Editorial

The APC/C and CK1 in the developing brain

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Casein Kinase 1δ (CK1 δ) is a serione/threonine kinase required for cell cycle progression, circadian rhythm, vesicle trafficking, and neurite outgrowth [1]. $CK1\delta$ is also a therapeutic target in various cancers, Alzheimer's disease, alcoholism, and sleep disorders [1]. To examine the role of CK1 δ in brain development, we used cerebellar granule cell progenitors (GCPs) as a model system. GCPs are the most abundant neurons in the mammalian brain and are one of two principal neurons in the cerebellar circuitry [2]. CK1 δ is expressed in GCPs during peak times of proliferation (postnatal day 6-postnatal day 8). To probe a role for CK1 δ in proliferation of GCPs during this time, we assayed proliferation in GCPs lacking CK18, after knockdown of CK18 by RNAi methodology or in purified GCPs treated with highly specific CK1 δ inhibitor [3]. In all three cases, ³H-thymidine incorporation assays showed reduced levels of proliferation. Given CK18's role in GCP neurogenesis, we anticipated that CK1 δ levels would decrease as GCPs exit the cell cycle. Indeed, we found that CK1 δ protein but not mRNA levels dropped during cell cycle exit, which suggested that CK1 δ is targeted for degradation during this time.

Importantly, biochemical assays demonstrate that CK1 δ is targeted for degradation via the Anaphase Promoting Complex/cyclosome (APC/C), a multisubunit E3 ubiquitin ligase, which has well-established roles in mitotic exit and G1 progression [2]. APC/C is also active in differentiating and differentiated cells [4]. APC/C associates with one of two activators termed Cdc20 or Cdh1, which recruit substrates to bring them into close proximity of the E2 enzyme bound to APC/C [4]. We report that Cdh1 binds to CK1 δ to initiate APC/C dependent ubiquination. In vitro ubiquitination assays containing purified APC/C and CK18 demonstrate that APC/C mediates CK1δ polyubiquitination in vitro. APC/C mediate ubiquitination of CK18 was dependent on two N-terminal destruction boxes in CK18 as mutation of these sites abrogated ubiquitination in vitro [2]. To demonstrate a requirement for CK1 δ in vivo we deleted CK1 δ in GCPs in the cerebellum [2]. Deletion of the APC/C activator Cdh1 in GCPs increased CK18 levels, suggesting that $CK1\delta$ is turned over in GCPs [2]. Collectively, these studies suggest that APC/C targets CK18 for destruction in vitro and in vivo and that APC/C^{Cdh1} is an important regulator of GCP proliferation by controlling CK18.

Our studies therefore suggest that APC/C-mediated degradation of CK1 δ functions in multiple steps in CNS

neuronal differentiation. CK18 has been linked to neurite outgrowth [5] and thus it will be important to determine whether APC/C mediated degradation of CK18 occurs in axons or dendrites. Prior studies demonstrated that APC/C inhibition in postmitotic neurons [4] increases neurite outgrowth while CK1 δ inhibition reduces neurite outgrowth in cell lines [5]. Thus, CK18 could be one of the substrates, which APC/C targets during neurite outgrowth, and whose levels rise during APC/C inhibition or depletion. It will be important to determine whether CK1 δ protein levels are modulated by APC/C active in postmitotic neurons. Interestingly, there are two forms of the APC/C that are active in postmitotic neurons, APC/ C^{Cdh1} and APC/C^{Cdc20} [4]. APC/C^{Cdh1} represses axonal growth [4] while APC/C^{Cdc20} activity controls dendritic morphogenesis [4]. APC/C^{Cdc20} is localized to centrosomes in postmitotic neurons. Given the finding that $CK1\delta$ is localized to centrosomes [5] it will be interesting to determine whether APC/C^{Cdc20} is able to induce CK1 δ destruction at centrosomes. An alternative could be that centrosome bound CK1 δ is protected from APC/C mediated degradation as other APC/C substrates cannot be ubiquitinated and degraded when bound to microtubules [6].

As centrosomal proteins often have roles in migration it will be important to determine whether

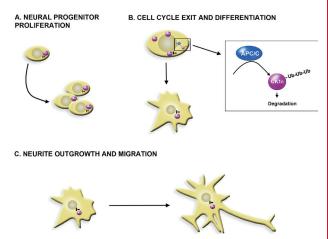


Figure 1: Model of CK1δ and APC/C during proliferation and differentiation of GCPs. A. CK1δ activity is required for GCP proliferation. **B.** CK1δ is degraded via APC/C mediated ubiquitination during GCP cell cycle exit and differentiation. Some centrosome bound CK1δ may be protected from APC/C mediated degradation. **C.** Centrosomal CK1δ is required for neurite outgrowth and migration.

APC/C mediated control of CK1 δ is linked to migration of neuronal precursors. Consistent with a role for CK1 δ in neuronal migration we found that CK1 δ inhibition reduced GCP migration *ex vivo* (unpublished observations).

In addition, since CK1 δ has a role in ciliogenesis¹ and APC/C^{Cdc20} has been reported to be required for primary cilia formation [7], the APC/C may interact with $CK1\delta$ in primary cilia. It will be interesting to determine whether APC/C^{Cdc20} induces CK1 δ degradation within cilia. Interestingly, since the primary cilium is required for Hedgehog (Hh) pathway signaling as we found that CK18 inhibition or disruption reduced Hh signaling in GCPs [2], it will be essential to determine whether the APC/C-CK18 interaction is important for Hh signaling in GCPs. Future studies will determine the importance of the APC/C-CK18 interaction in various signaling pathways including Hh and WNT, where CK18 has been implicated [1]. Furthermore, it will be critical to determine whether the APC/C-CK18 interaction is dysregulated in various neurological diseases.

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REFERENCES

- 1. Knippschild U, et al. Frontiers in oncology. 2014; 4: 96.
- 2. Penas C, et al. Cell reports. 2015; 11: 249-60.
- Penas C, et al. The Journal of biological chemistry. 2014; 289: 18893-903.
- 4. Penas C, et al. Frontiers in oncology. 2011; 1: 60.
- 5. Greer YE, et al. The Journal of cell biology. 2011; 192: 993-1004.
- 6. Song L, et al. Molecular cell. 2014; 53: 179-92.
- 7. Wang W, et al. eLife 3. 2014; e0308.