6-Shogaol attenuates LPS-induced inflammation in BV2 microglia cells by activating PPAR-γ

Qinghe Han¹, Qinghai Yuan² and Guanghong Xie¹

¹College of Veterinary Medicine, Jilin University, Changchun 130062, China
²The Second Hospital of Jilin University, Changchun 130000, China

Correspondence to: Guanghong Xie, email: Xiegh@jlu.edu.cn

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ABSTRACT

6-Shogaol, a pungent agent isolated from Zingiber officinale Roscoe, has been known to have anti-tumor and anti-inflammatory effects. However, the anti-inflammatory effects and biological mechanism of 6-Shogaol in LPS-activated BV2 microglia remains largely unknown. In this study, we evaluated the anti-inflammatory effects of 6-Shogaol in LPS-activated BV2 microglia. 6-Shogaol was administrated 1 h before LPS treatment. The production of inflammatory mediators were detected by ELISA. The expression of NF-κB and PPAR-γ were detected by western blot analysis. Our results revealed that 6-Shogaol inhibited LPS-induced TNF-α, IL-1β, IL-6, and PGE2 production in a concentration dependent manner. Furthermore, 6-Shogaol inhibited LPS-induced NF-κB activation by inhibiting phosphorylation and nuclear translocation of NF-κB p65. In addition, 6-Shogaol could increase the expression of PPAR-γ. Moreover, inhibition of PPAR-γ by GW9662 could prevent the inhibition of 6-Shogaol on LPS-induced inflammatory mediator production. In conclusion, 6-Shogaol inhibits LPS-induced inflammation by activating PPAR-γ.

INTRODUCTION

The incidence of neurodegenerative disease, particularly Parkinson disease (PD) and Alzheimer’s disease, increased markedly in the last decades [1, 2]. Microglia, the major immune cells in the brain, plays a key role in host defence response to injury or infectious agents [3]. Microglia is exquisitely sensitive to brain injury and disease [4]. Overactivation of microglia leads to the production of inflammatory mediators which plays a critical role in the development of neuroinflammation [5, 6]. Neuroinflammation has recently been implicated as an important mechanism responsible for the pathological processes of neurodegenerative diseases [7, 8]. Therefore, the identification of agents to inhibit neuroinflammation might be an effective approach for the treatment of neurodegenerative diseases.

6-Shogaol, a pungent agent from Zingiber officinale Roscoe, has been reported to have anti-tumor and anti-inflammatory effects. 6-Shogaol has been reported to protect against LPS-induced acute lung injury in mice. Also, 6-Shogaol was found to attenuate neuroinflammation and cognitive deficits in animal models of dementia [9]. Furthermore, 6-Shogaol has been reported to inhibit LPS-induced iNOS and COX-2 expression in macrophages [10]. In addition, studies showed that 6-Shogaol could protect against LPS-induced toxicity in murine astrocytes [11]. However, whether 6-Shogaol could inhibit LPS-induced anti-inflammatory response in activated microglial cells remains unclear. In the present study, we evaluated the anti-inflammatory effects of 6-Shogaol in LPS-stimulated BV2 microglia.

RESULTS

Effects of 6-Shogaol on cell viability

To test whether 6-Shogaol has cytotoxicity on BV2 microglia, MTT assay were used in this study. The results showed that 6-Shogaol had no cytotoxicity on BV2
microglia at the concentration of 0 to 20 μg/mL (Figure 1). Therefore, 6-Shogaol (5, 10, 20 μg/mL) were used in the following experiments.

6-Shogaol inhibited LPS-induced TNF-α, IL-1β, IL-6, and PGE₂ production

To investigate the anti-inflammatory effects of 6-Shogaol, the expression of inflammatory mediators were detected in this study by ELISA. As shown in Figure 2, LPS dramatically increased the production of TNF-α, IL-1β, IL-6, and PGE₂. However, 6-Shogaol concentration dependently down-regulated the production of TNF-α, IL-1β, IL-6, and PGE₂ induced by LPS.

6-Shogaol inhibited LPS-induced NF-κB activation

NF-κB has been known to be involved in the regulation of inflammatory mediators. To investigate the anti-inflammatory mechanism of 6-Shogaol, LPS-induced NF-κB activation were detected in the present study. The results showed that LPS significantly up-regulated the phosphorylation levels of NF-κB p65 and IκBα. Pretreatment of 6-Shogaol concentration dependently inhibited LPS-induced NF-κB p65 phosphorylation and IκBα phosphorylation and degradation (Figure 3).

Effects of 6-Shogaol on PPAR-γ expression

Previous studies showed that activation PPAR-γ could inhibit LPS-induced NF-κB activation. Thus, we detected whether 6-Shogaol could up-regulated the expression of PPAR-γ. As shown in Figure 4, 6-Shogaol increased the expression of PPAR-γ in a concentration dependent manner (Figure 4).

GW9662 prevented the anti-inflammatory effects of 6-Shogaol

To further evaluate the anti-inflammatory mechanism of 6-Shogaol, PPAR-γ was blocked by its inhibitor GW9662. As shown in Figure 5, our results indicated that the inhibition of 6-Shogaol on TNF-α, IL-1β, IL-6 and PGE₂ production were prevented by GW9662. These results suggested that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia by activating PPAR-γ.

DISCUSSION

Microglia has been known to play an important role in neurodegenerative diseases [12]. Increasing evidences suggested that controlling the activation of microglia may have protective effects against neurodegenerative diseases [13]. In this study, the results showed that 6-Shogaol inhibited LPS-induced microglia activation by activating PPAR-γ.

Microglia, the prime effector cells in the brain, plays a critical role in immune defense and inflammatory responses [14]. However, overactivation of microglia could lead to the pathological process of neurodegenerative diseases [15]. LPS has the ability to induce microglia activation, which lead to the release of inflammatory mediators [12, 16]. These inflammatory mediators, such as TNF-α, IL-1β, IL-6 and PGE₂, play an important role in the pathological process of...
neurodegenerative diseases [17]. In the present study, our results showed that 6-Shogaol significantly inhibited LPS-induced inflammatory mediators production in BV2 microglia. The results indicated that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia.

It has been reported that NF-κB played a critical role in neuroinflammation [18]. LPS could induce NF-κB activation and inflammatory cytokines release [19]. Inhibition of LPS-induced NF-κB activation could attenuate neuroinflammation [20]. To clarify the anti-inflammatory mechanism of 6-Shogaol, NF-κB activation were measured in this study. We demonstrated that 6-Shogaol significantly inhibited LPS-induced NF-κB activation. PPAR-γ, belongs to a nuclear receptor superfamily, is a ligand-activated transcription factor [21]. Activation of PPAR-γ could regulate metabolism

Figure 2: Effects of 6-Shogaol on LPS-induced TNF-α, IL-1β, IL-6 and PGE₂ production. The production of TNF-α, IL-1β, IL-6 and PGE₂ were measured by ELISA. The data presented are the means ± SEM of three independent experiments. *p < 0.05 vs. control group; **p < 0.01 vs. LPS group.

Figure 3: Effects of 6-Shogaol on NF-κB expression. The values presented are the means ± SEM of three independent experiments. *p < 0.05 vs. control group; **p < 0.01 vs. LPS group.
and inflammation [22]. Previous studies PPAR-γ agonists inhibited LPS-induced airway inflammation [23]. Also, PPAR-γ agonists could suppress LPS-induced inflammatory response in RAW264.7 cells [24]. Furthermore, PPAR-γ agonists have been reported to have therapeutic role in diabetes, inflammation, and cancer [25]. In this study, our results showed that 6-Shogaol increased the expression of PPAR-γ. And GW9662, a PPAR-γ agonist, suppressed the expression of PPAR-γ in RAW264.7 cells.

**Figure 4:** Effects of 6-Shogaol on PPAR-γ expression. The values presented are the means ± SEM of three independent experiments. *p < 0.05 vs. control group; *p < 0.05, **p < 0.01 vs. LPS group.

**Figure 5:** Effects of PPAR-γ inhibitor GW9662 on the anti-inflammatory effects of 6-Shogaol. Cells were treated with GW9662 for 12 h. Then, the cells were treated with 6-Shogaol and stimulated by LPS. The productions of inflammatory mediator were detected 24 h after LPS treatment. The values presented are the means ± SEM of three independent experiments. *p < 0.05 vs. control group; *p < 0.05, **p < 0.01 vs. LPS group.
inhibitor, could prevent the inhibition of 6-Shogaol on TNF-α, IL-1β, IL-6 and PGE₂ production. These results indicated that 6-Shogaol exhibited anti-inflammatory effects in BV2 microglia by activating PPAR-γ.

In conclusion, our results demonstrated that 6-Shogaol suppressed LPS-induced inflammatory mediators production by activating PPAR-γ, which subsequently inhibited LPS-induced NF-κB activation. 6-Shogaol might be an effective agent for the treatment of neurodegenerative diseases.

MATERIALS AND METHODS

Materials

6-Shogaol (purity > 98%) was obtained from the National Institute for the Control of Pharmaceutical and Biological Products (Beijing, China). LPS (Escherichia coli O55:B5) and MTT were purchased from Sigma (St. Louis, MO, USA). ELISA kits of PGE₂, TNF-α, IL-6 and IL-1β were purchased from BioLegend (San Diego, CA). PPAR-γ monoclonal antibody was obtained from Santa Cruz Biotechnology (Heidelberg, Germany). NF-κB p65, IκBα, and β-actin monoclonal antibodies were obtained from Cell Signaling Technology Inc (Boston, MA, USA).

Cell culture

Murine BV2 microglia cells were purchased from China Center for Type Culture Collection (CCTCC, Wuhan, China). The cells were cultured in DMEM with 5% fetal bovine serum, 100 U/ml penicillin, and 100 μg/ml streptomycin. The cells were treated with 6-Shogaol 1 h before LPS treatment.

Cell viability

For determination of cell viability, MTT assay was applied in this study. BV2 microglia was incubated with 6-Shogaol alone and with LPS for 18 h. Then, the cells were treated with MTT for 4 h and the formazan formed was dissolved with DMSO (150 μl/well). The optical density was determined at 570 nm using a Bio-Rad spectrophotometer.

ELISA assay

24 h after LPS treatment, the levels of TNF-α, IL-1β, IL-6, and PGE2 in culture media were tested using commercially available ELISA kits (BioLegend, San Diego, CA). The assay was performed following the instructions provided by the manufacturers.

Western blot analysis

The cells were lysed using RIAP lysis buffer and the concentration was measured by BCA method. Equal amount of protein was resolved using 12% SDS-polyacrylamide gel. The proteins were transferred onto PVDF membranes. The membranes were blocked with 5% skimmed milk and incubated with primary antibodies and HRP-conjugated goat anti-rabbit IgG. The proteins were tested using the chemiluminescence detection system (Amersham, Berkshire, UK). Finally, the bands were analyzed using ImageJ software.

Statistical analysis

Data were presented as means ± SEM. Statistical comparison of the data were analyzed by one-way ANOVA with post-test Neuman-Keuls. A p value < 0.05 was considered as significant.

CONFLICTS OF INTEREST

All authors declare that they have no conflicts of interest.

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REFERENCES


