www.impactjournals.com/oncotarget

Research Paper

Quantitative image variables reflect the intratumoral pathologic heterogeneity of lung adenocarcinoma

E-Ryung Choi¹, Ho Yun Lee¹, Ji Yun Jeong², Yoon-La Choi³, Jhingook Kim⁴, Jungmin Bae¹, Kyung Soo Lee¹, Young Mog Shim⁴

¹Department of Radiology and Center for Imaging Science, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

²Department of Pathology, Kyungpook National University Hospital, Kyungpook National University School of Medicine, Daegu, Korea

³Department of Pathology, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

⁴Department of Thoracic and Cardiovascular Surgery, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

Correspondence to: Ho Yun Lee, email: hoyunlee96@gmail.com

Keywords:lung adenocarcinoma, heterogeneity, radiomics, quantitative image variables, dual energy CTReceived:May 12, 2016Accepted:July 19, 2016Published:August 30, 2016

ABSTRACT

We aimed to compare quantitative radiomic parameters from dual-energy computed tomography (DECT) of lung adenocarcinoma and pathologic complexity.

A total 89 tumors with clinical stage I/II lung adenocarcinoma were prospectively included. Fifty one radiomic features were assessed both from iodine images and noncontrast images of DECT datasets. Comprehensive histologic subtyping was evaluated with all surgically resected tumors. The degree of pathologic heterogeneity was assessed using pathologic index and the number of mixture histologic subtypes in a tumor. Radiomic parameters were correlated with pathologic index. Tumors were classified as three groups according to the number of mixture histologic subtypes and radiomic parameters were compared between the three groups.

Tumor density and 50th through 97.5th percentile Hounsfield units (HU) of histogram on non-contrast images showed strong correlation with the pathologic heterogeneity. Radiomic parameters including 75th and 97.5th percentile HU of histogram, entropy, and inertia on 1-, 2- and 3 voxel distance on non-contrast images showed incremental changes while homogeneity showed detrimental change according to the number of mixture histologic subtypes (all *Ps* < 0.05).

Radiomic variables from DECT of lung adenocarcinoma reflect pathologic intratumoral heterogeneity, which may help in the prediction of intratumoral heterogeneity of the whole tumor.

INTRODUCTION

Lung cancer is the most commonly diagnosed cancer worldwide and the leading cause of cancer-related death [1], and adenocarcinoma is the most common histologic subtype of lung cancer in most countries [2]. In an attempt to reflect the widely divergent pathologic spectrum of lung adenocarcinoma, lung adenocarcinoma classification criteria were proposed by the International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society (IASLC/ATS/ ERS) [3]. This classification was devised to understand the histological subtypes and their histo-molecular correlations. Since the release of this classification, many studies have investigated the possible correlations among the most predominant subtypes, driver mutations and patient prognosis [4, 5]. However, prognostic stratification considering only the most predominant subtypes has shown substantial limitation due to the fact that more than 80% of invasive lung adenocarcinomas show mixed type including two or more of histologic subtypes [6–9].

Recently, scientists have performed quantitative imaging of lung cancer using primarily a radiomic approach, demonstrating that radiomic values quantifying spatial variation in architecture have shown prognostic significance [10–12]. Based on these results, they suggested that imaging features depicting spatial heterogeneity in tumors might reflect genomic and phenotypic intratumoral heterogeneity, which has significant implications for treatment, resistance, and, ultimately, prognosis [13–15]. Surprisingly, there have not been any studies directly evaluating the relationship between radiologic heterogeneity values and pathologic complexity within lung adenocarcinoma. Thus, we aimed to correlate various quantitative radiomic parameters from dual-energy computed tomography (DECT) of lung adenocarcinomas with pathologic complexity to ultimately identify the role of quantitative image variables in predicting pathologic heterogeneity.

RESULTS

Clinical characteristics of patients and tumors

All 92 consecutive patients were enrolled and underwent DECT for work-up (Figure 1). We excluded three patients who were shown to have benign disease after percutaneous lesion biopsy and five patients who were found to have unresectable stage III or IV lung cancer through further studies. Eight-four patients underwent complete resection for ninety-three lesions. Of those, we excluded four who had one benign disease, one who had mucinous adenocarcinoma, and two who had insufficient pathologic slides for detailed pathologic review. In total, 80 patients with 89 lung adenocarcinomas were included in our study.

There were total 6 patients who have more than one primary lesion arising in the lung at the same time. Three patients had 2 tumors in the lung and three patients had 3 tumors in the lung. Among these patients, four had nodules in the same lobe. However, multiple tumors in the same lobe showed complete different solidity on CT scan suggesting intendent relationships more likely than the metastasis. Pathological result revealed these tumors were synchronous tumors. The most predominant histologic subtype was the acinar subtype (59.6%) followed by lepidic subtype (20.2%) (Table 1). The most frequent combinations in mixed type were lepidic and acinar subtypes (14.6%). Final pathologic staging revealed 84 stage I tumors, one stage II tumor, and four stage III tumors. The clinicopathologic characteristics of the 89 lung adenocarcinomas included in this study are summarized in Table 1. Also, the relationship between size and volume of tumors are included in Supplementary Figure S1.

Correlation between radiomic features and pathology heterogeneity index

There was strong relationship between the prediction model made with selected radiomic variables and pathologic heterogeneity index (R = 0.936, p = 0.001).

The relationships between 51 radiomic parameters and the pathologic heterogeneity index are described

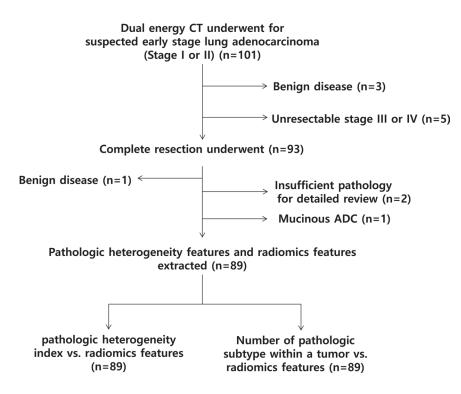


Figure 1: Flow diagram of the patient cohort.

in Table 2. Among the radiomic parameters of physical features, density on non-contrast images showed a strong correlation with pathologic heterogeneity (R =0.654, P < 0.001). Also, pathologic heterogeneity was robustly associated with 50th percentile, 75th percentile and 97.5th percentile of HU among histogram CT parameters on non-contrast images (R = 0.648, R =0.663, R = 0.626, respectively; all Ps < 0.001). The size of the tumor on iodine-contrast images shows moderate positive correlation (R = 0.527, P < 0.001). The skewness of histogram features showed a moderate negative correlation with histologic complexity on both non-contrast and iodine-contrast images (R = -0.532, R = -0.531, respectively; all Ps < 0.001). There was no strong association between regional features. Among local features, variance and inertia on non-contrast image were moderately associated with pathologic complexity (variance on 1 voxel distance, R = -0.522; variance on 2 voxel distance, R = -0.517; variance on 3 voxel distance, R = -0.521; inertia on 1 voxel distance, R = 0.549; inertia on 2 voxel distance, R = 0.550; inertia on 3 voxel distance, R = 0.574; all Ps < 0.001).

Comparison of three different groups based on pathologic heterogeneity and radiomic parameters

Among histogram features on non-contrast CT images, the 75th percentile and 97.5th percentile HU of the histogram showed significantly increased mean CT values according to the number of histologic subtypes (P = 0.003, P = 0.002, respectively) (Table 3).

Of all regional features, entropy showed an escalation of the mean CT values with an increased number of histologic subtypes on non-contrast images with statistical significance (P = 0.003). Although those were not statistically significant, intensity variability and size-zone variability on the non-contrast images and entropy on the iodine images presented increased mean CT values according to the number of histologic subtypes.

Among local features on non-contrast images, the mean CT values of inertia increased while those of homogeneity demonstrate a declined as the number of histologic subtypes increased with statistical significance (Inertia on 1 voxel distance, P = 0.027; Inertia on 2 voxel distance, P = 0.024; Inertia on 3 voxel distance, P = 0.018; homogeneity on 1 voxel distance, P = 025; homogeneity on 2 voxel distance, P = 023; homogeneity on 3 voxel distance, P = 0.008). On iodine-enhanced images, the mean CT values of energy and maximum probability value increased while those of contrast, sum mean, cluster shade, and homogeneity showed a decline when the number of histologic subtypes increased. However, the tendencies of these values were failed to show the statistical significance. All radiomic parameters of the three different groups based on pathologic heterogeneity are described in Table 3.

DISCUSSION

Recent lung cancer research has demonstrated that cancerous cells not only undergo clonal evolution from a single progenitor, but also exhibit branched evolution, whereby each tumor develops and preserves multiple distinct subclonal compositions [16-19]. This intratumoral genetic heterogeneity consequently leads to phenotypic differences in histopathological divergence, containing regions demarcated by various degrees of differentiation, proliferation, vascularity, and invasiveness [20]. Given the existence of such heterogeneity in tumors in advanced metastatic disease, the efficacy of therapies targeting somatic driver aberrations may be limited [13]. Recent oncology research has mostly focused on the intratumoral variation in gene mutation and expression, whereas few studies have explored the spatial relationship among imaging, genomics, and histopathology.

While imaging is a main tool in tumor staging, progression assessment and recurrence detection, most radiologic approaches deal only with tumor size or average parameter values [12] in routine oncologic practice and research. Considerable effort has explored sophisticated and robust analyses to quantify tumor spatial complexity with tumor imaging data [10, 21–25]. This approach usually uses a texture analysis method to quantify the spatial variation as the remaining spatial arrangement of voxel values in imaging data [12], which could serve as a potential prognostic biomarker [11, 26]. Most studies of radiomic analyses have evaluated the role of radiomics in the prognostic stratification [12], but there have not yet been any studies correlating intratumoral heterogeneity on CT images with pathology heterogeneity. Radiomic analysis is emerging as a method to quantify spatial variation within tumors. Establishment of a histologic correlation would obviously be an important step in the validation of this approach. Our study is the first study to determine a direct relationship between the histological complexity of lung adenocarcinoma and intratumoral heterogeneity expressed as CT radiomic values.

We adopted histogram, local and regional features to measuring degree of intratumoral heterogeneity. Histogram features were derived from the density distribution of the tumor reflecting heterogeneity of intratumoral density. The spatial arrangement of voxel values is obtained from local features. We included tumor size as a routine practical measurement variable and compared with other all radiomic variables in the analysis. Radiomic variables were more strongly correlated with pathologic heterogeneity rather than tumor size. In

Chara	cteristics		
Sex (%	(0)		
Mal	e	39 (48.8)	
Fem	ale	41 (51.2)	
Age, r	ange (median)	37-78 (59)	
	r size (mm), (median)	4-64 (21)	
Stagin	g		
Ι		84 (94.4)	
II		1 (1.1)	
III		4 (4.5)	
T cate	gory		
1		73 (82.1)	
2		14 (15.7)	
3		2 (2.2)	
N cate	gory		
0		82 (92.1)	
1		5 (5.6)	
2		2 (2.2)	
Exten	t of resection (%)		
Seg	mentectomy	50 (56.2)	
Lobectomy		39 (43.8)	
	predominant ogic subtype		
Lep	idic	18 (20.2)	
Acir	nar	53 (59.6)	
Papi	illary	11 (12.4)	
Soli	d	1 (1.1)	
Mic	ropapillary	6 (6.7)	
No. of	histologic subtype	s within a tumor	
1	19 (21.3)	Lepidic	5 (26.3)
		Acinar	11 (57.9)
		Papillary	3 (15.8)
2	54 (60.7)	Lepidic + Acinar	31 (57.4)
		Acinar + Papillary	13 (24.1)
		Acinar + Micropapillary	1 (1.9)
		Acinar + Solid	9 (16.7)
3	14 (15.7)	Lepidic + Acinar + Papillary	6 (42.9)
		Lepidic + Acinar + Solid	2 (14.3)
		Acinar + Papillary + Micropapillary	4 (28.6)
		Acinar + Micropapillary + Solid	2 (14.3)
4	2 (2.2)	Lepidic + Acinar + Papillary + Micropapillary	2 (100)

Table 1: Clinicopathologic characteristics of l	ing adenocarcinoma	(89 tumors from 80 patients)
---	--------------------	------------------------------

CT nonomotors	Non-contras	Iodine-contrast images		
CT parameters	R	Р	R	Р
Physical features				
Volume (cm ³)	0.233	0.028*	0.177	0.096
Density (HU)	0.653**	< 0.001*	0.139	0.195
Mass (g)	0.352	0.001*	0.183	0.086
Size (mm)	0.252	0.017	0.527**	< 0.001*
Histogram features				
Skewness	-0.532**	< 0.001*	-0.531**	< 0.001*
Kurtosis	-0.059	0.584	-0.313	0.003*
2.5th percentile (HU)	0.424	< 0.001*	-0.423	< 0.001*
25th percentile (HU)	0.584**	< 0.001*	0.099	0.356
50th percentile (HU)	0.648**	< 0.001*	0.254	0.016*
75th percentile (HU)	0.663**	< 0.001*	0.246	0.020*
97.5th percentile (HU)	0.626**	< 0.001*	-0.180	0.091
Regional features				
Uniformity	-0.195	0.067	-0.140	0.190
Entropy	0.297	0.005*	0.131	0.223
Intensity variability	0.276	0.009*	0.368	< 0.001*
Size-zone variability	0.245	0.021*	0.171	0.108
Local features				
Energy on 1 voxel distance	0.311	0.003*	-0.073	0.496
Energy on 2 voxel distance	0.249	0.020*	-0.060	0.579
Energy on 3 voxel distance	0.184	0.087	-0.134	0.211
Entropy on 1 voxel distance	-0.192	0.073	0.132	0.219
Entropy on 2 voxel distance	-0.112	0.298	0.117	0.276
Entropy on 3 voxel distance	-0.065	0.550	0.193	0.070
Correlation on 1 voxel distance	-0.127	0.239	-0.100	0.350
Correlation on 2 voxel distance	0.009	0.934	0.029	0.791
Correlation on 3 voxel distance	0.112	0.299	0.063	0.559
Contrast on 1 voxel distance	-0.032	0.770	0.061	0.568
Contrast on 2 voxel distance	0.012	0.914	-0.028	0.793
Contrast on 3 voxel distance	0.073	0.500	-0.039	0.716
Variance on 1 voxel distance	-0.522**	< 0.001*	-0.385	< 0.001*
Variance on 2 voxel distance	-0.517**	< 0.001*	-0.382	< 0.001*
Variance on 3 voxel distance	-0.521**	< 0.001*	-0.409	< 0.001*
Sum mean on 1 voxel distance	0.238	0.025*	0.027	0.804
Sum mean on 2 voxel distance	0.222	0.038*	-0.014	0.9

Table 2: Correlation of radiomic features with pathologic heterogeneity index

(Continued)

CT.	Non-contras	t images	Iodine-contrast images		
CT parameters –	R	Р	R	Р	
Sum mean on 3 voxel distance	0.235	0.027*	-0.017	0.875	
Inertia on 1 voxel distance	0.549**	< 0.001*	0.391	< 0.001*	
Inertia on 2 voxel distance	0.550**	< 0.001*	0.384	< 0.001*	
Inertia on 3 voxel distance	0.574**	< 0.001*	0.401	< 0.001*	
Cluster shade on 1 voxel distance	-0.032	0.770	0.061	0.568	
Cluster shade on 2 voxel distance	0.012	0.914	-0.028	0.793	
Cluster shade on 3 voxel distance	0.073	0.500	-0.039	0.716	
Cluster tendency on 1 voxel distance	-0.454	< 0.001*	-0.434	< 0.001*	
Cluster tendency on 2 voxel distance	-0.438	< 0.001*	-0.420	< 0.001*	
Cluster tendency on 3 voxel distance	-0.448	< 0.001*	-0.470	< 0.001*	
Homogeneity on 1 voxel distance	0.406	< 0.001*	-0.207	0.052	
Homogeneity on 2 voxel distance	0.398	< 0.001*	-0.129	0.230	
Homogeneity on 3 voxel distance	0.411	< 0.001*	-0.109	0.308	
Maximum probability on 1 voxel distance	0.447	< 0.001*	-0.041	0.700	
Maximum probability on 2 voxel distance	0.403	< 0.001*	-0.042	0.698	
Maximum probability on 3 voxel distance	0.323	0.002*	-0.090	0.400	
Inverse variance on 1 voxel distance	-0.336	0.001*	-0.102	0.343	
Inverse variance on 2 voxel distance	-0.124	0.250	-0.005	0.963	
Inverse variance on 3 voxel distance	-0.048	0.659	-0.023	0.833	

R indicates the Spearman correlation coefficient. *P < 0.05, ** R > 0.5

addition, these radiomic features may offer minimal interand intra-reader variability with high reproducibility of imaging features.

Our results clearly demonstrate histopathologic complexity correlated with radiomic parameters such as texture features within CT images of lung adenocarcinoma. Quantifying the spatial complexity of tumor images can explain the spatial complexity in pathology, or in other words, the phenotypic intratumoral heterogeneity.

Radiomic features derived from non-contrast images reflect tumor cellularity and density, whereas iodineenhanced image even allows additional information about tumor vascularity. Therefore, we originally expected that radiomic features of iodine-contrast images might have associated more strongly to the tumor complexity related with heterogeneity of tumor density and tumor vascularity. However, in our analysis, the most of radiomic features which correlated to pathologic heterogeneity index were features derived from non-contrast image, not from iodine-enhanced image. This result can be explained in part by the concept of tumor microenvironment where whole tumor volume consists of real tumor component as well as nontumorous stromal component. Thus, radiomic variables extracted from non-contrast CT covering completely density or cellularity of ROI of the tumor may reflect more actually spatial heterogeneity in the entire tumor ROI, compared to radiomic variables from iodine image limitedly enhancing heterogeneity of only tumorous component. The problem is that results from small biopsy tissues in non-resectable lung adenocarcinomas do not represent the pathologic heterogeneity of the whole tumor. On the other hand, quantifying imaging data using a radiomic approach indirectly enables assessment of whole tumor heterogeneity. Quantifying spatial heterogeneity on a tumor image may help with determining the prognosis and stratifying patients with non-resectable lung adenocarcinoma for which whole tumor pathology is not available. Furthermore, intratumoral heterogeneity may have important consequences for personalized medicine approaches that typically rely on a single tumor-biopsy to portray the tumor mutational landscape. Quantitative radiomic variables could allow physicians to deliver more optimized, patient-specific treatment by allowing them to identify patients who need aggressive treatment.

	Non-contrast images			Iodine-contrast images			_	
	One subtype	Two subtypes	Three or four subtypes	р	One subtype	Two subtypes	Three or four subtypes	р
Physical features								
Volume (cm ³)	6.97±2.14	8.29±2.81	7.93±5.900	0.966	7.08±12.62	4.67±5.85	6.50±5.10	0.432
Density (HU)	0.57±0.23	0.566±0.22	0.75±0.155	0.008*	1.17±0.016	1.16±0.019	1.17±0.01	0.618
Mass (g)	5.30±10.35	4.572±9.00	6.00±4.484	0.832	8.25±14.71	5.44±6.78	7.61±5.95	0.426
Size (mm)	23.11±16.86	21.02±9.26	24.41±10.89	0.488	12.44±18.26	9.83±12.02	17.18±8.85	0.130
Histogram features								
Skewness	-0.21±0.66	0.012±0.721	-0.489±0.616	0.037*	0.461±0.822	0.782±1.04	0.056±0.598	0.024
Kurtosis	2.75±0.96	2.76±1.01	2.47±1.108	0.598	4.39±2.32	5.60±4.33	4.51±3.29	0.377
2.5 th percentile (HU)	-797.76±71.95	-814.99±70.14	-769.28 ±74.37	0.080	-21.07±14.38	-23.42±16.68	-31.39 ±14.34	0.131
25 th percentile (HU)	-576.08±193.86	-604.91±181.72	-484.64 ±181.10	0.078	25.31±13.29	21.15±16.143	26.20±15.66	0.397
50 th percentile (HU)	-431.21±232.06	-450.78±234.06	-251.37 ±171.56	0.009*	52.50±16.20	47.94±19.68	56.62±5.35	0.220
75 th percentile (HU)	-316.05±241.84	-305.18±232.75	-91.90±127.34	0.003*	79.94±17.36	76.38±22.41	83.68±16.05	0.429
97.5 th percentile (HU)	-181.87±241.84	-128.25±189.22	32.48±39.52	0.002*	152.31±39.92	156.08±35.30	143.65±27.89	0.464
Regional features								
Uniformity	0.003±0.002	0.002±0.001	0.002±0.001	0.042*	0.009±0.002	0.009±0.002	0.008±0.002	0.365
Entropy	8.76±0.73	9.08±0.50	9.41±0.37	0.003*	7.17±0.37	7.24±0.32	7.36± 0.31	0.258
Intensity variability	7.56±3.71	8.49±3.8	9.57±3.53	0.294	7.27±3.18	6.1563.29	8.03±3.27	0.099
Size-zone variability	13.14±12.21	15.33±11.86	18.59±7.81	0.367	14.42±14.94	14.22±9.44	17.99±8.60	0.454
Local features								
Energy on 1 voxel distance	0.040±0.050	0.034±0.033	0.051±0.067	0.410	0.037±0.017	0.046±0.030	0.049±0.036	0.394
Energy on 2 voxel distance	0.036±0.046	0.027±0.028	0.042 ± 0.064	0.356	0.033±0.014	0.040±0.025	0.043 ± 0.033	0.493
Energy on 3 voxel distance	0.031±0.025	0.024±0.026	0.039± 0.061	0.337	0.036±0.014	0.040±0.025	0.042 ± 0.033	0.784
Entropy on 1 voxel distance	1.74±0.23	1.79±0.19	1.76± 0.24	0.580	1.65±0.15	1.62±0.18	1.59±0.22	0.697
Entropy on 2 voxel distance	1.82±0.24	1.91±0.20	1.89±0.26	0.306	1.66±0.13	1.66±0.17	1.63±0.22	0.869
Entropy on 3 voxel distance	1.81±0.22	1.93±0.22	1.92±0.28	0.128	1.62±0.15	1.65±0.16	1.63±0.22	0.798
Correlation on 1 voxel distance	0.089±0.037	0.091±0.046	0.079±0.036	0.658	0.109±0.062	0.121±0.059	0.154±0.14	0.231
Correlation on 2 voxel distance	0.047±0.030	0.053±0.024	0.056±0.027	0.555	0.034±0.027	0.036±0.023	0.052±0.057	0.190
Correlation on 3 voxel distance	0.025±0.038	0.030±0.019	0.040 ± 0.020	0.231	0.021±0.019	0.017±0.015	0.027±0.030	0.203
Contrast on 1 voxel distance	5.30±2.23	4.76±2.04	4.47±1.90	0.478	5.51±2.83	4.98±3.13	4.32±2.06	0.492
Contrast on 2 voxel distance	11.16±4.60	10.45±4.24	10.62±5.01	0.834	8.13±4.25	7.29±3.87	6.09± 2.88	0.290
Contrast on 3 voxel distance	14.71±6.75	14.29±5.67	15.48± 8.17	0.807	8.62±4.36	7.94±4.19	6.38± 2.86	0.248
Variance on 1 voxel distance	0.17±0.11	0.163±0.054	0.129± 0.029	0.114	0.176±0.053	0.185±0.052	0.175±0.054	0.704
Variance on 2 voxel distance	0.17±0.10	0.161±0.054	0.129± 0.032	0.121	0.177±0.052	0.186±0.051	0.177± 0.056	0.718
Variance on 3 voxel distance	0.16±0.06	0.162±0.054	0.131 ± 0.038	0.111	0.179±0.050	0.188±0.051	0.179± 0.058	0.765
Sum mean on 1 voxel distance	8.83±3.47	9.60±3.41	11.544±3.509	0.061	4.37±4.05	4.05±1.92	3.48±1.43	0.351
Sum mean on 2 voxel distance	8.90±3.51	9.68±3.48	11.354± 3.407	0.111	4.37±1.92	4.02±1.87	3.40±1.46	0.285

Table 3: Radiomic features according to the number of histologic subtypes within a tumor

(*Continued*)

	Non-contrast images				Iodine-contrast images			
	One subtype	Two subtypes	Three or four subtypes	р	One subtype	Two subtypes	Three or four subtypes	р
Sum mean on 3 voxel distance	8.46±3.06	9.79±3.57	11.369± 3.468	0.051	4.27±1.75	4.02±1.77	3.36± 1.44	0.267
Inertia on 1 voxel distance	8.37±2.23	8.43±2.28	$10.057{\pm}1.532$	0.027*	6.87±1.71	6.50±1.64	6.85±1.71	0.617
Inertia on 2 voxel distance	8.45±2.26	8.54±2.31	10.20±1.54	0.024*	6.82±1.66	6.47±1.60	6.82±1.70	0.621
Inertia on 3 voxel distance	8.39±2.26	8.54±2.31	10.247 ± 1.554	0.018*	6.70±1.56	6.44±1.56	6.78±1.69	0.672
Cluster shade on 1 voxel distance	5.30±2.23	4.76±2.04	4.479±1.903	0.478	5.51±2.83	4.98±3.13	4.33±2.07	0.492
Cluster shade on 2 voxel distance	11.16±4.60	10.43±4.24	10.629±5.011	0.834	8.14±4.26	7.30±5.68	6.10±2.88	0.290
Cluster shade on 3 voxel distance	14.71±6.75	14.29±5.67	15.480 ± 8.177	0.807	8.62±4.37	7.941±3.87	6.38± 2.86	0.248
Cluster tendency on 1 voxel distance	-43.74±16.58	-42.39±50.95	-108.582±29.33	0.244	14.25±29.92	14.76±27.81	3.07±20.57	0.309
Cluster tendency on 2 voxel distance	-37.71±86.04	-36.77±116.02	-82.633 ± 04.72	0.318	7.57±18.33	8.30±16.88	1.58±15.04	0.374
Cluster tendency on 3 voxel distance	-33.78±64.46	-32.49±95.11	-70.196±88.10	0.314	7.34±15.93	6.95±13.80	1.56±14.53	0.388
Homogeneity on 1 voxel distance	4532.07±1777.15	3387.64±2166.78	2629.69±181.00	0.025*	629.72±389.37	565.77±348.42	386.11± 232.51	0.094
Homogeneity on 2 voxel distance	3480.25±1460.08	2523.03±1782.39	1899.41±1225.18	0.023*	375.41±229.52	341.68±234.28	243.52±173.64	0.197
Homogeneity on 3 voxel distance	2895.88±1303.93	2039.34±1521.84	1380.43±1060.43	0.008*	314.92±192.11	289.38±191.10	215.53±177.30	0.273
Maximum probability on 1 voxel distance	0.109±0.110	0.092±0.079	0.137 ± 0.125	0.248	0.085±0.041	0.110±0.064	$0.116{\pm}\ 0.067$	0.230
Maximum probability on 2 voxel distance	0.095±0.102	0.072±0.071	0.117 ± 0.123	0.189	0.076±0.033	0.095±0.055	$0.101{\pm}0.062$	0.308
Maximum probability on 3 voxel distance	0.081±0.0619	0.064±0.065	0.107 ± 0.120	0.154	0.078±0.029	0.092±0.051	$\textbf{0.096}{\pm}~\textbf{0.061}$	0.491
Inverse variance on 1 voxel distance	0.381±0.063	0.402±0.045	$0.381{\pm}0.051$	0.153	0.413±0.050	0.417±0.045	0.416±0.031	0.952
Inverse variance on 2 voxel distance	0.316±0.060	0.339±0.049	0.322 ± 0.047	0.185	0.377±0.066	0.386±0.054	0.396±0.046	0.598
Inverse variance on 3 voxel distance	0.294±0.055	0.305±0.051	0.289 ± 0.049	0.487	0.372±0.059	0.375±0.056	0.390±0.047	0.558

Features showing uniform tendency are expressed in bold font. *P < 0.05.

In addition, radiomic parameters might be useful for noninvasive *in vivo* monitoring of longitudinal changes in tumor heterogeneity. Lung cancer cells within tumors show homogeneous cell populations until relatively late in tumor progression, when hyperproliferation and increased genetic instability result in distinct clonal subpopulations [18, 19]. Several studies have shown that mechanisms of acquired drug resistance to EGFR inhibitors are related to several genotypic and phenotypic changes [27, 28]. Using a radiogenomic approach to quantify tumor heterogeneity on images may allow longitudinal studies of clonal evolution during treatment as well.

Our study had several limitations. First, the proportion of the tumor with three or four mixtures of histologic subtypes was relatively small. The relationship of the quantitative radiomic variables and pathologic heterogeneity and the values of the quantitative radiomic variables may be influenced by the sample size. Second, all cases were collected from a single institute. Larger prospective studies from multiple centers are needed. Third, we excluded patients with variant subtypes including mucinous pattern. However, we decided to exclude this subtype because reports regarding the survival of mucinous lung adenocarcinomas are limited [29, 30].

In conclusion, various radiomic variables from DECT of lung adenocarcinoma reflect pathologic intratumoral heterogeneity, which may be a helpful predictor of intratumoral heterogeneity of the whole tumor, considering that current genomic analyses are limited by the fact that they rely on a single tumor biopsy.

MATERIALS AND METHODS

Patients

This study was performed as part of an ongoing prospective clinical trial aimed at determining the value of imaging biomarkers for the prediction of tumor aggressiveness and prognosis in patient with operable lung adenocarcinoma (NCT01482585). This study was approved by the institutional review board (SMC 2011-09-083) and written informed consent was obtained.

From November 2011 to December 2012, a total of 92 patients with operable lung adenocarcinoma were eligible for our study. The inclusion criteria of our study were as follows: (1) Clinically and radiologically suspected lung adenocarcinoma, (2) Newly diagnosed stage I or II disease from clinical work-up including F-18fluorodeoxyglucose (FDG) positron emission tomography (PET)/CT, (3) Eastern Cooperative Oncology Group (ECOG) performance status of 0 or 1 and eligible for surgery, (4) Age 20 years or older, (5) Able to tolerate DECT imaging as required per protocol, and (6) Able to give study-specific informed consent. The exclusion criteria were: (1) Prior malignancy, (2) Scheduled for definitive radiation therapy or neoadjuvant concurrent chemoradiation therapy, and (3) Poor cardiopulmonary reserve (a contraindication for surgery).

CT imaging and analysis

Patients underwent CT examination using a dualsource CT scanner (Somatom Definition Flash; Siemens Medical Solutions, Forchheim, Germany) with the dualenergy technique. Three types of data set were generated from the DECT scanning: 80 kV, 140 kV, and enhanced weighted-average images. See the supplement and Supplementary Figure S2 for further details about the image acquisition protocol and reconstruction process.

Virtual non-enhanced images and iodine-enhanced images were generated using the liver Virtual Non-Contrast (VNC) application node of dedicated dual-energy post-processing software (Syngo Dual Energy; Siemens Medical Solution, Forchheim, Germany). To obtain the iodine value of both the solid and ground-glass opacity (GGO) component in each tumor, post-processing was performed with two different types of software. Image data were reconstructed with a section thickness of 1 mm using a D30f (medium smooth) kernel for the iodineenhanced image and a D45f (medium smooth) kernel for the virtual non-enhanced image.

In the quantitative analysis, regions of interest (ROIs) were delineated on the axial images to generate a volume of interest which included the entire tumor. Initially, we assessed the stability of all 51 radiomic parameters for which we performed the concordance correlation coefficients (CCC) regarding radiomic values extracted from two ROIs drawn by two radiologists in 25 randomly selected patients. From the stability test, we found that all radiomic parameters were stable, where all CCCs were very high or high (range, 0.888 – 0.999; mean, 0.934; SD, 0.008). As a next step, all included patient were handled by one radiologist.

Fifty one quantitative radiomic features were derived from this ROI to evaluate the heterogeneity of the tumors (Figure 2). The physical features included volume, size, density and mass. The histogram features included skewness, kurtosis, and Hounsfield units (HU) at the 2.5th, 25th, 50th, 75th and 97.5th percentiles. The regional features included uniformity, entropy, intensity variability and size-zone variability. There were 36 local texture features, including energy, entropy, correlation, contrast, variance, sum mean, inertia, cluster shade, cluster tendency, homogeneity, maximum probability, and inverse variance. Each local texture feature was derived from 13 directions according to 1-voxel, 2-voxel, and 3-voxel distances at the gray value for each voxel. These radiomic features were evaluated both on non-contrast images and iodine contrast images. See the supplement for further details regarding the extraction of quantitative radiomic parameters.

Pathologic evaluation and analysis

For tumor sampling, tumor tissues with an interval of per approximately 10 mm were taken from the tumor specimen and placed on a slide. All slides were scanned to produce a high- resolution digital image (0.25 µm/pixel at 40× magnification) using the Aperio Slide Scanning System (ScanScope T3; Aperio Technologies Inc., Vista, CA, USA). Two experienced lung pathologists with 13 and 18 years of experience (J.Y.J., Y.C.) interpreted all tissue sections by virtual slides using ImageScope viewing software (Aperio Technologies, Inc.) and a high-resolution monitor. For each case, the specimens were reviewed according to the International Association for the Study of Lung Cancer (IASLC), the American Thoracic Society (ATS), and the European Respiratory Society (ERS) International Multidisciplinary Lung Adenocarcinoma Classification Criteria [3] and comprehensive histologic subtyping was performed for a whole primary tumor in a semi-quantitative manner. The extent of existent tumor histologic subtypes and central fibrosis was quantified to the nearest 5% level, adding up to a total of 100% of the subtype components per tumor. They reported each of tumors as a relative ratio among total 100% in terms of all five with its whole histologic subtype.

The pathologic heterogeneity index was calculated from each tumor to evaluate the degree of pathologic heterogeneity, which was confirmed to reflect the survival predictive value according to the proportion of mixed histologic subtypes that were derived from the hazard ratio (HR) of each subtype using the disease-free survival curve of a large-scale study [4]. Further details about the pathologic heterogeneity index are described in the supplement.

Statistical analysis

A multivariate logistic regression model with the stepwise variable selection procedure based on Akaike's Information Criterion (AIC) was applied to validate the performance of selected radiomic features to predict actually pathologic heterogeneity. Ten-fold crossvalidation [31] was used to evaluate the performance of this prediction model. After then, Pearson correlation analysis was applied to compare the predictive values from made prediction model with pathologic heterogeneity in each. The Spearman correlation coefficient was used to determine the relationship between radiomic features and pathologic heterogeneity index. When statistically significant, an absolute Rho (ρ) value between 0.00 and 0.19 was considered very weak correlation, 0.20-0.39, weak, 0.40–0.59, moderate, 0.60-0.79 strong and greater than 0.80, strong [32]. Radiomic parameters were compared among the three different groups that had been

classified according to the number of histologic subtype within a tumor (e.g., tumor consistent with a single histologic subtype, two histologic subtypes, or three or four histologic subtypes) using ANOVA with Tukey's post hoc test. In the cases with multiple tumors in a patient, we did not take into account within-patient correlation because each tumor was considered an independent synchronous lesion [33].

Statistical significance was evaluated with software (SPSS, version 19.0, 2010; SPSS, Chicago, Ill). A P value less than 0.05 was considered to represent a statistically significant difference.

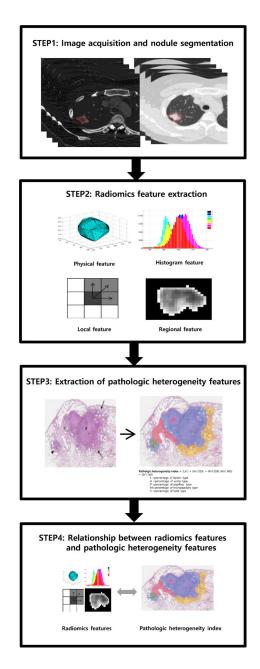


Figure 2: Radiomic data extraction and analysis workflow.

ACKNOWLEDGMENTS

None.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

GRANT SUPPORT

This study was supported by a grant from the Korean Foundation for Cancer Research (KFCR-CB-2011-02-02).

REFERENCES

- 1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. CA Cancer J Clin. 2013; 63:11-30.
- 2. Curado MP, International Agency for Research on C and International Association of Cancer R. Cancer incidence in five continents, Volume IX. Lyon; Geneva: International Agency for Research on Cancer ; Distributed by WHO Press, World Health Organization; 2008.
- 3. Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger KR, Yatabe Y, Beer DG, Powell CA, Riely GJ, Van Schil PE, Garg K, Austin JH, Asamura H, et al. International association for the study of lung cancer/ american thoracic society/european respiratory society international multidisciplinary classification of lung adenocarcinoma. J Thorac Oncol. 2011; 6:244-285.
- Hung JJ, Yeh YC, Jeng WJ, Wu KJ, Huang BS, Wu YC, Chou TY, Hsu WH. Predictive value of the international association for the study of lung cancer/American Thoracic Society/European Respiratory Society classification of lung adenocarcinoma in tumor recurrence and patient survival. J Clin Oncol. 2014; 32:2357-2364.
- Campos-Parra AD, Aviles A, Contreras-Reyes S, Rojas-Marin CE, Sanchez-Reyes R, Borbolla-Escoboza RJ, Arrieta O. Relevance of the novel IASLC/ATS/ERS classification of lung adenocarcinoma in advanced disease. Eur Respir J. 2014; 43:1439-1447.
- Motoi N, Szoke J, Riely GJ, Seshan VE, Kris MG, Rusch VW, Gerald WL, Travis WD. Lung adenocarcinoma: modification of the 2004 WHO mixed subtype to include the major histologic subtype suggests correlations between papillary and micropapillary adenocarcinoma subtypes, EGFR mutations and gene expression analysis. Am J Surg Pathol. 2008; 32:810-827.
- Kadota K, Colovos C, Suzuki K, Rizk NP, Dunphy MP, Zabor EC, Sima CS, Yoshizawa A, Travis WD, Rusch VW, Adusumilli PS. FDG-PET SUVmax combined with IASLC/ ATS/ERS histologic classification improves the prognostic stratification of patients with stage I lung adenocarcinoma. Ann Surg Oncol. 2012; 19:3598-3605.

- Lee HY, Jeong JY, Lee KS, Kim HJ, Han J, Kim BT, Kim J, Shim YM, Kim JH, Song I. Solitary pulmonary nodular lung adenocarcinoma: correlation of histopathologic scoring and patient survival with imaging biomarkers. Radiology. 2012; 264:884-893.
- Sica G, Yoshizawa A, Sima CS, Azzoli CG, Downey RJ, Rusch VW, Travis WD, Moreira AL. A grading system of lung adenocarcinomas based on histologic pattern is predictive of disease recurrence in stage I tumors. Am J Surg Pathol. 2010; 34:1155-1162.
- Win T, Miles KA, Janes SM, Ganeshan B, Shastry M, Endozo R, Meagher M, Shortman RI, Wan S, Kayani I, Ell PJ, Groves AM. Tumor heterogeneity and permeability as measured on the CT component of PET/CT predict survival in patients with non-small cell lung cancer. Clin Cancer Res. 2013; 19:3591-3599.
- Ganeshan B, Panayiotou E, Burnand K, Dizdarevic S, Miles K. Tumour heterogeneity in non-small cell lung carcinoma assessed by CT texture analysis: a potential marker of survival. Eur Radiol. 2012; 22:796-802.
- O'Connor JP, Rose CJ, Waterton JC, Carano RA, Parker GJ, Jackson A. Imaging intratumor heterogeneity: role in therapy response, resistance, and clinical outcome. Clin Cancer Res. 2015; 21:249-257.
- Jamal-Hanjani M, Quezada SA, Larkin J, Swanton C. Translational implications of tumor heterogeneity. Clin Cancer Res. 2015; 21:1258-1266.
- Gerlinger M, Rowan AJ, Horswell S, Larkin J, Endesfelder D, Gronroos E, Martinez P, Matthews N, Stewart A, Tarpey P, Varela I, Phillimore B, Begum S, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. N Engl J Med. 2012; 366:883-892.
- 15. Meacham CE, Morrison SJ. Tumour heterogeneity and cancer cell plasticity. Nature. 2013; 501:328-337.
- Ding L, Ley TJ, Larson DE, Miller CA, Koboldt DC, Welch JS, Ritchey JK, Young MA, Lamprecht T, McLellan MD, McMichael JF, Wallis JW, Lu C, et al. Clonal evolution in relapsed acute myeloid leukaemia revealed by wholegenome sequencing. Nature. 2012; 481:506-510.
- 17. Greaves M, Maley CC. Clonal evolution in cancer. Nature. 2012; 481:306-313.
- Zhang J, Fujimoto J, Zhang J, Wedge DC, Song X, Zhang J, Seth S, Chow CW, Cao Y, Gumbs C, Gold KA, Kalhor N, Little L, et al. Intratumor heterogeneity in localized lung adenocarcinomas delineated by multiregion sequencing. Science. 2014; 346:256-259.
- de Bruin EC, McGranahan N, Mitter R, Salm M, Wedge DC, Yates L, Jamal-Hanjani M, Shafi S, Murugaesu N, Rowan AJ, Gronroos E, Muhammad MA, Horswell S, et al. Spatial and temporal diversity in genomic instability processes defines lung cancer evolution. Science. 2014; 346:251-256.

- 20. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011; 144:646-674.
- Ganeshan B, Goh V, Mandeville HC, Ng QS, Hoskin PJ, Miles KA. Non-small cell lung cancer: histopathologic correlates for texture parameters at CT. Radiology. 2013; 266:326-336.
- 22. Tixier F, Hatt M, Le Rest CC, Le Pogam A, Corcos L, Visvikis D. Reproducibility of tumor uptake heterogeneity characterization through textural feature analysis in 18F-FDG PET. J Nucl Med. 2012; 53:693-700.
- 23. Willaime JM, Turkheimer FE, Kenny LM, Aboagye EO. Quantification of intra-tumour cell proliferation heterogeneity using imaging descriptors of 18F fluorothymidine-positron emission tomography. Phys Med Biol. 2013; 58:187-203.
- 24. Alic L, Niessen WJ, Veenland JF. Quantification of heterogeneity as a biomarker in tumor imaging: a systematic review. PLoS One. 2014; 9:e110300.
- 25. Aerts HJ, Velazquez ER, Leijenaar RT, Parmar C, Grossmann P, Carvalho S, Bussink J, Monshouwer R, Haibe-Kains B, Rietveld D, Hoebers F, Rietbergen MM, Leemans CR, Dekker A, Quackenbush J, Gillies RJ, Lambin P. Decoding tumour phenotype by noninvasive imaging using a quantitative radiomics approach. Nat Commun. 2014; 5:4006.
- 26. Ganeshan B, Abaleke S, Young RC, Chatwin CR, Miles KA. Texture analysis of non-small cell lung cancer on unenhanced computed tomography: initial evidence for a relationship with tumour glucose metabolism and stage. Cancer Imaging. 2010; 10:137-143.
- 27. Sequist LV, Waltman BA, Dias-Santagata D, Digumarthy S, Turke AB, Fidias P, Bergethon K, Shaw AT, Gettinger

S, Cosper AK, Akhavanfard S, Heist RS, Temel J, et al. Genotypic and Histological Evolution of Lung Cancers Acquiring Resistance to EGFR Inhibitors. Science Translational Medicine. 2011; 3:75ra26-75ra26.

- 28. Bai H, Wang Z, Chen K, Zhao J, Lee JJ, Wang S, Zhou Q, Zhuo M, Mao L, An T, Duan J, Yang L, Wu M, Liang Z, Wang Y, Kang X, Wang J. Influence of chemotherapy on EGFR mutation status among patients with non-small-cell lung cancer. J Clin Oncol. 2012; 30:3077-3083.
- 29. Ichinokawa H, Ishii G, Nagai K, Yoshida J, Nishimura M, Hishida T, Suzuki K, Ochiai A. Clinicopathological characteristics of primary lung adenocarcinoma predominantly composed of goblet cells in surgically resected cases. Pathol Int. 2011; 61:423-429.
- 30. Warth A, Muley T, Meister M, Stenzinger A, Thomas M, Schirmacher P, Schnabel PA, Budczies J, Hoffmann H, Weichert W. The novel histologic International Association for the Study of Lung Cancer/American Thoracic Society/ European Respiratory Society classification system of lung adenocarcinoma is a stage-independent predictor of survival. J Clin Oncol. 2012; 30:1438-1446.
- Simon RM, Subramanian J, Li M-C, Menezes S. Using cross-validation to evaluate predictive accuracy of survival risk classifiers based on high-dimensional data. Briefings in bioinformatics. 2011; 12:203-214.
- Taylor R. Interpretation of the correlation coefficient: a basic review. Journal of diagnostic medical sonography. 1990; 6:35-39.
- Kim HK, Choi YS, Kim K, Shim YM, Jeong SY, Lee KS, Kwon OJ, Kim J. Management of ground-glass opacity lesions detected in patients with otherwise operable nonsmall cell lung cancer. J Thorac Oncol. 2009; 4:1242-1246.