**Research Perspective** 

# Immunomodulatory and immunotherapeutic implications of tobacco smoking in squamous cell carcinomas and normal airway epithelium

# Jingming Wang<sup>1,2</sup>, Maximilian Linxweiler<sup>2,4</sup>, Wei Yang<sup>1,2</sup>, Timothy A. Chan<sup>1,2,3</sup> and Luc G.T. Morris<sup>1,4</sup>

<sup>1</sup>Immunogenomics Precision Oncology Platform, Memorial Sloan Kettering Cancer Center, New York, NY, USA

<sup>2</sup>Human Oncology and Pathogenesis Program, Memorial Sloan Kettering Cancer Center, New York, NY, USA

<sup>3</sup>Department of Radiation Oncology, Memorial Sloan Kettering Cancer Center, New York, NY, USA

<sup>4</sup>Department of Surgery, Memorial Sloan Kettering Cancer Center, New York, NY, USA

Correspondence to: Timothy A. Chan, email: chant@mskcc.org Luc G.T. Morris, email: morrisl@mskcc.org

Keywords: immune; microenvironment

Received: April 22, 2019 Accepted: May 13, 2019 Published: June 11, 2019

**Copyright:** Wang et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## ABSTRACT

The mutagenic effects of tobacco smoking increase the risk of the development of cancers of the lung, head and neck, and other anatomic sites. In a comparison of squamous cell carcinomas of the lung and the head and neck, we find that the immunomodulatory effects of smoking differ based on anatomic site. In both sites, the mutational signature of smoking is strongly associated with somatic mutational load. In head and neck squamous cell carcinoma, the mutational signature of tobacco exposure is associated with a strongly immunosuppressive tumor microenvironment. In contrast, in lung squamous cell carcinoma, the opposite effect is seen, with the tumor immune microenvironment significantly more inflamed. These effects are mirrored in rates of response to immune checkpoint inhibitor immunotherapy, which tend to be higher in smokers with lung cancer, but lower in smokers with head and neck cancer. We find a similarly strong immunosuppressive effect of smoking in non-cancerous lung epithelium. Taken together, our findings show that the effects of mutational signatures on the immune microenvironment and response to immunotherapy can be affected by context such as cancer type, anatomic site, and histology.

#### **INTRODUCTION**

Traditionally, cancers have been categorized by features such as anatomic site and tissue histology. Our understanding of cancer biology has subsequently evolved to recognize that tumors arising from diverse anatomic sites may share oncogenic molecular drivers or biomarkers. In recent years, targeted and immunotherapeutic drugs have demonstrated activity across multiple cancer types that are defined by a shared oncogenic alteration, such as a driver gene or fusion, or a shared biological process, such as DNA mismatch repair deficiency. This has led some to speculate that oncology drug treatments may be effective, and can receive regulatory approval, based only on molecular markers, in a "tissue-agnostic" fashion. It remains unknown how broadly tissue-agnostic approaches are applicable, and whether context such as anatomic site or tissue histology may alter the implications of these molecular markers [1–3].

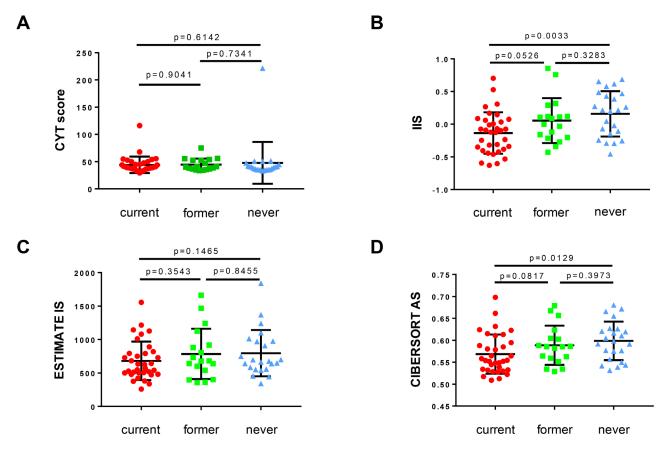
Tobacco smoking contributes to the development of multiple cancers, including head and neck and lung cancer [4, 5]. Tobacco smoke contains many carcinogenic chemicals that disrupt DNA such as polycyclic aromatic hydrocarbons and nitrosamines, which can cause  $G \rightarrow T$ transversions and distinct mutational signatures [6, 7]. Apart from increasing the somatic mutational burden associated with unrepaired DNA damage, tobacco carcinogens also appear to alter both the innate and the adaptive immune system, which may contribute to tumorigenesis [8–10]. The likelihood of a tumor responding to immunotherapy treatments such as immune checkpoint blockade (ICB) is known to be affected by these same factors, namely tumor mutational burden (TMB) and the degree of immune infiltration in the tumor microenvironment [11–15].

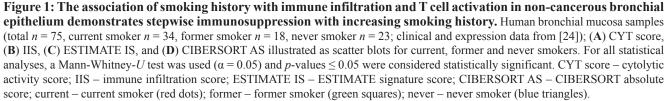
Immune checkpoint inhibitors are a class of cancer immunotherapy drugs that seek to inhibit T-cell checkpoints [16]. Pivotal trials have shown that a subset of patients with advanced melanoma, non-small cell lung cancer (NSCLC), renal cell carcinoma (RCC), bladder cancer, head and neck squamous cell carcinoma (HNSC), and many other tumor types, experience clinical responses following ICB therapy [17–23]. However, there are still a large majority of patients who do not experience tumor response to ICB, with response rates of 10–20% in HNSC and NSCLC. Interestingly, early clinical observations demonstrated that lung cancer patients with a smoking

history tended to have a higher likelihood of ICB response; however, for HNSC patients, the likelihood of response appeared to be lower in smokers [4, 5, 11, 18, 19].

In order to understand how tobacco smoking affects the tumor immune microenvironment and potentially identify the biomarkers to guide treatment options, we analyzed RNA and DNA sequencing data from cases studied as part of The Cancer Genome Atlas (TCGA), as well as two independent gene expression datasets of lung squamous cell carcinoma (LUSC) and HNSC tumors [4]. We found that the mutational signature of tobacco smoking was evident in both lung and head and neck tumors. In both LUSC and HNSC, we found that a higher mutational smoking signature was associated with a higher TMB, as would be expected.

We then examined the association between the mutational signature of smoking and RNA sequencingderived measures of tumor immune infiltration and T cell activation. In LUSC, a higher mutational smoking signature was positively associated with levels of immune infiltration, cytolytic activity and interferon- $\gamma$  pathway





signaling, indicating that smoking was associated with a more inflamed tumor immune microenvironment. In HNSC, these associations were all strongly in the opposite direction.

These findings have been apparent from clinical data. It was first observed by the Chan group that the genetic smoking signature was associated with a higher TMB and a higher response to anti-PD-1 immunotherapy [11]. It is possible that this higher response rate was also, in part, driven by a more T cell inflamed microenvironment in patients with a heavy smoking history. In HNSC, the reverse trend appears to emerge – we found that HNSC patients with a clinical smoking history tended to have a lower likelihood benefit from anti-PD-1 immunotherapy. These results underscore that contextual factors such as cancer type and anatomic site may play a role in modulating the interaction between the tumor genome and anti-tumor immunity.

Tobacco smoking has both immunosuppressive effects, such as a pro-apoptotic effect on T cells, as well as inflammatory effects. The balance of these sequelae may differ in the epithelium of the lung and bronchus versus upper aerodigestive tract mucosa. Furthermore, they may differ in time - one effect may predominate in normal epithelium, prior to cancer initiation, and the other effect may be more relevant during the development of an earlystage cancer. To better understand the effects of tobacco smoke in sculpting the immune microenvironment of noncancerous normal cells, we analyzed gene expression data from normal human airway epithelial cells in volunteers undergoing bronchoscopy [24]. In this study, Spira and colleagues obtained bronchoscopic biopsies of normal lower airway epithelium from a group of 34 current smokers, 18 former smokers and 23 never smokers. We analyzed these data and deconvolved gene expression data to measure immune infiltration and T cell activation [25, 26]. We determined levels of immune activation and infiltration by calculating the cytolytic (CYT) score (incorporating cytolytic effectors of CD8+ T cells: GZMA, and PRF1) [27], immune infiltration score (IIS) [28], ESTIMATE signature score [29] and CIBERSORT absolute score [30] for each sample (Figure 1). In these samples, we observed that smoking was associated with a significantly immunosuppressive microenvironment in normal human airway epithelium, most notably in current smokers, and less so in former smokers.

These findings will require further mechanistic investigation, perhaps with *in vivo* immunocompetent models of smoking carcinogenesis [31, 32]. We believe that these findings are consistent with profoundly immunosuppressive effects of tobacco smoke on the local immune microenvironment, which together with the mutagenic effects of tobacco, can lead to the initiation of cancer. In certain sites or anatomic locations such as the lung, the higher tumor mutational load associated with smoking may lead to an enhancement of T cell infiltration that becomes evident after tumor initiation. This effect is not seen in the head and neck mucosa, possibly because the immunosuppressive effects of smoking are more profound, or the degree of T cell infiltration responding to elevated mutational load is less marked. These effects would then potentially affect the probability of response to immunotherapies [33]. It is important to note that this latter point at present speculative and that the mechanistic aspects of carcinogenesis are likely to be far more complex. Indeed, there are several different carcinogenic compounds in cigarette smoke which are likely to exert differing immunosuppressive and inflammatory effects in different parts of the upper (HNSC) or lower (LUSC) airways.

The effect of smoking on cancer immunity and immunotherapy response needs to be explored across cancer types more broadly. A recent study of melanoma patients found that smoking had a strong negative prognostic effect in highly immune infiltrated tumors [34]. The systemic effects of tobacco smoke on smoking-associated cancers that are not directly exposed to smoke, such as bladder cancer, will also be important to dissect further. Future mechanistic and clinical studies will be needed to elucidate the importance of this risk factor, and its associated mutational signature, to the shaping of the tumor immune microenvironment, and the development of response and resistance to immunotherapies. These data indicate that it is very likely that cancer-causing processes, and mutational signatures, exert different effects on anti-tumor immunity in different contexts, and that further research will need to consider the interaction between molecular carcinogenesis and cancer type/location. Together with TMB, PD-L1 staining, measures of immune infiltration, HLA status, and other factors, it is likely that smoking history and/or the smoking mutational signature will add predictive value to our efforts to define biomarkers of response to ICB.

# **ACKNOWLEDGMENTS AND FUNDING**

The authors wish to acknowledge our patients and their families who selflessly contributed time and samples to support this research, the patients and investigators who contributed to the TCGA studies analyzed here, members of the Timothy Chan Lab for insightful discussions, and members of the MSKCC Immunogenomics and Precision Oncology Platform. This work was supported by the NIH/ NCI Cancer Center Support Grant P30 CA008748 (to MSKCC), Cycle for Survival (R.J.W., L.G.T.M., T.A.C.), The Frederick Adler Chair at MSKCC, The Jayme Flowers Fund, The Sebastian Nativo Fund, The Damon Runyon Cancer Research Foundation, NIH K08 DE024774, NIH R01 DE027738 (L.G.T.M.), The Adenoid Cystic Carcinoma Cancer Research Foundation, (T.A.C., A.L.H., L.G.T.M.), NIH R01 CA205426, The Pershing Square Sohn Cancer Research Alliance, the STARR Cancer Consortium, and The PaineWebber Chair at MSKCC (T.A.C.).

## **CONFLICTS OF INTEREST**

TAC is a co-founder of Gritstone Oncology and holds equity. TAC acknowledges grant funding from Bristol-Myers Squibb, AstraZeneca, Illumina, Pfizer, An2H, and Eisai. TAC has served as an advisor for Bristol-Myers Squibb, Illumina, Eisai, and An2H. L.G.T.M. received consulting fees from Rakuten Aspyrian and speaker fees from Physician Educational Resources.

# REFERENCES

- U.S. Food and Drug Administration. FDA approves first cancer treatment for any solid tumor with a specific genetic feature. 2017. <u>https://www.fda.gov/newsevents/newsroom/</u> pressannouncements/ucm560167.htm.
- Bhangoo MS, Sigal D. TRK Inhibitors: Clinical Development of Larotrectinib. Curr Oncol Rep. 2019; 21:14. <u>https://doi.org/10.1007/s11912-019-0761-y</u>. [PubMed]
- Marcus L, Lemery SJ, Keegan P, Pazdur R. FDA Approval Summary: Pembrolizumab for the treatment of microsatellite instability-high solid tumors. Clin Cancer Res. 2019 Feb 20. <u>https://doi.org/10.1158/1078-0432.CCR-18-4070</u>. [Epub ahead of print]. [PubMed]
- Desrichard A, Kuo F, Chowell D, Lee KW, Riaz N, Wong RJ, Chan TA, Morris LGT. Tobacco Smoking-Associated Alterations in the Immune Microenvironment of Squamous Cell Carcinomas. J Natl Cancer Inst. 2018; 110:1386–92. https://doi.org/10.1093/jnci/djy060. [PubMed]
- Cancer Genome Atlas Network. Comprehensive genomic characterization of head and neck squamous cell carcinomas. Nature. 2015; 517:576–82. <u>https://doi.org/10.1038/nature14129</u>. [PubMed]
- Hecht SS. Tobacco carcinogens, their biomarkers and tobacco-induced cancer. Nat Rev Cancer. 2003; 3:733–44. <u>https://doi.org/10.1038/nrc1190. [PubMed]</u>
- Alexandrov LB, Ju YS, Haase K, Van Loo P, Martincorena I, Nik-Zainal S, Totoki Y, Fujimoto A, Nakagawa H, Shibata T, Campbell PJ, Vineis P, Phillips DH, Stratton MR. Mutational signatures associated with tobacco smoking in human cancer. Science. 2016; 354:618–22. <u>https://doi. org/10.1126/science.aag0299</u>. [PubMed]
- Qiu F, Liang CL, Liu H, Zeng YQ, Hou S, Huang S, Lai X, Dai Z. Impacts of cigarette smoking on immune responsiveness: up and down or upside down? Oncotarget. 2017; 8:268–84. <u>https://doi.org/10.18632/oncotarget.13613</u>. [PubMed]
- Jaspers I. Cigarette smoke effects on innate immune mechanisms in the nasal mucosa. Potential effects on the microbiome. Ann Am Thorac Soc. 2014; 11:S38–42. <u>https:// doi.org/10.1513/AnnalsATS.201306-154MG</u>. [PubMed]
- Sopori M. Effects of cigarette smoke on the immune system. Nat Rev Immunol. 2002; 2:372–77. <u>https://doi.org/10.1038/nri803</u>. [PubMed]

- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS, Miller ML, Rekhtman N, Moreira AL, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science. 2015; 348:124–28. <u>https://doi.org/10.1126/science.aaa1348</u>. [PubMed]
- Tumeh PC, Harview CL, Yearley JH, Shintaku IP, Taylor EJ, Robert L, Chmielowski B, Spasic M, Henry G, Ciobanu V, West AN, Carmona M, Kivork C, et al. PD-1 blockade induces responses by inhibiting adaptive immune resistance. Nature. 2014; 515:568–71. <u>https://doi.org/10.1038/</u> <u>nature13954</u>. [PubMed]
- Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS, Hollmann TJ, Bruggeman C, Kannan K, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med. 2014; 371:2189–99. <u>https://doi.org/10.1056/ NEJMoa1406498</u>. [PubMed]
- Samstein RM, Lee CH, Shoushtari AN, Hellmann MD, Shen R, Janjigian YY, Barron DA, Zehir A, Jordan EJ, Omuro A, Kaley TJ, Kendall SM, Motzer RJ, et al. Tumor mutational load predicts survival after immunotherapy across multiple cancer types. Nat Genet. 2019; 51:202–06. https://doi.org/10.1038/s41588-018-0312-8. [PubMed]
- Takada K, Kashiwagi S, Asano Y, Goto W, Takahashi K, Fujita H, Takashima T, Tomita S, Hirakawa K, Ohira M. Clinical verification of the relationship between smoking and the immune microenvironment of breast cancer. J Transl Med. 2019; 17:13. <u>https://doi.org/10.1186/s12967-019-1773-y</u>. [PubMed]
- Wei SC, Duffy CR, Allison JP. Fundamental Mechanisms of Immune Checkpoint Blockade Therapy. Cancer Discov. 2018; 8:1069–86. <u>https://doi.org/10.1158/2159-8290.</u> <u>CD-18-0367. [PubMed]</u>
- Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, Patnaik A, Aggarwal C, Gubens M, Horn L, Carcereny E, Ahn MJ, Felip E, et al, and KEYNOTE-001 Investigators. Pembrolizumab for the treatment of nonsmall-cell lung cancer. N Engl J Med. 2015; 372:2018–28. https://doi.org/10.1056/NEJMoa1501824. [PubMed]
- Carbone DP, Reck M, Paz-Ares L, Creelan B, Horn L, Steins M, Felip E, van den Heuvel MM, Ciuleanu TE, Badin F, Ready N, Hiltermann TJ, Nair S, et al, and CheckMate 026 Investigators. First-Line Nivolumab in Stage IV or Recurrent Non-Small-Cell Lung Cancer. N Engl J Med. 2017; 376:2415–26. <u>https://doi.org/10.1056/ NEJMoa1613493</u>. [PubMed]
- Ferris RL, Blumenschein G Jr, Fayette J, Guigay J, Colevas AD, Licitra L, Harrington K, Kasper S, Vokes EE, Even C, Worden F, Saba NF, Iglesias Docampo LC, et al. Nivolumab for Recurrent Squamous-Cell Carcinoma of the Head and Neck. N Engl J Med. 2016; 375:1856–67. <u>https://doi. org/10.1056/NEJMoa1602252. [PubMed]</u>

- Gettinger SN, Horn L, Gandhi L, Spigel DR, Antonia SJ, Rizvi NA, Powderly JD, Heist RS, Carvajal RD, Jackman DM, Sequist LV, Smith DC, Leming P, et al. Overall Survival and Long-Term Safety of Nivolumab (Anti-Programmed Death 1 Antibody, BMS-936558, ONO-4538) in Patients With Previously Treated Advanced Non-Small-Cell Lung Cancer. J Clin Oncol. 2015; 33:2004–12. <u>https://</u> doi.org/10.1200/JCO.2014.58.3708. [PubMed]
- Larkin J, Chiarion-Sileni V, Gonzalez R, Grob JJ, Cowey CL, Lao CD, Schadendorf D, Dummer R, Smylie M, Rutkowski P, Ferrucci PF, Hill A, Wagstaff J, et al. Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. N Engl J Med. 2015; 373:23–34. https://doi.org/10.1056/NEJMoa1504030. [PubMed]
- Motzer RJ, Rini BI, McDermott DF, Redman BG, Kuzel TM, Harrison MR, Vaishampayan UN, Drabkin HA, George S, Logan TF, Margolin KA, Plimack ER, Lambert AM, et al. Nivolumab for Metastatic Renal Cell Carcinoma: Results of a Randomized Phase II Trial. J Clin Oncol. 2015; 33:1430– 37. https://doi.org/10.1200/JCO.2014.59.0703. [PubMed]
- Wolchok JD, Kluger H, Callahan MK, Postow MA, Rizvi NA, Lesokhin AM, Segal NH, Ariyan CE, Gordon RA, Reed K, Burke MM, Caldwell A, Kronenberg SA, et al. Nivolumab plus ipilimumab in advanced melanoma. N Engl J Med. 2013; 369:122–33. <u>https://doi.org/10.1056/</u> NEJMoa1302369. [PubMed]
- Spira A, Beane J, Shah V, Liu G, Schembri F, Yang X, Palma J, Brody JS. Effects of cigarette smoke on the human airway epithelial cell transcriptome. Proc Natl Acad Sci U S A. 2004; 101:10143–48. <u>https://doi.org/10.1073/ pnas.0401422101</u>. [PubMed]
- Riaz N, Havel JJ, Makarov V, Desrichard A, Urba WJ, Sims JS, Hodi FS, Martín-Algarra S, Mandal R, Sharfman WH, Bhatia S, Hwu WJ, Gajewski TF, et al. Tumor and Microenvironment Evolution during Immunotherapy with Nivolumab. Cell. 2017; 171:934–949.e16. <u>https://doi. org/10.1016/j.cell.2017.09.028</u>. [PubMed]
- Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, Hoang CD, Diehn M, Alizadeh AA. Robust enumeration of cell subsets from tissue expression profiles. Nat Methods. 2015; 12:453–57. <u>https://doi.org/10.1038/nmeth.3337</u>. [PubMed]

- Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N. Molecular and genetic properties of tumors associated with local immune cytolytic activity. Cell. 2015; 160:48–61. <u>https://doi.org/10.1016/j.cell.2014.12.033</u>. [PubMed]
- Şenbabaoğlu Y, Gejman RS, Winer AG, Liu M, Van Allen EM, de Velasco G, Miao D, Ostrovnaya I, Drill E, Luna A, Weinhold N, Lee W, Manley BJ, et al. Tumor immune microenvironment characterization in clear cell renal cell carcinoma identifies prognostic and immunotherapeutically relevant messenger RNA signatures. Genome Biol. 2016; 17:231. <u>https://doi.org/10.1186/s13059-016-1092-z</u>. [PubMed]
- Shen R, Seshan VE. FACETS: allele-specific copy number and clonal heterogeneity analysis tool for high-throughput DNA sequencing. Nucleic Acids Res. 2016; 44:e131. https://doi.org/10.1093/nar/gkw520. [PubMed]
- Chen B, Khodadoust MS, Liu CL, Newman AM, Alizadeh AA. Profiling Tumor Infiltrating Immune Cells with CIBERSORT. Methods Mol Biol. 2018; 1711:243–59. <u>https://doi.org/10.1007/978-1-4939-7493-1\_12</u>. [PubMed]
- Zhou H, Saliba J, Sandusky GE, Sears CR. XPC protects against smoking- and carcinogen-induced lungadenocarcinoma. Carcinogenesis. 2019; 40:403–411. <u>https://doi.org/10.1093/carcin/bgz003</u>. [PubMed]
- Balansky R, Ganchev G, Iltcheva M, Steele VE, D'Agostini F, De Flora S. Potent carcinogenicity of cigarette smoke in mice exposed early in life. Carcinogenesis. 2007; 28:2236– 43. <u>https://doi.org/10.1093/carcin/bgm122</u>. [PubMed]
- Woo SR, Corrales L, Gajewski TF. The STING pathway and the T cell-inflamed tumor microenvironment. Trends Immunol. 2015; 36:250–56. <u>https://doi.org/10.1016/j.</u> it.2015.02.003. [PubMed]
- 34. Poźniak J, Nsengimana J, Laye JP, O'Shea SJ, Diaz JM, Droop AP, Filia A, Harland M, Davies JR, Mell T, Randerson-Moor JA, Muralidhar S, Hogan SA, et al. Genetic and Environmental Determinants of Immune Response to Cutaneous Melanoma. Cancer Res. 2019; 79:2684–96. <u>https://doi.org/10.1158/0008-5472.CAN-18-2864. [PubMed]</u>