network also promotes angiogenesis and facilitates cancer cell intravasation. Specifically, EMT-TFs such as Snail and Slug, can promote blood vessel formation by inducing the expression of vascular endothelial growth factor A (VEGF-A) in subcutaneous xenograft tumor models [44–46]. Other factors in the regulatory network, such as Notch and HGF, can also promote angiogenesis via a similar mechanism [47-49]. Remarkably, VEGF and HIF1- α were also expressed on CTCs [50]. Moreover, EMT-induced and TGF- β -induced proteases, especially MMPs, can also promote angiogenesis and intravasation [51, 52]. Moreover, newly formed, tumor-associated vessels are often malformed and leaky, which facilitates tumor cell invasion [53]. Specifically, EMT-TFs or EMT-related factors have been shown to enhance transendothelial migration, which further supports that EMT promotes tumor cell intravasation [54, 55].

CTCs do not easily survive in the peripheral system because they may encounter strong anoikis signals and chemotherapy or radiotherapy. Nevertheless, EMT can facilitate tumor cell survival in the peripheral system by allowing cancer cells to avoid apoptosis, anoikis, and senescence and promote drug resistance [56, 57]. EMT-TFs, such as Snail, Slug, Twist and SIP1, can protect CTCs from anoikis by disturbing normal apoptotic cascades, resisting senescence and/or cooperating with TrkB [58–62]. EMT-TFs can reportedly endow tumor cells with resistance to chemotherapy and radiotherapy in several cancer types [6, 8, 63]. For example, Snail and Slug directly contribute to cisplatin resistance in ovarian cancer [64, 65]. Moreover, the inhibition of EMT can restore the chemosensitivity, indicating that the EMT-TFs may be a potential target for the treatment of therapy resistance [7, 66, 67].

Remarkably, many regulatory loops exist between the components of the EMT regulatory network [43, 45, 68–71], as shown in Figure 1. For example, EMT-TFs can induce the expression of MMPs, whereas some proteases can reverse EMT [69, 70]. VEFG-A plays a similar role in EMT [45, 71]. In summary, the EMT-related regulatory network is an interactive, integrated and precisely regulated network due to the association between EMT, extracellular factors (TGF- β , FGF, Notch, etc.) and hypoxia, which are all important aspects in cancer progression. As the central part of the regulation network, EMT, accompanied by the related factors and pathways, plays an important role in the generation of CTCs by promoting cell invasion, angiogenesis, intravasation, therapy resistance and survival.

EMT's role in metastasis

The contribution of EMT to tumor cell invasiveness and CTC generation has been confirmed, but its role in the metastatic cascade remains debated. Two metastatic models involving EMT have been proposed (Figure 2). The first and most widely known model proposes that cancer cells must first undergo EMT to become invasive and generate CTCs and then undergo the reverse of EMT, MET (mesenchymal-to-epithelial transitions), to restore epithelial properties after extravasation into secondary sites and facilitate metastatic growth [72–75]. This process of EMT and subsequent MET is very common and important

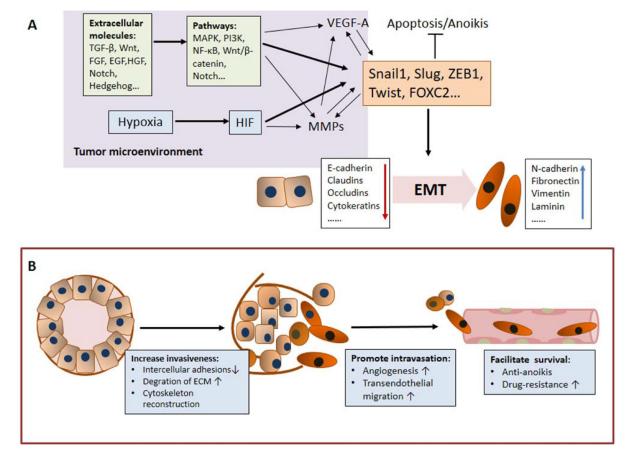


Figure 1: The mechanisms underlying the role of EMT in CTC generation. (A) The EMT-related regulatory network. EMTinducing transcription factors (EMT-TFs), including Snail 1, Snail 2 (Slug), ZEB1, and Twist, play a central role in this network and regulate molecular changes during EMT. Some important extracellular molecules in the tumor microenvironment, such as TGF β , HGF, FGF, Wnt and Notch, bind to their respective receptors to induce EMT and are consequently also important components in the EMT regulatory network. Hypoxia, a significant aspect in cancer progression, triggers EMT and participates in the EMT regulatory network. Notably, the EMT regulatory network is an interactive, integrated and precisely regulated network that is involved in the generation of CTCs. (**B**) EMT promotes CTC generation by increasing tumor cell invasiveness, promoting tumor cell intravasation and facilitating tumor cell survival in the peripheral system.

during embryonic development and some postnatal gland development. Thus, this process can be expected to play a role in cancer progression [73, 76]. Specifically, this EMT/MET metastatic model can explain the fact that the histopathological features of secondary tumor sites resemble those of the primary site [73]. Furthermore, the switch between EMT and MET and phenotypic plasticity have been reported in some cancer types [77, 78]. For example, Chao et al. have reported that E-cadherin was reexpressed due to MET and may play a role in promoting cell survival at metastatic sites [79, 80]. Although clinical studies have indicated that MET occurs in distant sites, a lack of in vivo evidence of MET at distant sites reduces the credibility of this model. In recent years, increasing evidence from both in vitro and in vivo experiments has supported this metastatic model. For example, Chaffer et al. used a series of bladder cancer cell lines to study the role MET in the metastasis cascade and found that reversion to epithelial characteristics in a mesenchymal-like cancer cell line, i.e., regaining an epithelial phenotype, was favorable in the latter stages of the metastatic cascade; this finding confirmed the suspected role of MET in secondary cancer growth [81, 82]. Moreover, Banyard, J and colleagues used an in vivo cycling strategy to select metastatic cancer cells from the lymph nodes of mice bearing orthotopic DU145 human prostate tumors and observed a shift to an epithelial phenotype in progressive lymphatic cancer cells, providing evidence for spontaneous MET *in vivo* [83]. Ocana, O and colleagues induced MET by silencing Prrx1, an EMT inducer, and found that the loss of Prrx1 was required for migratory cells to colonize a secondary organ *in vivo* [84]. Tsai, JH et al. came to a similar conclusion: they found that turning off Twist 1, an important EMT-TF, to allow the reversion of EMT was essential for cancer cells to form metastases in distant sites [85].

However, the classic EMT/MET model has been challenged in recent years [11–13]. Although the importance of EMT in endowing cancer cell with enhanced invasiveness and motility has been confirmed, the role of MET in the metastatic cascade remains debated. Increasing evidence suggests that MET plays a role in the latter stages of the metastatic cascade, but some researchers question the necessity of MET to complete colonization at distant sites. Based on the concept of "collective" or "cohort" migration [86], they introduced the second metastatic model discussed in this review: epitheliallike and mesenchymal-like cancer cells can cooperate with each other during collective migration to achieve metastasis instead of undergoing the difficult process of EMT and subsequent MET [86, 87]. EMT is not an on/

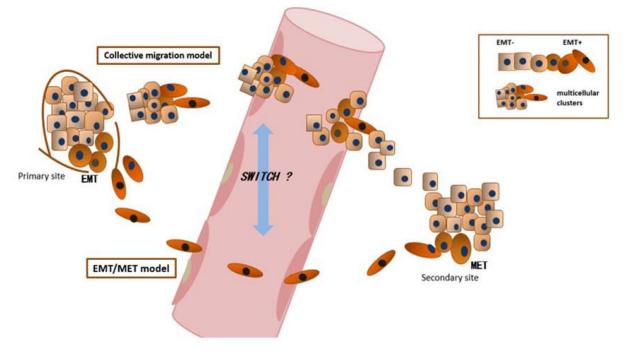


Figure 2: EMT and metastatic models. EMT/MET model: epithelial cancer cells must first undergo EMT to become invasive and motile and generate CTCs; CTCs circulate around the body and extravasate to distant sites; after extravasation to secondary sites, cancer cells must undergo the reverse process of EMT, MET (mesenchymal-to-epithelial transition) to restore epithelial properties, allowing them to ultimately colonize distant sites and form metastases. Collective migration model: instead of migrating as a single cell, cancer cells that have undergone various degrees of EMT coexist as multicellular clusters and migrate collectively, with the more motile invasive mesenchymal-like cells aggregating at the invasive front of multicellular clusters to "pave" the way, whereas epithelial-like cells retain their epithelial properties, follow behind and seize the opportunity to proliferate and colonize at distant sites after extravasation. Notably, the EMT/MET model and collective migration model may be not independent or diametrically opposed. Tumor cells may switch between the two mechanisms under certain circumstances, or the two mechanisms may synergistically effect metastases.

off switch, and the phenotypic transition from epithelial to mesenchymal in cancer cells consequently would give rise to many hybrid phenotypes that possess the properties of both cell types [88]; this phenotype plasticity facilitates collective migration. The collective migration model hypothesizes that many hybrid phenotypes coexist as multicellular clusters and migrate collectively, with the more motile invasive mesenchymal-like cells aggregating at the invasive front of multicellular clusters to "pave" the way, whereas epithelial-like cells follow behind and seize the opportunity to proliferate and colonize distant sites after extravasation [87]. Accordingly, multicellular cancer fragments, so-called tumor micro-emboli or CTC clusters, have long been observed in the clinic and correlate with distant metastasis [13, 89]. Moreover, several in vivo studies support this model. For example, Tsuji and colleagues established an animal model by subcutaneously and intravenously injecting EMT-derived cells and non-EMT cells, respectively, and found that EMT-derived cells were able to invade the blood stream but failed to establish lung metastases, whereas non-EMT cells could not invade but were able to establish lung metastases when injected intravenously. Surprisingly, lung metastases formed when a mixture of EMT-derived and non-EMT cells were co-inoculated subcutaneously, suggesting cooperation between epithelial and mesenchymal cancer cells during metastasis [90, 91]. Banyard, J and colleagues also reported that the epithelial-like DU145-LN4 cell line invaded in a collective migration pattern into lymphatic vessels in their model [92]. Fischer, KR et al. established an in vivo EMT lineage tracing system to monitor EMT in a triple-transgenic mouse model, in which breast-to-lung metastasis occurred spontaneously and EMT was "visible", if it occurred. Notably, lung metastases mainly consisted of non-EMT cancer cells, which maintained epithelial phenotype during metastasis in their study, contradicting the original EMT/MET hypothesis [6]. Surprisingly, when they inhibited EMT by overexpressing miR-200, lung metastases were not affected, leading them to conclude that EMT was not required for lung metastasis [6]. Zheng, XF drew a similar conclusion: the suppression of EMT by deleting Snail or Twist in genetically engineered mouse models of pancreatic ductal adenocarcinoma (PDAC) did not affect the development of metastases [8], which also suggested that EMT does not play a role in metastasis. However, the contribution of EMT to the metastatic cascade should not yet be discounted because EMT is complex and precisely controlled. Therefore, the effects of EMT in vivo cannot be easily and completely eliminated. And the role of partial EMT may have been underestimated in these aforementioned models.

Notably, although the classic EMT/MET model and collective migration model are conflicting, these two models both state that the mesenchymal phenotype is more invasive and motile than the epithelial phenotype and that the epithelial phenotype favors proliferation compared with the mesenchymal phenotype. Thus, the assumption that the tumor takes advantage of the mesenchymal phenotype in the early stage of the metastatic cascade, when invasion is the main concern, and then exploits the proliferative epithelial phenotype to complete colonization is reasonable. The classic EMT/ MET model hypothesizes that cancer cells must undergo this phenotype transition twice, i.e., EMT and subsequent MET, to ensure the optimal phenotype in each stage of the metastatic cascade. Thus, EMT plays a central role in metastasis. Conversely, collective migration model cleverly circumvents these troublesome phenotype transitions and proposes that epithelial and mesenchymal phenotypes cooperate to accomplish metastases. Thus, EMT assists the metastatic cascade in this model. In fact, Chaffer et al., who support the EMT/MET model [82], and Tsuji et al., who back the collective migration model [90], reported similar but slightly different results, which has led to disagreement. Currently, identifying the correct model may be difficult because both are supported by an increasing body of evidence. Specifically, both singlecell motility and collective migration are observed in the circulation. Interestingly, Giampieri, S et al. found that TGF β -1 acted as a switch between the cohesive phenotype and single cell motility via a transcriptional program, and collective migration seems more practical in lymphatic metastasis than blood-borne metastasis [93]; however, this finding warrants further confirmation. Banyard, J also hypothesized that EMT may not be essential during lymphatic metastasis [75]. Thus, the EMT/MET model and/or collective migration model may not be independent or diametrically opposed; the two models may co-exist and synergistically facilitate the metastatic cascade. Tumor cells may switch between the two mechanisms under certain circumstances or prefer a mechanism for a specific type of metastasis to maximize their colonization of distant sites.

CLINICAL APPLICATIONS OF EMT MARKERS IN CTCS DETECTION

Circulating tumor cells are highly heterogeneous, and the molecular features of CTC often differ by subpopulation [94–96]; thus, these subpopulations may play different roles in cancer progression. A better understanding of CTC heterogeneity has demonstrated that the molecular features of different CTC subpopulations need to be studied to identify the "evil" CTC subpopulations that are responsible for lethal cancer progression. Therefore, an increasing number of studies no longer focus on simple CTC enumeration and have begun to explore the genotypic and phenotypic characterization of CTCs. For example, researchers have investigated the value of HER 2 expression on CTCs in patients with breast cancer [97, 98], AR gene status of CTCs in prostate cancer [99], EGFR mutations in lung cancer and KRAS mutations in colorectal cancer [100, 101]. Besides these cancer type specific biomarkers, EMT markers can be promising biomarkers of CTCs in different cancers. EMT can endow cells with enhanced invasiveness and motility, drug-resistance, and even stemness properties [2, 30], the CTC subpopulations positive for EMT markers may be responsible for cancer progression [76, 96]. Thus, delineating the expression of EMT markers in CTCs is of great clinical interest.

EMT is believed to generate CTCs with stem cell properties [9], and some studies reported that the overexpression of EMT markers in CTCs is often accompanied by the expression of stem cell markers, such as ALDH 1 and CD133 [102–104]. Below, we review the expression of EMT markers in CTCs and focus on their clinical applications while also examining stem cell markers (Supplementary Table 1).

EMT markers

The molecular changes during EMT are characterized by the down-regulation of epithelial proteins and up-regulation of mesenchymal proteins, and these changes are regulated by EMT-TFs and related pathways, as stated above. Thus, we can categorize the EMT markers into three types: i) epithelial makers, ii) mesenchymal markers and iii) regulators (Table 1).

Epithelial markers are molecular biomarkers that are highly expressed in epithelial cells but not expressed or expressed at low levels in mesenchymal cells, such as EpCAM, E-cadherin, cytoketatins, and Zonula occludins (ZO). Epithelial markers are often used to detect CTCs and ensure their epithelial origin. Epithelial cellular adhesion molecule (EpCAM) is an adhesive molecule between epithelial cells and believed to be involved in epithelial malignancies [105]. EpCAM has long been widely used to detect and enumerate CTCs [106, 107], but EpCAM expression is down-regulated if the cancer cells have undergone EMT because it is an epithelial marker. Thus, the use of EpCAM for CTC detection is likely to exclude CTCs that have undergone EMT [108]. Cytokeratins (CKs) are a group of intermediate filaments that comprise the cytoskeleton and have also been used to detect CTCs. Like EpCAM, CKs are down-regulated during EMT and may be not appropriate for CTC detection when CTCs have undergone EMT [108]. Moreover, E-cadherin is an essential component of adherent junctions, and ZOs are proteins of tight junction, which are both involved in the maintenance of epithelial intercellular adhesions and are down-regulated during EMT [109, 110].

Although mesenchymal markers, such as N-cadherin, vimentin, and fibronectin, are highly expressed in mesenchymal cells expressed at low levels in epithelial cells, they are up-regulated during EMT, and the expression of mesenchymal markers by CTC can reflect their EMT status. N-cadherin and E-cadherin both belong to the cadherin family. During EMT, N-cadherin is overexpressed, whereas E-cadherin is downregulated—this switch from E-cadherin to N-cadherin is a hallmark of CTCs that have undergone EMT [111]. Vimentin is a component of intermediate filaments whose expression facilitates the reconstruction of the cancer cell cytoskeleton during EMT and endows the cancer cell with a spindle-like shape suitable for migration [112, 113].

EMT-TFs and their related pathways regulate the molecular changes during EMT, and these elements themselves can also be markers of EMT, such as Twist, Snail, ZEB and Akt/PI3K. Twist 1 is a transcription factor that acts on the E-box of E-cadherin to down-regulate the expression of E-cadherin, which is associated with cancer progression and a useful marker to evaluate the EMT status of CTCs [114]. The mechanisms of Snail 1, Slug and ZEB 1 and their role in EMT are similar to that of Twist [22]. The PI3K/AKT/m TOR pathway is an important regulator of the cell cycle and related to proliferation, cancer and EMT [115]. As the central elements in this pathway, PI3K and Akt have already been used in some studies as mesenchymal markers of CTCs [102, 116].

The detection of these EMT markers in CTCs would help to evaluate their EMT status and distinguish CTC subpopulations.

The expression of EMT markers in CTCs

In recent years, an increasing number of studies have attempted to detect EMT markers in CTCs. For example, Kasimir-Bauer, S et al. studied three EMT markers (Twist 1, Akt2, and PI3Ka) and one stemcell marker (ALDH 1) in CTCs from 502 patients with primary breast cancer and detected at least one of the EMT markers and ALDH 1 in up to 72% and 46% of the CTC-positive group, respectively [102]. Kallergi, G et al. reported that over 80% of patients with both early and metastatic breast cancer harbored CTCs expressing phospho-Akt and phsopho-PI3K [117]. Moreover, Aktas, B et al. detected EMT markers and stem cell markers in 62% and 69% of the CTC-positive patients with metastatic breast cancer, respectively [103], and Li, YM et al. showed that the 80.4% and 84.8% of CTC isolated from patients with hepatocellular carcinoma express vimentin and Twist, respectively [118]. Similar findings have been reported in colorectal cancer, pancreatic cancer and prostate cancer [119–122].

Based on these data, we can conclude that EMT and stem cell markers are frequently overexpressed in CTCs, irrespective of cancer type. Remarkably, in addition to detecting EMT markers and ALDH 1 in the CTC-positive group, Kasimir-Bauer detected these respective markers in 18% and 5% of the CTC-negative group [102]. Aktas, B also reported the expression of EMT markers and stem cell markers in the CTC-negative group [103]. Gradilone, A et al. reported similar findings [123]. The three studies all relied on EpCAM-based capturing methods, i.e., AdnaTest

Categorization	Marker	Features
Epithelial markers	ЕрСАМ	Down-regulated during EMT.
	E-cadherin	Often used to detect CTCs.
	Cytokeratins (CK)	
	Zonula occludins (ZO)	
Mesenchymal markers	N-cadherin	Highly expressed in mesenchymal cells. The switch fron E-cadherin to N-cadherin is a hallmark of EMT.
	Vimentin	Highly expressed in mesenchymal cells, induce mesenchymal morphology.
	Fibronectin	Regulates cell shape.
Regulators	Twist 1	bHLH factor, represses E-cadherin expression.
	Snail 1	Zinc-finger protein, directly represses E-cadherin expression
	Snail 2 (Slug)	
	ZEB 1/ZEB 2	Zinc-finger E-box-binding homeobox protein, transcriptiona repressor.
	Akt and PI3K	The PI3K/AKT/m TOR pathway is associated with proliferation and EMT.
	FoxC 2	Transcriptional activator, induces EMT.

Table 1: The categorization of EMT markers

and CELLection, to detect CTC and consequently may have missed CTC subpopulations that had undergone EMT, indicating that epithelial marker-based (such as EpCAM) detection methods may be unsuitable for detecting EMT CTCs. This notion is supported by several studies showing that combining EMT markers with epithelial markers increases the CTC detection rate compared with the AdnaTest approach [124].

The expression of EMT markers in CTC correlates with tumor stage and metastasis

Kallergi, G et al. investigated the expression of the EMT markers Twist 1 and vimentin in CTCs from 25 patients with metastatic and 25 patients with early breast cancer and identified Twist 1(+) and vimentin(+)CTCs in 73% and 77% of patients with early breast cancer, respectively, whereas CTCs from all patients with metastatic breast cancer were positive for these markers [125]. Papadaki, M.A et al. also detected high ALDH1 expression (ALDH 1^{high}) and nuclear Twist (Twist^{nuc}) CTCs in 80% of patients with metastatic breast cancer but only 30.8% of patients with early breast cancer [126]. Moreover, Gradilone, A et al. evaluated the expression of vimentin, fibronectin and ALDH 1 in CTCs from 92 female patients with breast cancer and found that 91% (10/11) of the triple-positive samples (ALDH1/ vimentin/ fibronectin) were isolated from patients with stage IV disease, and the expression of ALDH 1 by CTCs significantly correlated with the stage of disease [123]. Li, Y.M and colleges confirmed the correlation between Twist/ vimentin expression and portal vein tumor thrombus or cancer stage in hepatocellular carcinoma, but they failed to find similar positive results with EMT-TFs, ZEB1, ZEB2 and Snail [118]. Nevertheless, Alonso-Alconada L et al. found that ZEB1 expression correlated with lymph node metastasis in endometrial cancer [127]. Similarly, Kulemann B reported this relationship in pancreatic ductal adenocarcinoma (PDCA) [120]. Although several meaningful findings have been made in several cancer types, Kasimir-Bauer, S et al. did not identify a correlation between EMT marker expression and cancer stage in their study [102].

The above studies suggest that the expression of EMT and stem cell markers in CTC may indicate a later stage and more aggressive disease but the clinical relevance of these findings needs to be further evaluated in more prospective trials.

The expression of EMT markers in CTC correlates with therapeutic response

Aktas, B and colleges evaluated the expression of EMT and stem cell markers in CTCs isolated from patients with breast cancer during follow-up and detected EMT marker and ALDH1 expression in only 10% and 5% of responders, respectively, whereas these percentages were much higher in non-responders (62% and 44%, respectively) [103]. Mego, M et al. also reported that EMT marker positivity correlated with a lack of response to therapy and relapse [129]. Satelli, A et al. defined a threshold of 5 EMT CTCs to reflect therapeutic response in patients with metastatic colon cancer who received adjuvant chemotherapy after surgery: the patients with < 5 EMT CTCs responded to therapy, whereas patients with ≥ 5 EMT CTCs tended to experience disease progression [134]. Moreover, Chang K et al. found that stem cell marker expression indicated a worse response rate to chemotherapy in metastatic castration-resistant prostate cancer (mCRPC), but similar correlations with EMT markers were not identified [128]. Similarly, Polioudaki H et al. also failed to identify a correlation between EMT marker expression and therapeutic response in metastatic breast cancer [130].

These studies suggest that EMT and stem cell markers may be negatively correlated with therapeutic response, and the detection of EMT and stem cell markers in CTC may consequently be more convenient and faster than conventional methods to assess therapy response and help doctors manage patients with cancer better. However, the utility of EMT and stem cell markers in assessing therapeutic response needs to be fully confirmed and strictly standardized prior to clinical application.

The expression of EMT markers in CTC correlates with prognosis

Lindsay, CR and colleges found that overall survival (OS) was significantly longer in patients with mCRPC whose CTCs were positive for the EMT marker vimentin than in patients whose CTCs lacked vimentin expression (453 days vs 305 days) [121]. Furthermore, Polioudaki H et al. detected keratin expression in CTCs from patients with metastatic breast cancer and found that low keratin expression correlated with worse OS [130]. Likewise, Ning, Y et al. showed that progression-free survival (PFS) (3.0 vs 7.7 months) and OS (10.0 vs 26.8 + months) were significantly shorter in patients with metastatic colorectal cancer with CTCs expressing ALDH1, PI3a and/or Akt-2 [119]. Mego, M also identified a correlation between EMT marker positivity and prognosis in patients with metastatic breast cancer [129]. Although an increasing number of studies have indicated that EMT marker-positive CTCs are valuable for prognosis, some studies failed to obtain similar results. Chang K et al. found that the expression of stem cell markers indicated poor prognosis in mCRPC patients, but EMT marker expression failed to show any prognostic value [128]. Moreover, Kulemann B's study of pancreatic ductal adenocarcinoma indicated a prognostic value for the KRAS mutation in CTCs but not the EMT marker ZEB1 [120]. Kasimir-Bauer, S et al. also failed to correlate the expression of EMT markers in CTCs with prognosis [102].

These studies indicate that the expression of EMT and stem cell markers may correlate with poor prognosis, but their prognostic value has to be further evaluated in additional clinical trials.

Classifying CTCs based on the expression of EMT markers

EMT is likely to result in many hybrid phenotypes of CTCs that possess both epithelial and mesenchymal features because the extent of EMT may differ among CTCs [88]. Thus, in addition to the information provided by the detection of EMT markers in CTCs on cancer stage, treatment response and prognosis, some studies have attempted to classify CTCs based on EMT status to research the potential value of CTC subpopulations and further explore the relationship between the EMT in CTCs and cancer progression.

After detecting the expression levels of keratin and vimentin in breast cell lines and patients with breast cancer, Polioudaki H and colleges introduced a novel parameter, the vimentin/keratin ratio (Vim/K ratio), to reflect the EMT status of CTCs: a low Vim/K ratio suggested an epithelial phenotype, whereas a high Vim/K ratio indicated a mesenchymal phenotype in CTCs. Based on the Vim/K ratio of CTCs, they classified the CTCs into epithelial, biophenptypic and mesenchymal CTCs [130]. Moreover, Nel, I et al. counted the CK-positive, vimentinpositive and N-cadherin-positive CTCs and calculated the vimentin+ cells/CK+ cell ratio and N-cadherin+ cells/ CK+ cell ratio in patients with hepatocellular carcinoma. They found that a change in the ratio of epithelial to mesenchymal CTCs was associated with a longer median time to progression (TTP) (1 vs 15 months), and the N-cadherin/CK ratio significantly correlated with cirrhosis [135]. Furthermore, Liu, YK et al. evaluated the expression of several EMT markers, including CK8/18/19, EpCAM, vimentin and Twist, in the CTCs of patients with hepatocellular carcinoma patients and classified CTCs into epithelial, biophenptypic and mesenchymal phenotypes based on the expression of EMT markers, which is the approach used by most studies to classify CTCs. Liu and colleges further studied the potential values of different CTC phenotypes and found that the number of epithelial CTCs was related to tumor size; biophenotypic CTCs were related to tumor number and mesenchymal CTCs were associated with metastasis [131]. Similarly, Zhao, R et al. analyzed the CTCs of patients with colorectal cancer and found that both biophenotypic and mesenchymal CTCs, but not epithelial CTCs, correlated with a late clinical stage, lymph node metastasis and distant metastasis, suggesting that CTCs with mesenchymal properties denote more aggressive disease and metastatic potential [132]. Li, TT classified CTCs from patients with gastric cancer into five types based on a similar principle and observed that the proportion of mesenchymal CTCs in the post-treatment blood specimens increased in a patient who experienced disease progression [133]. Accordingly, Yu, M et al. followed the EMT status of CTCs in patients with breast cancer and found that the EMT status of CTCs dynamically changed in response to therapy and disease progression [136], suggesting that the EMT status of CTCs may be used to monitor therapy response and cancer progression [133, 136].

CONCLUSIONS AND PROSPECTIVE

EMT promotes the generation of circulating tumor cells (CTCs) in epithelial cancers by increasing tumor cell invasiveness and motility to promote tumor cell intravasation and facilitate tumor cell survival in the peripheral system. Although the contribution of EMT to tumor cell invasiveness has been confirmed, its role in metastasis remains debated. Two metastatic models involving EMT have been proposed: the classic EMT/MET model, in which EMT comprises the core of the metastatic cascade, and the collective migration model, in which EMT assists the early stage of the metastatic cascade, as discussed above. The metastatic cascade is a complex process, and its individual steps need to be elucidated before we can determine which of these models is correct. Based on limited current evidence, we herein hypothesized that the two models mentioned above may co-exist and synergistically facilitate the metastatic cascade; tumor cells may switch between mechanisms to maximize the rate of metastasis. However, several areas remain to be explored: the validity of the co-existence of the two models, the circumstances inducing cells to switch between motility patterns, and the routes of metastasis favored by each model. To this end, better-designed assays that can distinguish the two models and truly delineate each step of the metastatic cascade are needed. In conclusion, the extent to which EMT is necessary and the model responsible for the metastatic cascade remain to be determined.

Liquid biopsy is undoubtedly superior to conventional methods for dynamically monitoring cancer status, and the detection of CTCs is likely to gain popularity in the clinic. As a supplement to CTC enumeration, evaluating the expression of EMT and stem cell markers in CTCs may provide information about the tumor stage, metastasis, therapeutic response and prognosis, and using EMT markers to classify CTCs can elucidate CTC heterogeneity. However, standard and optimized approaches are lacking and studies of CTCs generally suffer from small sample sizes. Therefore, larger well-designed clinical trials are needed to further illuminate the potential values of EMT markers in CTCs.

ACKNOWLEDGMENTS AND FUNDING

This work was supported by funding from Key Project for Major Diseases of Health System of Shanghai Municipality (2013ZYJB0201), National Key Research and Development Program (2016YFC1303100) and National Natural Sciences Foundation of China (81472424).

CONFLICTS OF INTEREST

None.

REFERENCES

- 1. Thiery JP. Epithelial–mesenchymal transitions in tumour progression. Nat Rev Cancer. 2002; 2:442–454.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelialmesenchymal transitions in development and disease. Cell. 2009; 139:871–890.
- 3. Kalluri R, Weinberg RA. The basics of epithelialmesenchymal transition. J Clin Invest. 2009; 119:1420–1428.
- 4. De Wever O, Pauwels P, De Craene B, Sabbah M, Emami S, Redeuilh G, Gespach C, Bracke M, Berx G. Molecular and pathological signatures of epithelial-mesenchymal transitions at the cancer invasion front. Histochem Cell Biol. 2008; 130:481–494.
- 5. Gupta GP, Massague J. Cancer metastasis: building a framework. Cell. 2006; 127:679–695.
- Fischer KR, Durrans A, Lee S, Sheng J, Li F, Wong STC, Choi H, El Rayes T, Ryu S, Troeger J, Schwabe RF, Vahdat LT, Altorki NK, et al. Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. Nature. 2015; 527:472–476.
- Zhuo W, Wang Y, Zhuo X, Zhang Y, Ao X, Chen Z. Knockdown of Snail, a novel zinc finger transcription factor, via RNA interference increases A549 cell sensitivity to cisplatin via JNK/mitochondrial pathway. Lung Cancer. 2008; 62:8–14.
- Zheng X, Carstens JL, Kim J, Scheible M, Kaye J, Sugimoto H, Wu C, LeBleu VS, Kalluri R. Epithelial-tomesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. Nature. 2015; 527:525–530.
- Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Brisken C, et al. The epithelialmesenchymal transition generates cells with properties of stem cells. Cell. 2008; 133:704–715.
- Liu S, Cong Y, Wang D, Sun Y, Deng L, Liu Y, Martin-Trevino R, Shang L, McDermott SP, Landis MD, Hong S, Adams A, D'Angelo R, et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. Stem Cell Reports. 2014; 2:78–91.
- 11. Christiansen JJ, Rajasekaran AK. Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. Cancer Res. 2006; 66:8319–8326.
- Garber K. Epithelial-to-mesenchymal transition is important to metastasis, but questions remain. J Natl Cancer Inst. 2008; 100:232–233, 239.
- 13. Chui MH. Insights into cancer metastasis from a clinicopathologic perspective: Epithelial-Mesenchymal

Transition is not a necessary step. Int J Cancer. 2013; 132:1487–1495.

- 14. Alix-Panabieres C, Pantel K. Circulating tumor cells: liquid biopsy of cancer. Clin Chem. 2013; 59:110–118.
- 15. Ferreira MM, Ramani VC, Jeffrey SS. Circulating tumor cell technologies. Mol Oncol. 2016; 10:374–394.
- 16. Cohen SJ, Punt CJ, Iannotti N, Saidman BH, Sabbath KD, Gabrail NY, Picus J, Morse M, Mitchell E, Miller MC, Doyle GV, Tissing H, Terstappen LW, et al. Relationship of circulating tumor cells to tumor response, progressionfree survival, and overall survival in patients with metastatic colorectal cancer. J Clin Oncol. 2008; 26:3213–3221.
- Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, Reuben JM, Doyle GV, Allard WJ, Terstappen LW, Hayes DF. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. N Engl J Med. 2004; 351:781–791.
- de Bono JS, Scher HI, Montgomery RB, Parker C, Miller MC, Tissing H, Doyle GV, Terstappen LW, Pienta KJ, Raghavan D. Circulating tumor cells predict survival benefit from treatment in metastatic castration-resistant prostate cancer. Clin Cancer Res. 2008; 14:6302–6309.
- Danila DC, Morris MJ, de Bono JS, Ryan CJ, Denmeade SR, Smith MR, Taplin ME, Bubley GJ, Kheoh T, Haqq C, Molina A, Anand A, Koscuiszka M, et al. Phase II multicenter study of abiraterone acetate plus prednisone therapy in patients with docetaxel-treated castrationresistant prostate cancer. J Clin Oncol. 2010; 28:1496–1501.
- Wang J, Wang K, Xu J, Huang J, Zhang T. Prognostic significance of circulating tumor cells in non-small-cell lung cancer patients: a meta-analysis. PLoS One. 2013; 8:e78070.
- 21. Christofori G. New signals from the invasive front. Nature. 2006; 441:444–450.
- 22. Peinado H, Olmeda D, Cano A. Snail, Zeb and bHLH factors in tumour progression: an alliance against the epithelial phenotype? Nat Rev Cancer. 2007; 7:415–428.
- 23. Yang J, Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. Dev Cell. 2008; 14:818–829.
- Katoh Y, Katoh M. Hedgehog signaling, epithelial-tomesenchymal transition and miRNA (review). Int J Mol Med. 2008; 22:271–275.
- De Craene B, Berx G. Regulatory networks defining EMT during cancer initiation and progression. Nat Rev Cancer. 2013; 13:97–110.
- Huber MA, Beug H, Wirth T. Epithelial-mesenchymal transition: NF-kappaB takes center stage. Cell Cycle. 2004; 3:1477–1480.
- Moustakas A, Heldin CH. Signaling networks guiding epithelial-mesenchymal transitions during embryogenesis and cancer progression. Cancer Sci. 2007; 98:1512–1520.
- 28. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. Direct regulation of TWIST

by HIF-1alpha promotes metastasis. Nat Cell Biol. 2008; 10:295–305.

- Imai T, Horiuchi A, Wang C, Oka K, Ohira S, Nikaido T, Konishi I. Hypoxia attenuates the expression of E-cadherin via up-regulation of SNAIL in ovarian carcinoma cells. Am J Pathol. 2003; 163:1437–1447.
- Bonnomet A, Brysse A, Tachsidis A, Waltham M, Thompson EW, Polette M, Gilles C. Epithelial-to-Mesenchymal Transitions and Circulating Tumor Cells. J Mammary Gland Biol Neoplasia. 2010; 15:261–273.
- Ikenouchi J, Matsuda M, Furuse M, Tsukita S. Regulation of tight junctions during the epithelium-mesenchyme transition: direct repression of the gene expression of claudins/occludin by Snail. J Cell Sci. 2003; 116:1959–1967.
- Vandewalle C, Comijn J, De Craene B, Vermassen P, Bruyneel E, Andersen H, Tulchinsky E, Van Roy F, Berx G. SIP1/ZEB2 induces EMT by repressing genes of different epithelial cell-cell junctions. Nucleic Acids Res. 2005; 33:6566–6578.
- Liberko M, Kolostova K, Bobek V. Essentials of circulating tumor cells for clinical research and practice. Crit Rev Oncol Hematol. 2013; 88:338–356.
- 34. Derksen PW, Liu X, Saridin F, van der Gulden H, Zevenhoven J, Evers B, van Beijnum JR, Griffioen AW, Vink J, Krimpenfort P, Peterse JL, Cardiff RD, Berns A, et al. Somatic inactivation of E-cadherin and p53 in mice leads to metastatic lobular mammary carcinoma through induction of anoikis resistance and angiogenesis. Cancer Cell. 2006; 10:437–449.
- 35. Hulit J, Suyama K, Chung S, Keren R, Agiostratidou G, Shan W, Dong X, Williams TM, Lisanti MP, Knudsen K, Hazan RB. N-cadherin signaling potentiates mammary tumor metastasis via enhanced extracellular signal-regulated kinase activation. Cancer Res. 2007; 67:3106–3116.
- 36. Yilmaz M, Christofori G. EMT, the cytoskeleton, and cancer cell invasion. Cancer Metastasis Rev. 2009; 28:15–33.
- Ota I, Li XY, Hu Y, Weiss SJ. Induction of a MT1-MMP and MT2-MMP-dependent basement membrane transmigration program in cancer cells by Snail1. Proc Natl Acad Sci USA. 2009; 106:20318–20323.
- 38. Huang CH, Yang WH, Chang SY, Tai SK, Tzeng CH, Kao JY, Wu KJ, Yang MH. Regulation of membrane-type 4 matrix metalloproteinase by SLUG contributes to hypoxiamediated metastasis. Neoplasia. 2009; 11:1371–1382.
- 39. Poincloux R, Lizarraga F, Chavrier P. Matrix invasion by tumour cells: a focus on MT1-MMP trafficking to invadopodia. J Cell Sci. 2009; 122:3015–3024.
- Labelle M, Begum S, Hynes RO. Direct signaling between platelets and cancer cells induces an epithelialmesenchymal-like transition and promotes metastasis. Cancer Cell. 2011; 20:576–590.
- Romero D, Al-Shareef Z, Gorrono-Etxebarria I, Atkins S, Turrell F, Chhetri J, Bengoa-Vergniory N, Zenzmaier C, Berger P, Waxman J, Kypta R. Dickkopf-3 regulates

prostate epithelial cell acinar morphogenesis and prostate cancer cell invasion by limiting TGF-beta-dependent activation of matrix metalloproteases. Carcinogenesis. 2016; 37:18–29.

- Wang L, Yu Y, Guan H, Liu T, Qiao C. 67-kDa Laminin receptor contributes to hypoxia-induced migration and invasion of trophoblast-like cells by mediating matrix metalloproteinase-9. Clin Exp Pharmacol Physiol. 2015; 42:549–558.
- Nakayama K. cAMP-response element-binding protein (CREB) and NF-kappaB transcription factors are activated during prolonged hypoxia and cooperatively regulate the induction of matrix metalloproteinase MMP1. J Biol Chem. 2013; 288:22584–22595.
- Peinado H, Marin F, Cubillo E, Stark HJ, Fusenig N, Nieto MA, Cano A. Snail and E47 repressors of E-cadherin induce distinct invasive and angiogenic properties *in vivo*. J Cell Sci. 2004; 117:2827–2839.
- 45. Fantozzi A, Gruber DC, Pisarsky L, Heck C, Kunita A, Yilmaz M, Meyer-Schaller N, Cornille K, Hopfer U, Bentires-Alj M, Christofori G. VEGF-mediated angiogenesis links EMT-induced cancer stemness to tumor initiation. Cancer Res. 2014; 74:1566–1575.
- 46. Niu RF, Zhang L, Xi GM, Wei XY, Yang Y, Shi YR, Hao XS. Up-regulation of Twist induces angiogenesis and correlates with metastasis in hepatocellular carcinoma. J Exp Clin Cancer Res. 2007; 26:385–394.
- 47. Jue C, Lin C, Zhisheng Z, Yayun Q, Feng J, Min Z, Haibo W, Youyang S, Hisamitsu T, Shintaro I, Shiyu G, Yanqing L. Notch1 promotes vasculogenic mimicry in hepatocellular carcinoma by inducing EMT signaling. Oncotarget. 2017; 8:2501–2513. https://doi.org/10.18632/ oncotarget.12388.
- 48. Chen QY, Jiao DM, Wu YQ, Chen J, Wang J, Tang XL, Mou H, Hu HZ, Song J, Yan J, Wu LJ, Chen J, Wang Z. MiR-206 inhibits HGF-induced epithelial-mesenchymal transition and angiogenesis in non-small cell lung cancer via c-Met /PI3k/Akt/mTOR pathway. Oncotarget. 2016; 7:18247–61. https://doi.org/10.18632/oncotarget.7570.
- 49. Wang Z, Banerjee S, Li Y, Rahman KM, Zhang Y, Sarkar FH. Down-regulation of notch-1 inhibits invasion by inactivation of nuclear factor-kappaB, vascular endothelial growth factor, and matrix metalloproteinase-9 in pancreatic cancer cells. Cancer Res. 2006; 66:2778–2784.
- 50. Kallergi G, Markomanolaki H, Giannoukaraki V, Papadaki MA, Strati A, Lianidou ES, Georgoulias V, Mavroudis D, Agelaki S. Hypoxia-inducible factorlalpha and vascular endothelial growth factor expression in circulating tumor cells of breast cancer patients. Breast Cancer Res. 2009; 11:R84.
- 51. Sounni NE, Roghi C, Chabottaux V, Janssen M, Munaut C, Maquoi E, Galvez BG, Gilles C, Frankenne F, Murphy G, Foidart JM, Noel A. Up-regulation of vascular endothelial growth factor-A by active membrane-type 1 matrix metalloproteinase through activation of Src-tyrosine kinases. J Biol Chem. 2004; 279:13564–13574.

- 52. van Hinsbergh VW, Engelse MA, Quax PH. Pericellular proteases in angiogenesis and vasculogenesis. Arterioscler Thromb Vasc Biol. 2006; 26:716–728.
- 53. Nagy JA, Chang SH, Shih SC, Dvorak AM, Dvorak HF. Heterogeneity of the tumor vasculature. Semin Thromb Hemost. 2010; 36:321–331.
- Qi J, Wang J, Romanyuk O, Siu CH. Involvement of Src family kinases in N-cadherin phosphorylation and betacatenin dissociation during transendothelial migration of melanoma cells. Mol Biol Cell. 2006; 17:1261–1272.
- 55. Drake JM, Strohbehn G, Bair TB, Moreland JG, Henry MD. ZEB1 enhances transendothelial migration and represses the epithelial phenotype of prostate cancer cells. Mol Biol Cell. 2009; 20:2207–2217.
- Singh A, Settleman J. EMT, cancer stem cells and drug resistance: an emerging axis of evil in the war on cancer. Oncogene. 2010; 29:4741–4751.
- 57. Tiwari N, Gheldof A, Tatari M, Christofori G. EMT as the ultimate survival mechanism of cancer cells. Semin Cancer Biol. 2012; 22:194–207.
- Cao Z, Livas T, Kyprianou N. Anoikis and EMT: Lethal "Liaisons" during Cancer Progression. Crit Rev Oncog. 2016; 21:155–168.
- 59. Frisch SM, Schaller M, Cieply B. Mechanisms that link the oncogenic epithelial-mesenchymal transition to suppression of anoikis. J Cell Sci. 2013; 126:21–29.
- 60. Smit MA, Peeper DS. Zeb1 is required for TrkB-induced epithelial-mesenchymal transition, anoikis resistance and metastasis. Oncogene. 2011; 30:3735–3744.
- Smit MA, Geiger TR, Song JY, Gitelman I, Peeper DS. A Twist-Snail axis critical for TrkB-induced epithelialmesenchymal transition-like transformation, anoikis resistance, and metastasis. Mol Cell Biol. 2009; 29:3722–3737.
- 62. Sayan AE, Griffiths TR, Pal R, Browne GJ, Ruddick A, Yagci T, Edwards R, Mayer NJ, Qazi H, Goyal S, Fernandez S, Straatman K, Jones GD, et al. SIP1 protein protects cells from DNA damage-induced apoptosis and has independent prognostic value in bladder cancer. Proc Natl Acad Sci USA. 2009; 106:14884–14889.
- 63. Cheng GZ, Chan J, Wang Q, Zhang W, Sun CD, Wang LH. Twist transcriptionally up-regulates AKT2 in breast cancer cells leading to increased migration, invasion, and resistance to paclitaxel. Cancer Res. 2007; 67:1979–1987.
- 64. Haslehurst AM, Koti M, Dharsee M, Nuin P, Evans K, Geraci J, Childs T, Chen J, Li J, Weberpals J, Davey S, Squire J, Park PC, et al. EMT transcription factors snail and slug directly contribute to cisplatin resistance in ovarian cancer. BMC Cancer. 2012; 12:91.
- 65. Kurrey NK, Jalgaonkar SP, Joglekar AV, Ghanate AD, Chaskar PD, Doiphode RY, Bapat SA. Snail and slug mediate radioresistance and chemoresistance by antagonizing p53-mediated apoptosis and acquiring a stemlike phenotype in ovarian cancer cells. Stem Cells. 2009; 27:2059–2068.

- Ren J, Chen Y, Song H, Chen L, Wang R. Inhibition of ZEB1 reverses EMT and chemoresistance in docetaxelresistant human lung adenocarcinoma cell line. J Cell Biochem. 2013; 114:1395–1403.
- 67. Feng B, Wang R, Song HZ, Chen LB. MicroRNA-200b reverses chemoresistance of docetaxel-resistant human lung adenocarcinoma cells by targeting E2F3. Cancer. 2012; 118:3365–76.
- Chen J, Yuan W, Wu L, Tang Q, Xia Q, Ji J, Liu Z, Ma Z, Zhou Z, Cheng Y, Shu X. PDGF-D promotes cell growth, aggressiveness, angiogenesis and EMT transformation of colorectal cancer by activation of Notch1/Twist1 pathway. Oncotarget. 2017; 8:9961–9973. https://doi.org/10.18632/ oncotarget.14283.
- Chen QK, Lee K, Radisky DC, Nelson CM. Extracellular matrix proteins regulate epithelial-mesenchymal transition in mammary epithelial cells. Differentiation. 2013; 86:126–132.
- 70. Lin CY, Tsai PH, Kandaswami CC, Lee PP, Huang CJ, Hwang JJ, Lee MT. Matrix metalloproteinase-9 cooperates with transcription factor Snail to induce epithelialmesenchymal transition. Cancer Sci. 2011; 102:815–827.
- 71. Gonzalez-Moreno O, Lecanda J, Green JE, Segura V, Catena R, Serrano D, Calvo A. VEGF elicits epithelialmesenchymal transition (EMT) in prostate intraepithelial neoplasia (PIN)-like cells via an autocrine loop. Exp Cell Res. 2010; 316:554–567.
- Chaffer CL, Thompson EW, Williams ED. Mesenchymal to epithelial transition in development and disease. Cells Tissues Organs. 2007; 185:7–19.
- 74. Chaffer CL, Weinberg RA. A perspective on cancer cell metastasis. Science. 2011; 331:1559–1564.
- Banyard J, Bielenberg DR. The role of EMT and MET in cancer dissemination. Connect Tissue Res. 2015; 56: 403–413.
- 76. Ksiazkiewicz M, Markiewicz A, Zaczek AJ. Epithelial-Mesenchymal Transition: A Hallmark in Metastasis Formation Linking Circulating Tumor Cells and Cancer Stem Cells. Pathobiology. 2012; 79:195–208.
- Brabletz T, Jung A, Reu S, Porzner M, Hlubek F, Kunz-Schughart LA, Knuechel R, Kirchner T. Variable betacatenin expression in colorectal cancers indicates tumor progression driven by the tumor environment. Proc Natl Acad Sci USA. 2001; 98:10356–10361.
- Hudson LG, Zeineldin R, Stack MS. Phenotypic plasticity of neoplastic ovarian epithelium: unique cadherin profiles in tumor progression. Clin Exp Metastasis. 2008; 25:643–655.
- 79. Chao YL, Shepard CR, Wells A. Breast carcinoma cells re-express E-cadherin during mesenchymal to epithelial reverting transition. Mol Cancer. 2010; 9:179.
- 80. Chao Y, Wu Q, Shepard C, Wells A. Hepatocyte induced re-expression of E-cadherin in breast and prostate cancer

cells increases chemoresistance. Clin Exp Metastasis. 2012; 29:39–50.

- Chaffer CL, Dopheide B, McCulloch DR, Lee AB, Moseley JM, Thompson EW, Williams ED. Upregulated MT1-MMP/TIMP-2 axis in the TSU-Pr1-B1/B2 model of metastatic progression in transitional cell carcinoma of the bladder. Clin Exp Metastasis. 2005; 22:115–125.
- Chaffer CL, Brennan JP, Slavin JL, Blick T, Thompson EW, Williams ED. Mesenchymal-to-epithelial transition facilitates bladder cancer metastasis: role of fibroblast growth factor receptor-2. Cancer Res. 2006; 66:11271–11278.
- Banyard J, Chung I, Wilson AM, Vetter G, Le Bechec A, Bielenberg DR, Zetter BR. Regulation of epithelial plasticity by miR-424 and miR-200 in a new prostate cancer metastasis model. Sci Rep. 2013; 3:3151.
- 84. Ocana OH, Corcoles R, Fabra A, Moreno-Bueno G, Acloque H, Vega S, Barrallo-Gimeno A, Cano A, Nieto MA. Metastatic colonization requires the repression of the epithelial-mesenchymal transition inducer Prrx1. Cancer Cell. 2012; 22:709–724.
- Tsai JH, Donaher JL, Murphy DA, Chau S, Yang J. Spatiotemporal regulation of epithelial-mesenchymal transition is essential for squamous cell carcinoma metastasis. Cancer Cell. 2012; 22:725–736.
- Friedl P, Wolf K. Tumour-cell invasion and migration: diversity and escape mechanisms. Nat Rev Cancer. 2003; 3:362–374.
- Friedl P, Gilmour D. Collective cell migration in morphogenesis, regeneration and cancer. Nat Rev Mol Cell Biol. 2009; 10:445–457.
- Klymkowsky MW, Savagner P. Epithelial-mesenchymal transition: a cancer researcher's conceptual friend and foe. Am J Pathol. 2009; 174:1588–1593.
- Wang C, Mu Z, Chervoneva I, Austin L, Ye Z, Rossi G, Palazzo JP, Sun C, Abu-Khalaf M, Myers RE, Zhu Z, Ba Y, Li B, et al. Longitudinally collected CTCs and CTC-clusters and clinical outcomes of metastatic breast cancer. Breast Cancer Res Treat. 2017; 161:83–94.
- Tsuji T, Ibaragi S, Hu GF. Epithelial-mesenchymal transition and cell cooperativity in metastasis. Cancer Res. 2009; 69:7135–7139.
- 91. Tsuji T, Ibaragi S, Shima K, Hu MG, Katsurano M, Sasaki A, Hu GF. Epithelial-mesenchymal transition induced by growth suppressor p12CDK2-AP1 promotes tumor cell local invasion but suppresses distant colony growth. Cancer Res. 2008; 68:10377–10386.
- 92. Banyard J, Chung I, Migliozzi M, Phan DT, Wilson AM, Zetter BR, Bielenberg DR. Identification of genes regulating migration and invasion using a new model of metastatic prostate cancer. BMC Cancer. 2014; 14:387.
- Giampieri S, Manning C, Hooper S, Jones L, Hill CS, Sahai E. Localized and reversible TGFbeta signalling switches breast cancer cells from cohesive to single cell motility. Nat Cell Biol. 2009; 11:1287–1296.

- 94. Massard C, Oulhen M, Le Moulec S, Auger N, Foulon S, Abou-Lovergne A, Billiot F, Valent A, Marty V, Loriot Y, Fizazi K, Vielh P, Farace F. Phenotypic and genetic heterogeneity of tumor tissue and circulating tumor cells in patients with metastatic castration-resistant prostate cancer: A report from the PETRUS prospective study. Oncotarget. 2016; 7:55069–55082. https://doi.org/10.18632/oncotarget.10396.
- 95. Powell AA, Talasaz AH, Zhang H, Coram MA, Reddy A, Deng G, Telli ML, Advani RH, Carlson RW, Mollick JA, Sheth S, Kurian AW, Ford JM, et al. Single cell profiling of circulating tumor cells: transcriptional heterogeneity and diversity from breast cancer cell lines. PLoS One. 2012; 7:e33788.
- Bednarz-Knoll N, Alix-Panabieres C, Pantel K. Plasticity of disseminating cancer cells in patients with epithelial malignancies. Cancer Metastasis Rev. 2012; 31:673–687.
- 97. Apostolaki S, Perraki M, Kallergi G, Kafousi M, Papadopoulos S, Kotsakis A, Pallis A, Xenidis N, Kalmanti L, Kalbakis K, Agelaki S, Kalykaki A, Stournaras C, et al. Detection of occult HER2 mRNApositive tumor cells in the peripheral blood of patients with operable breast cancer: evaluation of their prognostic relevance. Breast Cancer Res Treat. 2009; 117:525–534.
- 98. Ligthart ST, Bidard FC, Decraene C, Bachelot T, Delaloge S, Brain E, Campone M, Viens P, Pierga JY, Terstappen LW. Unbiased quantitative assessment of Her-2 expression of circulating tumor cells in patients with metastatic and non-metastatic breast cancer. Ann Oncol. 2013; 24:1231–1238.
- 99. Attard G, Swennenhuis JF, Olmos D, Reid AH, Vickers E, A'Hern R, Levink R, Coumans F, Moreira J, Riisnaes R, Oommen NB, Hawche G, Jameson C, et al. Characterization of ERG, AR and PTEN gene status in circulating tumor cells from patients with castration-resistant prostate cancer. Cancer Res. 2009; 69:2912–2918.
- 100. Maheswaran S, Sequist LV, Nagrath S, Ulkus L, Brannigan B, Collura CV, Inserra E, Diederichs S, Iafrate AJ, Bell DW, Digumarthy S, Muzikansky A, Irimia D, et al. Detection of mutations in EGFR in circulating lung-cancer cells. N Engl J Med. 2008; 359:366–377.
- 101. Yang MJ, Chiu HH, Wang HM, Yen LC, Tsao DA, Hsiao CP, Chen YF, Wang JY, Lin SR. Enhancing detection of circulating tumor cells with activating KRAS oncogene in patients with colorectal cancer by weighted chemiluminescent membrane array method. Ann Surg Oncol. 2010; 17:624–633.
- 102. Kasimir-Bauer S, Hoffmann O, Wallwiener D, Kimmig R, Fehm T. Expression of stem cell and epithelial-mesenchymal transition markers in primary breast cancer patients with circulating tumor cells. Breast Cancer Res. 2012; 14:R15.
- 103. Aktas B, Tewes M, Fehm T, Hauch S, Kimmig R, Kasimir-Bauer S. Stem cell and epithelial-mesenchymal transition markers are frequently overexpressed in circulating tumor cells of metastatic breast cancer patients. Breast Cancer Res. 2009; 11:R46.

- 104. Armstrong AJ, Marengo MS, Oltean S, Kemeny G, Bitting RL, Turnbull JD, Herold CI, Marcom PK, George DJ, Garcia-Blanco MA. Circulating tumor cells from patients with advanced prostate and breast cancer display both epithelial and mesenchymal markers. Mol Cancer Res. 2011; 9:997–1007.
- 105. Patriarca C, Macchi RM, Marschner AK, Mellstedt H. Epithelial cell adhesion molecule expression (CD326) in cancer: a short review. Cancer Treat Rev. 2012; 38:68–75.
- 106. Sieuwerts AM, Kraan J, Bolt J, van der Spoel P, Elstrodt F, Schutte M, Martens JW, Gratama JW, Sleijfer S, Foekens JA. Anti-epithelial cell adhesion molecule antibodies and the detection of circulating normal-like breast tumor cells. J Natl Cancer Inst. 2009; 101:61–66.
- 107. Allard WJ, Matera J, Miller MC, Repollet M, Connelly MC, Rao C, Tibbe AG, Uhr JW, Terstappen LW. Tumor cells circulate in the peripheral blood of all major carcinomas but not in healthy subjects or patients with nonmalignant diseases. Clin Cancer Res. 2004; 10:6897–6904.
- 108. Mikolajczyk SD, Millar LS, Tsinberg P, Coutts SM, Zomorrodi M, Pham T, Bischoff FZ, Pircher TJ. Detection of EpCAM-Negative and Cytokeratin-Negative Circulating Tumor Cells in Peripheral Blood. J Oncol. 2011; 2011:252361.
- 109. Runkle EA, Mu D. Tight junction proteins: from barrier to tumorigenesis. Cancer Lett. 2013; 337:41–48.
- 110. Gall TM, Frampton AE. Gene of the month: E-cadherin (CDH1). J Clin Pathol. 2013; 66:928–932.
- 111. Hazan RB, Qiao R, Keren R, Badano I, Suyama K. Cadherin switch in tumor progression. Ann N Y Acad Sci. 2004; 1014:155–163.
- 112. Satelli A, Li S. Vimentin in cancer and its potential as a molecular target for cancer therapy. Cell Mol Life Sci. 2011; 68:3033–3046.
- 113. Barriere G, Fici P, Gallerani G, Fabbri F, Zoli W, Rigaud M. Circulating tumor cells and epithelial, mesenchymal and stemness markers: characterization of cell subpopulations. Ann Transl Med. 2014; 2:109.
- 114. Wushou A, Hou J, Zhao YJ, Shao ZM. Twist-1 upregulation in carcinoma correlates to poor survival. Int J Mol Sci. 2014; 15:21621–21630.
- 115. Porta C, Paglino C, Mosca A. Targeting PI3K/Akt/mTOR Signaling in Cancer. Front Oncol. 2014; 4:64.
- 116. Barriere G, Riouallon A, Renaudie J, Tartary M, Rigaud M. Mesenchymal and stemness circulating tumor cells in early breast cancer diagnosis. BMC Cancer. 2012; 12:114.
- 117. Kallergi G, Agelaki S, Kalykaki A, Stournaras C, Mavroudis D, Georgoulias V. Phosphorylated EGFR and PI3K/Akt signaling kinases are expressed in circulating tumor cells of breast cancer patients. Breast Cancer Res. 2008; 10:R80.
- 118. Li YM, Xu SC, Li J, Han KQ, Pi HF, Zheng L, Zuo GH, Huang XB, Li HY, Zhao HZ, Yu ZP, Zhou Z, Liang P. Epithelial-mesenchymal transition markers expressed in

circulating tumor cells in hepatocellular carcinoma patients with different stages of disease. Cell Death Dis. 2013; 4:e831.

- 119. Ning Y, Zhang W, Hanna DL, Yang D, Okazaki S, Berger MD, Miyamoto Y, Suenaga M, Schirripa M, El-Khoueiry A, Lenz HJ. Clinical relevance of EMT and stemlike gene expression in circulating tumor cells of metastatic colorectal cancer patients. Pharmacogenomics J. 2016.
- 120. Kulemann B, Liss AS, Warshaw AL, Seifert S, Bronsert P, Glatz T, Pitman MB, Hoeppner J. KRAS mutations in pancreatic circulating tumor cells: a pilot study. Tumour Biol. 2016; 37:7547–7554.
- 121. Lindsay CR, Le Moulec S, Billiot F, Loriot Y, Ngo-Camus M, Vielh P, Fizazi K, Massard C, Farace F. Vimentin and Ki67 expression in circulating tumour cells derived from castrate-resistant prostate cancer. BMC Cancer. 2016; 16:168.
- 122. Pal SK, He M, Wilson T, Liu X, Zhang K, Carmichael C, Torres A, Hernandez S, Lau C, Agarwal N, Kawachi M, Yen Y, Jones JO. Detection and phenotyping of circulating tumor cells in high-risk localized prostate cancer. Clin Genitourin Cancer. 2015; 13:130–136.
- 123. Gradilone A, Raimondi C, Nicolazzo C, Petracca A, Gandini O, Vincenzi B, Naso G, Agliano AM, Cortesi E, Gazzaniga P. Circulating tumour cells lacking cytokeratin in breast cancer: the importance of being mesenchymal. J Cell Mol Med. 2011; 15:1066–1070.
- 124. Fina E, Reduzzi C, Motta R, Di Cosimo S, Bianchi G, Martinetti A, Wechsler J, Cappelletti V, Daidone MG. Did circulating tumor cells tell us all they could? The missed circulating tumor cell message in breast cancer. Int J Biol Markers. 2015; 30:e429-e433.
- 125. Kallergi G, Papadaki MA, Politaki E, Mavroudis D, Georgoulias V, Agelaki S. Epithelial to mesenchymal transition markers expressed in circulating tumour cells of early and metastatic breast cancer patients. Breast Cancer Res. 2011; 13:R59.
- 126. Papadaki MA, Kallergi G, Zafeiriou Z, Manouras L, Theodoropoulos PA, Mavroudis D, Georgoulias V, Agelaki S. Co-expression of putative stemness and epithelial-to-mesenchymal transition markers on single circulating tumour cells from patients with early and metastatic breast cancer. BMC Cancer. 2014; 14:651.
- 127. Alonso-Alconada L, Muinelo-Romay L, Madissoo K, Diaz-Lopez A, Krakstad C, Trovik J, Wik E, Hapangama D, Coenegrachts L, Cano A, Gil-Moreno A, Chiva L, Cueva J, et al. Molecular profiling of circulating tumor cells links plasticity to the metastatic process in endometrial cancer. Mol Cancer. 2014; 13:223.
- 128. Chang K, Kong YY, Dai B, Ye DW, Qu YY, Wang Y, Jia ZW, Li GX. Combination of circulating tumor cell

enumeration and tumor marker detection in predicting prognosis and treatment effect in metastatic castration-resistant prostate cancer. Oncotarget. 2015; 6:41825–41836. https://doi.org/10.18632/oncotarget.6167.

- 129. Mego M, Gao H, Lee BN, Cohen EN, Tin S, Giordano A, Wu Q, Liu P, Nieto Y, Champlin RE, Hortobagyi GN, Cristofanilli M, Ueno NT, et al. Prognostic Value of EMT-Circulating Tumor Cells in Metastatic Breast Cancer Patients Undergoing High-Dose Chemotherapy with Autologous Hematopoietic Stem Cell Transplantation. J Cancer. 2012; 3:369–380.
- 130. Polioudaki H, Agelaki S, Chiotaki R, Politaki E, Mavroudis D, Matikas A, Georgoulias V, Theodoropoulos PA. Variable expression levels of keratin and vimentin reveal differential EMT status of circulating tumor cells and correlation with clinical characteristics and outcome of patients with metastatic breast cancer. BMC Cancer. 2015; 15:399.
- 131. Liu YK, Hu BS, Li ZL, He X, Li Y, Lu LG. An improved strategy to detect the epithelial-mesenchymal transition process in circulating tumor cells in hepatocellular carcinoma patients. Hepatol Int. 2016; 10:640–646.
- 132. Zhao R, Cai Z, Li S, Cheng Y, Gao H, Liu F, Wu S, Liu S, Dong Y, Zheng L, Zhang W, Wu X, Yao X. Expression and clinical relevance of epithelial and mesenchymal markers in circulating tumor cells from colorectal cancer. Oncotarget. 2017; 8:9293–9302. https://doi.org/10.18632/ oncotarget.14065.
- 133. Li TT, Liu H, Li FP, Hu YF, Mou TY, Lin T, Yu J, Zheng L, Li GX. Evaluation of epithelial-mesenchymal transitioned circulating tumor cells in patients with resectable gastric cancer: Relevance to therapy response. World J Gastroenterol. 2015; 21:13259–13267.
- 134. Satelli A, Mitra A, Brownlee Z, Xia X, Bellister S, Overman MJ, Kopetz S, Ellis LM, Meng QH, Li S. Epithelial-mesenchymal transitioned circulating tumor cells capture for detecting tumor progression. Clin Cancer Res. 2015; 21:899–906.
- 135. Nel I, Baba HA, Ertle J, Weber F, Sitek B, Eisenacher M, Meyer HE, Schlaak JF, Hoffmann AC. Individual profiling of circulating tumor cell composition and therapeutic outcome in patients with hepatocellular carcinoma. Transl Oncol. 2013; 6:420–428.
- 136. Yu M, Bardia A, Wittner BS, Stott SL, Smas ME, Ting DT, Isakoff SJ, Ciciliano JC, Wells MN, Shah AM, Concannon KF, Donaldson MC, Sequist LV, et al. Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition. Science. 2013; 339:580–584.