Research Paper

Association between single nucleotide polymorphisms in the *TSPYL6* gene and breast cancer susceptibility in the Han Chinese population

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ABSTRACT

We investigated the associations between single nucleotide polymorphisms (SNPs) in the *testis-specific Y-encoded-like protein 6* (*TSPYL6*) gene and breast cancer (BC) susceptibility in the Han Chinese population. A total of 183 BC patients and 195 healthy women were included in the study. Six SNPs in *TSPYL6* were genotyped and the association with BC risk analyzed. Odds ratios (ORs) and 95% confidence intervals (95% CIs) were calculated using unconditional logistic regression analysis. Multivariate logistic regression analysis was used to identify SNPs that correlated with BC susceptibility. Rs11896604 was associated with a decreased risk of BC based on dominant and genotype models. Rs843706 was associated with an increased risk of BC based on a recessive model. Rs11125529 was associated with decreased BC susceptibility based on a genotype model. Finally, rs843711 inversely correlated with clinical stage III/IV BC. Our findings reveal a significant association between SNPs in the *TSPYL6* gene and BC risk in a Han Chinese population.

INTRODUCTION

Breast cancer (BC) is the most common type of cancer and the leading cause of cancer deaths among women worldwide (particularly in less developed regions including East Asian countries, which accounted for 324,000 deaths or 14.3% of the total) [1]. According to GLOBOCAN 2012, 187,213 individuals were diagnosed with BC in China in 2012, and 47,984 of these individuals died of the disease [2]. BC is a multifactorial disease that has been associated with various factors including age, gender, ethnicity, family history, personal history, lifestyle, as well as both hormonal and non-hormonal risk factors [3]. Hereditary BC clusters in families and is typically diagnosed at an earlier age [4]. Studies of twins have indicated that the risk of BC is higher for a monozygotic twin of a co-twin, suggesting that genetic factors play an important role in BC development [5]. Single nucleotide polymorphisms (SNPs) also play an important role in the genetic susceptibility to BC. Many genes have been associated with a moderate or high lifetime risk of BC including *BRCA1*, *BRCA2*, *PALB2*, *ATM*, and *CHEK2*. In addition, common variants at more than 70 loci have been identified through GWAS and large-scale replication studies [6–9].

The testis-specific Y-encoded-like protein 6 (*TSPYL6*) gene, located on human chromosome 2p16.2, is a member of the *TSPY/TSPYL/SET/NAP-1* (TTSN) superfamily that includes *TSPYL1*, *TSPYL2*, *TSPYL3*, *TSPYL4*, and *TSPYL5* [10]. Upregulation of *TSPYL6* has been observed in both benign and malignant cells. The *TSPYL6* protein has been associated with chromatin and

nucleosome assembly [11]. However, the specific functions of *TSPYL6* are not yet clear. Norling et al. [12] sequenced the *TSPYL6* gene in an entire Sweden patient cohort, but no inactivating mutations were identified. Additionally, no studies have investigated correlations between the *TSPYL6* gene and BC susceptibility. In this case-control study, we genotyped six SNPs in *TSPYL6*: rs843645, rs11125529, rs12615793, rs843711, rs11896604, and rs843706 and performed a comprehensive association analysis to identify SNPs associated with BC risk in Han Chinese women.

RESULTS

Participant characteristics

A total of 183 patients with BC and 195 healthy individuals were enrolled in the study. The participant characteristics are shown in Table 1. No significant differences in age, body mass index (BMI), or the menopause age were observed between patients in the case and control groups (p > 0.05). The mean age of the participants was 45.35 years in the control group and 46.40 years in the case group. The mean BMI was 22.53 in the control group and 23.08 in the case group.

Association between *TSPYL6* polymorphisms and BC risk

Detailed SNP data and the associations between various SNPs and BC risk are shown in Table 2. Our data indicated that all 6 SNPs investigated were in Hardy-Weinberg equilibrium in the control subjects (p > 0.05). No associations were observed between the alleles and BC risk in an allele model. We also performed a Bonferroni correction and determined that none of the SNPs showed statistical significant associations with BC risk.

We further assessed the association between each SNP and BC risk in an unconditional logistic regression analysis, which was performed using four models: additive, dominant, recessive, and genotype model (Tables 3 and 4). Rs11896604 was associated with a decreased risk of BC in a dominant model (odds ratio [OR] = 0.623, 95% confidence interval [95% CI] = 0.405-0.958, p = 0.031). Rs843706 was associated with an increased risk of BC under the recessive model (OR = 1.709, 95%CI = 1.055 - 2.770, p = 0.030) (Table 3). Rs11125529 was associated with a decreased risk of BC under the genotype model (OR = 0.612, 95% CI = 0.391–0.959, p = 0.032) (Table 4). Rs11896604 was associated with a decreased risk of BC in a genotype model (OR = 0.574, 95% CI = 0.370 - 0.891, p = 0.013). No statistical associations were detected under the other models. In addition, no positive results were observed after Bonferroni correction.

In order to assess the associations between SNP haplotypes and BC risk, a Wald test was performed

using an unconditional multivariate regression analysis. However, no positive results were observed (Table 5, Figure 1).

Association between *TSPYL6* polymorphisms and BC patient clinicopathological features

We next analyzed the association between TSPYL6 polymorphisms and BC patient clinicopathological features, which included age, age of menarche, BMI, breastfeeding duration, clinical stage, estrogen receptor status, family history of cancer, procreative time, progesterone receptor status, tumor location, tumor size (cm), tumor type, incipient recurrence, presence of lymph node metastasis, age of menopause, and prim parous age. Positive results are shown in (Table 6A, 6B). For rs11125529, we found that more recurrent BC patients had the AA + CA genotype than the CC genotype (OR = 2.321, 95% CI = 1.192-4.521, p = 0.012)(Table 6A). For rs843711, the CT + CC genotype was observed less frequently in patients with clinical stage III/ IV disease (OR = 0.411, 95% CI = 0.194-0.869, p = 0.018) and in patients with recurrent BC (OR = 0.458, 95%CI = 0.222-0.944, p = 0.032) than the TT genotype (Table 6A). Our results suggested that the frequency of recurrent BC patients with the CC genotype of rs11896604 was higher than the frequency of patients with the GG + CG genotype (OR = 2.471, 95% CI = 1.290-4.734, p = 0.006) (Table 6B). Finally, the CA + CC genotype of rs843706 was more frequently observed in patients with clinical stage III/IV disease (OR = 0.411, 95%CI = 0.194 - 0.869, p = 0.018) and in patients with recurrent BC (OR = 0.458, 95% CI = 0.222–0.944, *p* = 0.032) than the AA genotype (Table 6B). No statistical associations were detected between the other loci and the clinical parameters that were investigated.

DISCUSSION

In this study, we investigated the association between SNPs in the *TSPYL6* gene and BC risk in Han Chinese women. We found that four SNPs (rs11896604, rs843706, rs11125529, and rs843711) were associated with the risk of BC in this population. Rs11896604 was associated with a decreased risk of BC in a dominant and genotype model, but the various genotypes were associated with an increased risk of recurrence in BC patients. An association between this locus and other diseases has not been previously reported. Rs843706 was associated with an increased risk of BC in a recessive model, but there was a decreased association between the SNP and the risk of recurrence as well as with clinical stage III/IV BC.

We are the first to demonstrate an association between this locus and BC susceptibility. Rs11125529 was associated with a decreased risk of BC in a genotype model, but an increased risk of recurrence. Although

Characteristic		Cases (N = 183)	Controls ($N = 195$)	<i>P</i> -value
Mean age \pm SD		$46.40 \pm 9.383 \ (N = 183)$	$45.35 \pm 6.899 \ (N = 195)$	0.218ª
Mean BMI ± SD		$23.08 \pm 3.00 \ (N = 183)$	$22.53 \pm 2.55 \ (N = 195)$	0.056ª
Menopause	Premenopausal	115 (62.8%)	119 (61.0%)	0.716 ^b
	Postmenopausal	68 (37.2%)	76 (39.0%)	
Age of Menarche	≤ 12	25 (13.7%)		
	> 12	158 (86.3%)		
Breastfeeding Duration	≤6	12 (6.5%)		
	> 6	158 (93.5%)		
Clinical Stages	I/II	135 (73.8%)		
	III/IV	48 (26.2%)		
Estrogen Receptor	negative	60 (32.8%)		
	positive	123 (67.2%)		
Family Tumor History	no	156 (85.2%)		
	yes	27 (14.8%)		
Incipientw or Recurrence	Incipient	109 (59.9%)		
	Recurrence	73 (40.1%)		
Lymph node metastasis	no	105 (58.3%)		
	yes	75 (41.7%)		
Menopause	no	115 (62.8%)		
	yes	68 (37.2%)		
Primiparous Age	< 30	170 (96.6%)		
	\geq 30	6 (3.4%)		
Procreative Times	< 1	142 (81.1%)		
	≥ 1	33 (18.9%)		
Progestrone Receptor	negative	75 (41.0%)		
	positive	108 (59.0%)		
Tumor Location	left	84 (45.9%)		
	right	97 (53.0%)		
	both	2 (1.1%)		
Tumor Size (cm)	≤ 3	94 (51.4%)		
	> 3	89 (48.6%)		
Tumor Type	carcinoma	165 (90.2%)		
	others	18 (9.8%)		
Whether fertility	no	7 (3.8%)		
	yes	176 (96.2%)		

Table 1: Basic characteristics of the control individuals and patients with breast cancer

SD: Standard deviation. BMI: Body mass index (weight [kg]/height[m]²). ^a *P* value was calculated by Welch's *t* test. ^b *P* value was calculated by Pearson's χ^2 test.

SNPs	Position	Band	Alleles A/B	MAF-control	MAF-case	HWE-p	OR	95% CI	<i>p</i> -x ²
rs843645	54474664	2p16.2	G/T	0.297	0.279	0.4968	0.913	0.666-1.251	0.57
rs11125529	54475866	2p16.2	A/C	0.195	0.150	0.1695	0.731	0.499-1.069	0.105
rs12615793	54475914	2p16.2	A/G	0.201	0.161	0.1166	0.764	0.525-1.109	0.156
rs843711	54479117	2p16.2	C/T	0.487	0.544	0.1531	1.254	0.942-1.669	0.12
rs11896604	54479199	2p16.2	G/C	0.221	0.167	0.0929	0.707	0.491-1.018	0.062
rs843706	54480369	2p16.2	C/A	0.482	0.544	0.06103	1.281	0.962-1.705	0.090

Table 2: Basic information of candidate SNPs in this study

SNPs: Single nucleotide polymorphisms; MAF: Minor allele frequency; HWE: Hardy-Weinberg equilibrium; OR: Odds ratio; CI: Confidence interval. ^AMinor alleles. ^BMajor alleles.

Ding et al. reported neither the genotype nor the allele frequencies at rs11125529 in *ACYP2* differed significantly between coronary heart disease patients and normal controls [13]. The association between the telomere lengthrelated variant rs11125529 in *ACYP2* and gastric cancer risk was previously investigated in a Chinese population, but no significant association was identified [14]. We found that the rs843711 genotypes in the *TSPYL6* gene were inversely correlated with clinical stage III/IV BC. Finally, rs843645 and rs12615793 were not associated with the risk of BC.

The function of *TSPYL6* may be similar to those of other members of the TTSN superfamily. However, the molecular mechanisms underlying *TSPYL6* function have not been elucidated. Mutation of *TSPYL* can cause

sudden infant death with dysgenesis of the testes (SIDDT) in affected males, indicating that *TSPYL* is important for the development of the testis and other tissues such as the brain [15]. Although *TSPYL* is expressed in all tissues [16], the role of *TSPYL* in tumor cells is not clear. The *TSPYL4* gene is located 25 kb from *TSPYL*, however no coding variants were identified in affected individuals with direct sequencing. The *TSPYL1* gene does not contain any introns, but the exact composition has not been determined [17].

TSPYL2 gene and cyclin B can inhibit cell proliferation by arresting cell growth in response to DNA damage [18]. Thus, it has been suggested that *TSPYL2* is a negative regulator of cell cycle progression. The *TSPYL2* gene is silenced in glioma and malignant lung tissue, and in certain lung cancer cell lines [19]. Overexpression of

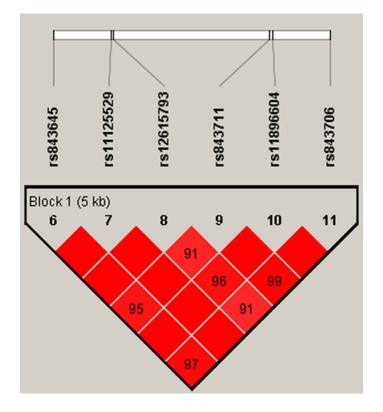


Figure 1: Haplotype block map for all the SNPs of the *TSPYL6* gene.

SNP	Model	Genotype	Cases	Controls	OR (95% CI)	Р
rs843645	Dominant model	T/T	99	94	1	0.246
		G/G-G/T	84	101	0.785 (0.521-1.182)	
	Recessive model	T/T-T/G	165	180	1	0.535
		G/G	18	15	1.259 (0.608-2.606)	
	Additive model	-	-	-	0.904 (0.658-1.241)	0.532
rs11125529	Dominant model	C/C	133	123	1	0.057
		A/A-A/C	50	72	0.652 (0.419-1.013)	
	Recessive model	C/C-C/A	178	191	1	0.618
		A/A	5	4	1.413 (0.364–5.488)	
	Additive model	-	-	-	0.730 (0.491-1.085)	0.120
rs12615793	Dominant model	G/G	129	120	1	0.085
		A/A-A/G	54	74	0.682 (0.441-1.054)	
	Recessive model	G/G-G/A	178	190	1	0.625
		A/A	5	4	1.402 (0.361-5.445)	
	Additive model	-	-	-	0.755 (0.510-1.117)	0.159
rs843711	Dominant model	T/T	39	46	1	0.532
		C/C-C/T	144	149	1.169 (0.405–0.958)	
	Recessive model	T/T-T/C	128	154	1	0.066
		C/C	55	41	1.563 (0.972–2.515)	
	Additive model	-	-	-	1.261 (0.937-1.700)	0.126
rs11896604	Dominant model	C/C	128	114	1	0.031
		G/G-G/C	55	81	0.623 (0.405-0.958)	
	Recessive model	C/C-C/G	177	190	1	0.644
		G/G	6	5	1.336 (0.391–4.564)	
	Additive model	-	-	-	0.709 (0.484–1.039)	0.078
rs843706	Dominant model	A/A	39	45	1	0.603
		C/C-C/A	144	149	1.140 (0.696–1.866)	
	Recessive model	A/A-A/C	128	156	1	0.030
		C/C	55	38	1.709 (1.055–2.770)	
	Additive model	-	-	-	1.294 (0.958-1.750)	0.093

Table 2. Single losi association with breast concerning	(adjusted by aga DMI and mononausa)
Table 3: Single loci association with breast cancer risk	(autusteu Dy age, Divit and menobause)

SNPs: Single nucleotide polymorphisms; OR: Odds ratio. CI: Confidence interval.

P value was calculated by Wald test. $\ast p < 0.05$ indicates statistical significant.

TSPYL2 can inhibit human lung and breast cancer cell lines [20]. However, there is limited evidence for a direct function of *TSPYL2* in cell cycle control. Interestingly, the *TSPYL5* gene has been reported to suppress gastric cancer development [21]. Further studies are required to characterize the function of *TSPYL6* and elucidate the mechanisms underlying the association between the *TSPYL6* and BC susceptibility. Currently, the relationship between clinical characteristics in BC patients and *TSPYL6* gene expression/function is not clear.

Our study is the first to demonstrate that polymorphisms in *TSPYL6* affect the pathogenesis of BC and are associated with clinicopathological characteristics

of BC patients. Collectively, the results provide insight into the pathogenesis of BC. Although this study had sufficient statistical power, there were still some intrinsic limitations. First, the sample size was relatively small (183 cases and 195 controls). Therefore, our findings must be confirmed in studies with larger sample sizes as well as in a meta-analysis. Additionally, we only analyzed Han Chinese women. Therefore, our results must be validated in studies of other populations. Finally, although we identified significant associations between four SNPs (rs11896604, rs843706, rs11125529, and rs843711) and BC susceptibility, the mechanisms responsible for the associations are still unclear. Further studies of *TSPYL6*

Genotype	Cases	Controls	OR (95% CI)	Р
rs843645				
TT	99	94	1.00 [Ref]	
GT	66	86	0.729 (0.475-1.117)	0.147
GG	18	15	1.139 (0.543–2.391)	0.730
rs11125529				
CC	133	123	1.00 [Ref]	
AC	45	68	0.612 (0.391-0.959)	0.032
AA	5	4	1.156 (0.304–4.404)	0.832
rs12615793				
GG	129	120	1.00 [Ref]	
AG	49	70	0.651 (0.419–1.013)	0.057
AA	5	4	1.163 (0.305–4.432)	0.825
rs843711				
TT	39	46	1.00 [Ref]	
СТ	89	108	0.972 (0.583-1.620)	0.913
CC	55	41	1.582 (0.879–2.848)	0.126
rs11896604				
CC	128	114	1.00 [Ref]	
GC	49	76	0.574 (0.37-0.891)	0.013
GG	6	5	1.069 (0.318-3.596)	0.915
rs843706				
AA	39	45	1.00 [Ref]	
CA	89	111	0.925 (0.555-1.543)	0.766
CC	55	38	1.670 (0.921-3.030)	0.092

Table 4: The association between the single-nucleotide polymorphisms and BC risk in Genotype
model (adjusted by age, BMI and menopause)

OR: odd ratio; CI: confidence interval;

p value was calculated by Wald test. *p < 0.05 indicates statistical significance.

Table 5: Haplotype frequency and their association with BC risk in case and control	ol subjects
(adjusted by age, BMI and menopause)	

SNPs	Hanlatuna	Freq%		P1	OR	95% CI		P2
5111 5	Haplotype	case	control	F1	UK	937	0 CI	Γ2
rs843645 rs11125529 rs12615793 rs8437 11 rs11896604 rs843706	TAATGA	0.150	0.192	0.126	0.745	0.501	1.108	0.146
	TCGTGA	0.016	0.023	0.510	0.741	0.256	2.149	0.581
	GCGTCA	0.276	0.292	0.619	0.912	0.662	1.257	0.574
	TCGCCC	0.530	0.474	0.126	1.266	0.939	1.707	0.122

**P*-value < 0.05 indicates statistical significance.

P1- values were calculated from two-sided Chi-squared test.

P2 -values were calculated by unconditional logistic regression.

The reference standard for each haplotype is the other haplotype.

Variables			rs1112	25529		rs843711				
variables	AA + CA	CC	OR ^a	95% CI	P ^b	CT+CC	TT	OR ^a	95% CI	Pb
Age	50	133				144	39			
≤ 40	14	43	1	(reference)		41	16	1	(reference)	
> 40	36	90	1.229	(0.600-2.515)	0.573	103	23	1.748	(0.839–3.639)	0.133
Age of Menarche	50	133				144	39			
≤ 12	5	20	1	(reference)		22	3	1	(reference)	
> 12	45	113	0.628	(0.222 - 1.775)	0.377	122	36	2.164	(0.612-7.646)	0.221
BMI	50	133				144	39			
≤ 24	34	87	1	(reference)		98	23	1	(reference)	
> 24	16	46	0.89	(0.445 - 1.780)	0.742	46	16	0.675	(0.326–1.397)	0.288
Breastfeeding Duration	46	124				133	37			
≤ 6	3	9	1	(reference)		10	2	1	(reference)	
> 6	43	115	0.891	(0.230-3.448)	0.868	123	35	1.423	(0.298-6.797)	0.657
Clinical Stages	50	133				144	39			
I/II	38	97	1	(reference)		112	23	1	(reference)	
III/IV	12	36	0.851	(0.401-1.807)	0.674	32	16	0.411	(0.194–0.869)	0.018
Estrogen Receptor	50	133				144	39			
negative	17	43	1	(reference)		49	11	1	(reference)	
positive	33	90	0.927	(0.466-1.847)	0.83	95	28	0.762	(0.350-1.658)	0.492
Family Tumor History	50	133				144	39			
no	8	114	1	(reference)		121	35	1	(reference)	
yes	42	19	1.143	(0.465-2.807)	0.771	23	4	1.663	(0.539-5.031)	0.372
Procreative Times	47	128				138	37			
< 1	37	105	1	(reference)		112	30	1	(reference)	
≥ 1	10	23	0.81	(0.353-1.862)	0.62	26	7	1.005	(0.398-2.539)	0.991
Progestrone Receptor	50	133				144	39			
negative	22	53	1	(reference)		59	16	1	(reference)	
positive	28	80	0.843	(0.437-1.627)	0.611	85	23	1.002	(0.488-2.058)	0.995
Tumor Location	50	133				144	39			
left	22	62	1	(reference)		66	18	1	(reference)	
right	28	69	1	(reference)		77	20	1	(reference)	
both	0	2			0.631	1	1			0.603
Tumor Size (cm)	50	133				144	39			
≤3	24	70	1	(reference)		76	18	1	(reference)	
> 3	26	63	1.204	(0.628-2.308)	0.576	68	21	0.767	(0.377-1.559)	0.463
Tumor Type	50	133				144	39			
Infiltrating ductal carcinoma	47	118	1	(reference)		128	37	1	(reference)	
others	3	15	1.992	(0.551-7.198)	0.285	16	2	0.432	(0.095-1.967)	0.266
Incipience/Recurrence	49	133				144	38		× ,	
Incipience	22	87	1	(reference)		92	17	1	(reference)	
Recurrence	27	46	2.321	(1.192-4.521)	0.012*	52	21	0.458	(0.222-0.944)	0.032
Lymph node metastasis	49	131		· · · · ·		141	39		× /	
no	29	76	1	(reference)		83	17	1	(reference)	
yes	20	55	0.953	(0.489–1.857)	0.887	58	22	0.904	(0.442–1.851)	0.783
Menopause	50	133		× ····)		144	39	-		
no	27	88	1	(reference)		87	28	1	(reference)	
yes	23	45	1.666	(0.859 - 3.230)	0.129	57	11	1.668	(0.770 - 3.614)	0.192
Primiparous Age	47	129		(139	37		(
< 30	45	125	1	(reference)		136	34	1	(reference)	
≥ 30	2	4	0.72	(0.127-4.006)	0.709	3	3	4	(0.773-20.70)	0.076

Table 6A: The Associations between *TSPYL6* polymorphisms and clinical characteristics of breast cancer patients

Variables			rs1189	06604		rs843706					
variables	GG + CG	CC	OR ^a	95% CI	P^b	CA+CC	AA	OR ^a	95% CI	P^b	
Age	55	128				144	39				
≤ 40	16	41	1	(reference)		41	16	1	(reference)		
> 40	39	87	1.149	(0.576–2.291)	0.694	103	23	1.748	(0.839–3.639)	0.133	
Age of Menarche	55	128				144	39				
≤ 12	6	19	1	(reference)		22	3	1	(reference)		
> 12	49	109	0.702	(0.264–1.868)	0.477	122	36	2.164	(0.612-7.646)	0.221	
BMI	55	128				144	39				
\leq 24	38	83	1	(reference)		98	23	1	(reference)		
> 24	17	45	0.825	(0.419–1.624)	0.578	46	16	0.675	90.326-1.397	0.288	
Breastfeeding Duration	51	119				133	37				
≤ 6	3	9	1	(reference)		10	2	1	(reference)		
> 6	48	110	0.764	(0.198–2.946)	0.695	123	35	1.423	(0.298-6.797)	0.657	
Clinical Stages	55	128				144	39				
I/II	43	92	1	(reference)		112	23	1	(reference)		
III/IV	12	36	0.713	(0.338–1.505)	0.374	32	16	0.411	(0.194–0.869)	0.018*	
Estrogen Receptor	55	128				144	39				
negative	19	41	1	(reference)		49	11	1	(reference)		
positive	36	87	0.893	(0.458 - 1.742)	0.74	95	28	0.762	(0.350-1.658)	0.492	
Family Tumor History	55	128				144	39				
no	45	111	1	(reference)		121	35	1	(reference)		
yes	10	17	1.151	(0.617-3.410)	0.391	23	4	1.663	(0.539–5.131)	0.372	
Procreative Times	52	123				138	37				
< 1	41	101	1	(reference)		112	30	1	(reference)		
≥ 1	11	22	0.812	(0.361-1.824)	0.614	26	7	1.005	(0.398-2.539)	0.991	
Progestrone Receptor	55	128				144	39				
negative	25	50	1	(reference)		59	16	1	(reference)		
positive	30	78	0.769	(0.406-1.457)	0.42	85	23	1.002	(0.488-2.058)	0.995	
Tumor Location	55	128				144	39				
left	25	59	1	(reference)		66	18	1	(reference)		
right	30	67	1	(reference)		77	20	1	(reference)		
both	0	2			0.638	1	1			0.603	
Tumor Size (cm)	55	128				144	39				
≤ 3	27	67	1	(reference)		76	18	1	(reference)		
> 3	28	61	1.139	(0.605 - 2.144)	0.686	68	21	0.767	(0.377–1.559)	0.463	
Tumor Type	55	128				144	39				
Infiltrating ductal carcinoma	52	113	1	(reference)		128	37				
others	3	15	2.301	(0.638-8.295)	0.192	16	2	0.432	(0.095–1.967)	0.266	
Incipience/Recurrence	54	128				144	38				
Incipience	24	85	1	(reference)		92	17	1	(reference)		
Recurrence	30	43	2.471	(1.290-4.734)	0.006*	52	21	0.458	(0.222-0.944)	0.032*	
Lymph node metastasis	54	126				141	39				
no	33	72	1	(reference)		83	22	1	(reference)		
yes	21	54	0.848	(0.442–1.627)	0.621	58	17	0.904	(0.442–1.851)	0.783	
Menopause	55	128				144	39				
no	32	83	1	(reference)		87	28	1	(reference)		
yes	23	45	1.326	(0.694–2.532)	0.393	57	11	1.668	(0.770–3.614)	0.192	
Primiparous Age	52	124				139	37				
< 30	50	120	1	(reference)		136	34	1	(reference)		
\geq 30	2	4	0.833	(0.148-4.697)	0.836	3	3	4	(0.773 - 20.70)	0.076	

Table 6B: The Associations between *TSPYL6* polymorphisms and clinical characteristics of breast cancer patients

BMI: body mass index; CI: confidence interval. OR: odds ratio.

^aAdjusted for Age, Age of Menarche, BMI, Breastfeeding Duration, Clinical Stages, Estrogen Receptor, Family Tumor History, Procreative Times, Progestrone Receptor, Tumor Location, Tumor Size (cm), Tumor Type, Incipient/Recurrence, Lymph node metastasis, Menopause and Primiparous Age.

^bTwo-sided Chi-square test for the distributions of genotype frequencies.

*p < 0.05 indicates statistical significance.

 Table 7: Primers used for this study

SNP_ID	1st-PCRP	2nd-PCRP	UEP_SEQ
rs843645	ACGTTGGATGGAAATCTGA	ACGTTGGATGACAGTGCCTTTA	TCATAGGCACTACT
	ATACCACCTAC	GCAAGGTG	GTATC
rs11125529	ACGTTGGATGGAGCTTAGTT	ACGTTGGATGCCGAAGAAAAG	AGAAAAGAAGATG
	GTTTACAGATG	AAGATGAC	ACTAAAACAT
rs12615793	ACGTTGGATGTTTGAGCTTAG	ACGTTGGATGATCTTGGCCCTT	AAATTGAGTGACAA
	TTGTTTAC	GAAGAA	ATATAAACTAC
rs843711	ACGTTGGATGGACAAAGGACC	ACGTTGGATGTGCCTTGTGGGA	gggaTCAGGGAACCA
	TTACAACTC	ATTAGAGC	GTGCAAA
rs11896604	ACGTTGGATGAAGTCAGAATA	ACGTTGGATGTGTCTCTGACCT	GTTAAGCTTGCAA
	GTGCTTAC	AGCATGTA	GGAG
rs843706	ACGTTGGATGTGAAAGCCAT	ACGTTGGATGTGAATAACTTGG	cACTTGGTCTTATCT
	AAATATTTTG	TCTTATC	GATGC

and other members of the TTSN superfamily are necessary to dissect the mechanisms by which polymorphisms in these genes contribute to BC risk. Hereditary, endocrine, environmental, and life style factors should be also considered.

We performed Bonferroni correction in our statistical analysis, but found no statistical significant associations between *TSPYL6* SNPs and risk of BC. This may be due to the relatively small sample size, the selection criteria for *TSPYL6* SNPs (minor allele frequency [MAF] > 5%), and the weakness of Bonferroni correction itself (the interpretation of a finding depends on the number of other tests performed). True differences may have been deemed non-significant given the likelihood of type II errors.

MATERIALS AND METHODS

Study participants

A total of 183 patients with BC and 195 healthy women were included in this study. The patients were treated at the Second Affiliated Hospital of Xi'an Jiao Tong University between January 2013 and November 2015. All demographic and related clinical data including residential region, age, ethnicity, and education status were collected through a face-to-face questionnaire and a review of medical records. The clinical and demographic characteristics of the patients are shown in Table 1. Patients who had been recently diagnosed with primary BC (confirmed by histopathological analysis) were included in the study. Patients diagnosed with other types of cancers or who underwent radiotherapy or chemotherapy were excluded. Control patients who had undergone annual health evaluations were recruited from health checkup centers affiliated with our institution. All controls were matched with cases based on age (p = 0.218) and ethnicity. All control patients had no history of cancer. Factors that could influence the mutation rate were minimized. The participants were women who were \geq 18 years old with good mental health and no blood relatives with BC going back three generations. This study was performed in accordance with the Chinese Department of Health and Human Services regulations for the protection of human research subjects. Informed consent was obtained from all participants and the study protocols were approved by the Institutional Review Board of Xi'an Jiao Tong University.

SNP selection and genotyping

Validated SNPs that had a MAF > 5% in the HapMap Asian population were selected for the association analysis [12, 20, 22, 23]. Venous blood samples (5 mL) were collected from each patient during a laboratory examination. DNA was extracted from whole blood samples using the Gold Mag-Mini Whole Blood Genomic DNA Purification Kit (version 3.0; TaKaRa, Japan) [24]. The DNA concentration was measured by spectrometry UV/VIS spectrophotometer, Beckman (DU530 Instruments, Fullerton, CA, USA). The Sequenom MassARRAY Assay Design 3.0 software (Sequenom, Inc, San Diego, CA, USA) was used to design the multiplexed SNP Mass EXTEND assay. Genotyping was performed using a Sequenom MassARRAY RS1000 (Sequenom, Inc.) according to the manufacturer's protocol [25]. The SequenomTyper 4.0 Software[™] (Sequenom, Inc.) was used to manage and analyze the data [26]. The primers corresponding to each SNP are shown in Table 7. Based on these results, the following six SNPs were selected: rs843645, rs11125529, rs12615793, rs843711, rs11896604, and rs843706. The SNP data are shown in Table 3.

Statistical analysis

Chi-squared tests (categorical variables) and Student's *t*-tests (continuous variables) were used to evaluate the differences in the demographic characteristics between the cases and controls [27]. The Hardy-Weinberg equilibrium of each SNP was assessed in order to compare the expected frequencies of the genotypes in the control patients. All of the minor alleles were regarded as risk alleles for BC susceptibility. To evaluate associations between the SNPs and risk of BC in the four models (genotype, dominant, recessive, and additive), ORs and 95% CIs were calculated using unconditional logistic regression analysis [28]. In multivariate analyses, unconditional logistic regression was used to assess the association between each SNP and the risk of BC after adjusting for BMI, age, and menopause [28]. Linkage disequilibrium analysis and SNP haplotypes were analyzed using the Haploview software package (version 4.2) and the SHEsi software platform (http://www.nhgg.org/ analysis/) [29]. All statistical analyses were performed using the SPSS version 17.0 statistical package (SPSS, Chicago, IL, USA) and Microsoft Excel. A p < 0.05 was considered statistically significant and all statistical tests were two-sided.

CONCLUSIONS

In summary, we have identified four novel associations between SNPs (rs11896604, rs843706, rs11125529, and rs843711) in *TSPYL6* and BC. Our results suggest that these SNPs may contribute to BC development and possibly other complex genetic traits. These SNPs may function as molecular markers of BC susceptibility, and could therefore be used as diagnostic and prognostic markers in clinical studies of BC patients.

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CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest.

REFERENCES

- Kamangar F, Dores GM, Anderson WF. Patterns of cancer incidence, mortality, and prevalence across five continents: defining priorities to reduce cancer disparities in different geographic regions of the world. J Clin Oncol. 2006; 24:2137–2150.
- Zhou L, He N, Feng T, Geng TT, Jin TB, Chen C. Association of five single nucleotide polymorphisms at 6q25.1 with breast cancer risk in northwestern China. Am J Cancer Res. 2015; 5:2467–75.

- Michailidou K, Beesley J, Lindstrom S, Canisius S, Dennis J, Lush MJ, Maranian MJ, Bolla MK, Wang Q, Shah M, Perkins BJ, Czene K, Eriksson M, et al. Genomewide association analysis of more than 120,000 individuals identifies 15 new susceptibility loci for breast cancer. Nat Genet. 2015; 47:373–380.
- 4. Michailidou K, Beesley J, Lindstrom S, Canisius S, Dennis J, Lush MJ, Maranian MJ, Bolla MK, Wang Q, Shah M, Perkins BJ, Czene K, Eriksson M, et al. Genomewide association analysis of more than 120,000 individuals identifies 15 new susceptibility loci for breast cancer. Nat Genet. 2015; 47:373–380.
- Peto J, Mack TM. High constant incidence in twins and other relatives of women with breast cancer. Nat Genet. 2000; 26:411–414.
- Garcia-Closas M, Couch FJ, Lindstrom S, Michailidou K, Schmidt MK, Brook MN, Orr N, Rhie SK, Riboli E, Feigelson HS, Le Marchand L, Buring JE, Eccles D, et al. Genome-wide association studies identify four ER negativespecific breast cancer risk loci. Nat Genet. 2013; 45:392– 398, 398e391–392.
- Bojesen SE, Pooley KA, Johnatty SE, Beesley J, Michailidou K, Tyrer JP, Edwards SL, Pickett HA, Shen HC, Smart CE, Hillman KM, Mai PL, Lawrenson K, et al. Multiple independent variants at the TERT locus are associated with telomere length and risks of breast and ovarian cancer. Nat Genet. 2013; 45:371–384.
- Milne RL, Burwinkel B, Michailidou K, Arias-Perez JI, Zamora MP, Menendez-Rodriguez P, Hardisson D, Mendiola M, Gonzalez-Neira A, Pita G, Alonso MR, Dennis J, Wang Q, et al. Common non-synonymous SNPs associated with breast cancer susceptibility: findings from the Breast Cancer Association Consortium. Hum Mol Genet. 2014; 23:6096–6111.
- 9. Cai Q, Zhang B, Sung H, Low SK, Kweon SS, Lu W, Shi J. Genome-wide association analysis in East Asians identifies breast cancer susceptibility loci at 1q32.1, 5q14.3 and 15q26.1. 2014; 46:886–890.
- Vinci G, Brauner R, Tar A, Rouba H, Sheth J, Sheth F, Ravel C, McElreavey K, Bashamboo A. Mutations in the TSPYL1 gene associated with 46,XY disorder of sex development and male infertility. Fertil steril. 2009; 92:1347–1350.
- Schayek H, Bentov I, Jacob-Hirsch J, Yeung C, Khanna C, Helman LJ, Plymate SR, Werner H. Global Methylation Analysis Identifies PITX2 as an Upstream Regulator of the Androgen Receptor and IGF-I Receptor Genes in Prostate Cancer. Horm Metab Res. 2012; 44:511–519.
- Norling A, Hirschberg AL, Rodriguez-Wallberg KA, Iwarsson E, Wedell A, Barbaro M. Identification of a duplication within the GDF9 gene and novel candidate genes for primary ovarian insufficiency (POI) by a customized high-resolution array comparative genomic hybridization platform. Hum Reprod (Oxford, England). 2014; 29:1818–1827.

- Ding H, Yan F, Zhou LL, Ji XH, Gu XN, Tang ZW, Chen RH. Association between previously identified loci affecting telomere length and coronary heart disease (CHD) in Han Chinese population. Clin Interv Aging. 2014; 9:857–861.
- Du J, Zhu X, Xie C, Dai N, Gu Y, Zhu M, Wang C, Gao Y, Pan F, Ren C, Ji Y, Dai J, Ma H, et al. Telomere length, genetic variants and gastric cancer risk in a Chinese population. Carcinogenesis. 2015; 36:963–970.
- 15. Puffenberger EG, Hu-Lince D, Parod JM, Craig DW, Dobrin SE, Conway AR, Donarum EA, Strauss KA, Dunckley T, Cardenas JF, Melmed KR, Wright CA, Liang W, et al. Mapping of sudden infant death with dysgenesis of the testes syndrome (SIDDT) by a SNP genome scan and identification of TSPYL loss of function. P Natl Acad Sci USA. 2004; 101:11689–11694.
- Grasberger H, Bell GI. Subcellular recruitment by TSG118 and TSPYL implicates a role for zinc finger protein 106 in a novel developmental pathway. Int J Biochem Cell B. 2005; 37:1421–1437.
- Javaher P, Stuhrmann M, Wilke C, Frenzel E, Manukjan G, Grosshenig A, Dechend F, Schwaab E, Schmidtke J, Schubert S. Should TSPYL1 mutation screening be included in routine diagnostics of male idiopathic infertility? Fertil Steril. 2012; 97:402–406.
- Tu Y, Wu W, Wu T, Cao Z, Wilkins R, Toh BH, Cooper ME, Chai Z. Antiproliferative autoantigen CDA1 transcriptionally up-regulates p21(Waf1/Cip1) by activating p53 and MEK/ERK1/2 MAPK pathways. J Biol Chem. 2007; 282:11722–11731.
- Kim TY, Zhong S, Fields CR, Kim JH, Robertson KD. Epigenomic profiling reveals novel and frequent targets of aberrant DNA methylation-mediated silencing in malignant glioma. Cancer Res. 2006; 66:7490–7501.
- Kandalaft LE, Zudaire E, Portal-Nunez S, Cuttitta F, Jakowlew SB. Differentially expressed nucleolar transforming growth factor-beta1 target (DENTT) exhibits an inhibitory role on tumorigenesis. Carcinogenesis. 2008; 29:1282–1289.

- Kim EJ, Lee SY, Kim TR, Choi SI, Cho EW, Kim KC, Kim IG. TSPYL5 is involved in cell growth and the resistance to radiation in A549 cells via the regulation of p21(WAF1/ Cip1) and PTEN/AKT pathway. Biochem Bioph Res C. 2010; 392:448–453.
- 22. Li Y, Zou L, Li Q, Haibe-Kains B, Tian R, Li Y, Desmedt C, Sotiriou C, Szallasi Z, Iglehart JD, Richardson AL, Wang ZC. Amplification of LAPTM4B and YWHAZ contributes to chemotherapy resistance and recurrence of breast cancer. Nat Med. 2010; 16:214–218.
- 23. Hu G, Chong RA, Yang Q, Wei Y, Blanco MA, Li F, Reiss M, Au JL, Haffty BG, Kang Y. MTDH activation by 8q22 genomic gain promotes chemoresistance and metastasis of poor-prognosis breast cancer. Cancer Cell. 2009; 15:9–20.
- 24. Carracedo An. Forensic DNA typing protocols. (Totowa, N.J.: Humana Press). 2005.
- 25. Gabriel S, Ziaugra L, Tabbaa D. SNP genotyping using the Sequenom MassARRAY iPLEX platform. Current protocols in human genetics / editorial board, Jonathan L Haines [et al]. 2009; Chapter 2:Unit 2 12.
- 26. Thomas RK, Baker AC, Debiasi RM, Winckler W, Laframboise T, Lin WM, Wang M, Feng W, Zander T, MacConaill L, Lee JC, Nicoletti R, Hatton C, et al. Highthroughput oncogene mutation profiling in human cancer. Nat Genet. 2007; 39:347–351.
- Adamec C. [Example of the Use of the Nonparametric Test. Test X2 for Comparison of 2 Independent Examples]. Ceskoslovenske zdravotnictvi. 1964; 12:613–619.
- Bland JM, Altman DG. Statistics notes. The odds ratio. Bmj. 2000; 320:1468.
- Shi YY, He L. SHEsis, a powerful software platform for analyses of linkage disequilibrium, haplotype construction, and genetic association at polymorphism loci. Cell Res. 2005; 15:97–98.